



**Addis Ababa University**  
**College of Health Sciences**  
**School of Medicine**  
**Department of Medical Biochemistry**

**Title: TOTAL OXIDANT-ANTIOXIDANT STATUS AMONG  
PSORIATIC PATIENTS ATTENDING AT ALERT  
HOSPITAL, ADDIS ABABA, ETHIOPIA**

*A thesis submitted to Addis Ababa University School of Graduate Studies,  
Department of Biochemistry in partial fulfillment of the requirements for the  
Degree of Master of Science in Medical Biochemistry.*

**By: Asrat Endrias (BSc)**

**Advisors**

**Daniel Seifu (PhD) (Department of Biochemistry)**

**Menakath Menon (PhD) (Department of Biochemistry)**

**Mihretu Wodeyes (MD +DV) (Department of Dermatology)**

**Tamrat Abebe (PhD)(Department of Microbiology, Parasitology and Immunology)**

**March 2016**

## **Declaration**

I declare that the thesis hereby submitted for the masters of sciences Degree at the University of Addis Ababa Collage of Health Sciences is my own work and has not been previously submitted by me at another university for any Degree. To the best of my knowledge and beliefs, this thesis contain no material previously published or written by another person, except where due reference is made. I give up copyright of the thesis in favor of the University of Addis Ababa.

-----

Asrat Endrias

March 2016

**College of Health Sciences**  
**School of Medicine**  
**Department of Medical Biochemistry**

This is to certify that the thesis prepared by Asrat Endrias entitled: *Total oxidant-antioxidant status among psoriatic patients attending at Alert Hospital* is submitted in partial fulfillment of the requirements for degree of masters in medical biochemistry complies with the regulations of the university and meets the accepted standards with respect to originality and quality.

Signed by the examining committee:

Examiner \_\_\_\_\_ signature \_\_\_\_\_ Date \_\_\_\_\_

Advisor \_\_\_\_\_ signature \_\_\_\_\_ Date \_\_\_\_\_

Advisor \_\_\_\_\_ signature \_\_\_\_\_ Date \_\_\_\_\_

Advisor \_\_\_\_\_ signature \_\_\_\_\_ Date \_\_\_\_\_

Advisor \_\_\_\_\_ signature \_\_\_\_\_ Date \_\_\_\_\_

Chair of Department or graduate coordinator \_\_\_\_\_

## **ACKNOWLEDGEMENTS**

I would like to express my gratitude and appreciation to my supervisor Dr. Daniel Seifu for his support, insightful knowledge, and encouragement throughout my research. I would like also to thank Dr. Menon for the insights and patience particularly in reviewing my thesis, and insightful comments. I wish to thank Dr. Tamrat for his permission and guidance to use ELISA micro plate reader. I would like to thank the Dr. Mihretu for all his help.

Special thanks to all participants, ALERT/HARI staff members, Nurses and physicians for their help in getting things done. Gratitude is also expressed to all of the people whom without their help this project would not have been possible. I would like to thank Desalegn Yibeltal, Mohamed Mehdi, Tewodros Mengesha and Solomon Tsegaye. Finally, I would like to thank my family for all of their love and support.

# TABLE OF CONTENTS

ACKNOWLEDGEMENTS .....	I
LIST OF TABLES .....	IV
LIST OF FIGURES .....	V
ABBREVIATION .....	VI
<i>Abstract</i> .....	VII
1.INTRODUCTION .....	1
1.1.Psoriasis .....	1
1.1.1. Epidemiology of psoriasis .....	2
1.1.2. Clinical and histological features of psoriasis .....	3
1.1.3. Co-morbidities of psoriasis.....	5
1.1.4. Treatment of psoriasis .....	5
1.1.5. Pathogenesis of psoriasis.....	5
1.1.5.1. Genetic factors.....	6
1.1.5.2. Environmental triggers .....	6
1.1.5.3. Immunopathogenesis of Psoriasis.....	7
1.2.Literature Review.....	9
1.2.1.Generation of Reactive Oxygen Species (ROS) in the skin.....	9
1.2.2.Role of Oxidative Stress in the skin .....	11
1.2.3.Antioxidants in the skin.....	13
1.2.4.Oxidative stress in the pathogenesis of psoriasis .....	15
1.2.4.1. MAPK\AP-1 pathway.....	16
1.2.4.2.NF- $\kappa$ B pathway modulation by ROS.....	16
1.2.4.3. Transactivation between MAPK/AP-1 and NF- $\kappa$ B.....	17
1.2.4.4. JAK–STAT pathway modulation by ROS.....	17
1.2.4.5. Protein kinase pathways .....	18
1.2.5. Evidence of oxidative stress in psoriasis patients .....	18
1.3.Statement of the problem .....	21
1.4.Significance of the study .....	22
1.5.Hypothesis.....	22
2.OBJECTIVE .....	23
2.1.General objective .....	23
2.1.1.Specific objectives.....	23
3.MATERIAL AND METHODS .....	24
3.1.Study design and period .....	24
3.2.Study area and study subjects.....	24

3.3. Inclusion criteria .....	24
3.4. Exclusion criteria .....	24
3.5. Sample size determination .....	25
3.6. Study variables .....	25
3.6.1. Dependent variables .....	25
3.6.2. Independent variable .....	25
3.7. Ethical Consideration .....	26
3.8. Data and Specimen collection handling and storage .....	26
3.9. Biochemical assays and Laboratory analysis .....	27
3.9.1. Determination of Total Oxidant Capacity (TOC) .....	27
3.9.2. Determination of Total Antioxidant Capacity (TAC) .....	28
3.9.3. Measurement of oxidative stress index (OSI) .....	29
3.10. Statistical Analysis .....	30
3.11. Dissemination of results .....	30
3.12. Quality control .....	30
4. RESULTS .....	31
4.1. Demographic characterization .....	31
4.2. Estimated biochemical parameters .....	32
4.2.1. Total oxidant- antioxidant between psoriasis and health control .....	32
4.2.2. Total oxidant- antioxidant status and severity of disease .....	33
4.2.3. Psoriasis disease duration and changes in Total oxidant- antioxidant status .....	34
4.2.4. Correlations between Total oxidant –antioxidant in psoriatic patients .....	35
4.2.5. Correlations between Total oxidant –Antioxidant and severity of disease .....	36
5. DISCUSSION .....	37
6. CONCLUSION .....	44
7. RECOMMENDATION .....	45
8. LIMITATION .....	46
9. REFERENCES .....	47
Appendix I: Questionnaire .....	57
Appendix II: Consent form .....	60
Appendix III -Informed consent .....	62
Appendix IV -Amharic Version .....	63
Appendix VII –PASI .....	68

## LIST OF TABLES

	Page
Table 4.1: Demographic characteristics .....	31
Table 4.2: TOC, TAC and OSI levels in patients versus controls .....	32
Table 4.3: TOC, TAC and OSI within different groups of psoriasis patients .....	33
Table 4.4: psoriasis disease duration and changes in levels of TOC, TAC and OSI.....	34
Table 4.5: Correlations between TOC levels and TAC, OSI levels in psoriatic patients.....	35
Table 4.6: Correlations between TOC levels and TAC.....	36

## LIST OF FIGURES

	Page
Figure 1.1: Clinical manifestation of plaque type psoriasis.....	4
Figure 3.1: Standard curve for estimation of Total oxidant status (TOS).....	27
Figure 3.2: Standard curve for estimation of Total antioxidant capacity (TAC).....	29
Figure 4.1: Regression fit of (a) TOC Vs TAC and (b) TOC Vs OSI.....	35



## ABBREVIATION

CAT	Catalase
ERK	Extracellular- regulated kinase
GP	Glutathione peroxidase
IFN- $\alpha$	Interferon-alpha
IL	Interlukin
iNOS	Inducible nitric oxide synthase
JAK-STAT	Janus kinase-Signal transducer and activator of transcription
MAPK	Mitogen-activated protein kinase
MDA	Manoldialdehyde
MEKK	Mitogen extracellular kinase kinase
MSK1	Mitogen- and- stress-activated protein kinase
NF-KB	Nuclear factor kB
NO	Nitric oxide
OSI	Oxidative stress index
PKC	Protein kinase C
PMN	Polymorphoneuclear
ROS	Reactive oxygen species
SOD	Superoxide dismutase
TAC	Total Anti-oxidant Capacity
TNF $\alpha$	Tumor Necrosis Factor alpha
TOC	Total Oxidant Capacity

## ***Abstract***

*Psoriasis is a chronic inflammatory skin disease characterized by the immune system activation in which the genetic and environmental factors are involved that trigger the development of skin lesions. Reactive oxygen species produced as a result of skin inflammation may cause disorders of the antioxidant defense systems and increased oxidative stress in psoriasis which was proposed to have to have a consequent function in psoriasis.*

***Aim of the study:*** *The presented research work was planned to evaluate oxidative stress by measuring Total Oxidant Capacity, Total Anti-oxidant capacity and Oxidative Stress Index in psoriatic patients.*

***Methods:*** *A comparative cross-sectional study was undertaken with 45 clinically diagnosed psoriatic patients without any drug therapy and 45 age-sexes matched healthy controls. The severity of psoriasis was determined by Psoriasis area severity index. Out of total 45 Psoriasis patients, 22 were mild, 15 moderate and remaining 8 were severe psoriatic patients. Levels of Total Oxidant Capacity, Total Anti-oxidant Capacity and Oxidative Stress Index were measured in patients and controls.*

***Results:*** *The present study showed significantly ( $p < 0.001$ ) increased levels of serum Total Oxidant Capacity as well as Oxidative Stress Index ( $p < 0.001$ ) in psoriasis patients as compared to controls and positively correlated with severity and duration of the disease. Total Anti-oxidant Capacity levels were significantly ( $p < 0.001$ ) lower in patients than in controls and negatively correlated with severity and duration of the disease.*

***Conclusion:*** *These results provide some evidence regarding the role of increased reactive oxygen species with decreased antioxidant activity in psoriatic patients.*

***Key Words:*** *Total Oxidant capacity, Total Antioxidant capacity, Oxidative stress index Psoriasis.*

# 1. INTRODUCTION

## 1.1. Psoriasis

The desire to have beautiful and healthy looking skin has been a centuries-old quest for humans. Skin with brighter complexion and smoother surface tends to be perceived as being healthier and more attractive. Beyond its obvious roles of keeping our insides in and the outside out, skin have many less conspicuous duties: its sweat glands help regulate temperature, its nerves provide the sense of touch, its deeper cells produce vitamin D, and its appearance advertises your age and health. For those who can read skin's language, a close examination reveals clues about the whole body's health. Changes in the skin color may indicate homeostatic imbalances in the body (Ng and Lau, 2015).

Skin makes up to 12–15% of an adult's body weight. Each square centimeter has 6 million cells, 5,000 sensory points, 100 sweat glands, and 15 sebaceous glands. It consists of three layers the epidermis, the dermis and the hypodermis. The epidermis is the protective skin layer in contact with the external environment. This skin layer consists mainly of a stratified squamous keratinized epithelium. The epidermis cells, the keratinocytes, divide in the basal layer and differentiate throughout their migration to the surface (Ojeh *et al.*, 2015). The epidermis is divided into five different layers: stratum basale, stratum spinosum, stratum granulosum, stratum lucidum and stratum corneum. The dermis is the feeder layer of the epidermis and provides most of the skin's mechanical resistance and elasticity. It is mainly composed of fibroblasts, epidermal appendages, blood vessels, nerves and nerve endings. The hypodermis is the deepest layer of the skin. It varies in size and content, but is usually composed of adipocytes which form the adipose tissue (Jean and Pouliot, 2010).

Skin disease is one of the most common human illnesses which pervades all cultures, occurs at all ages, and affects between 30% and 70% of individuals. Its detrimental effects on health range from physical incapacity to death. The International Classification of human disease lists more than 1,000 skin or skin-related illnesses, a pattern dominated by a few conditions accounting for most of the skin disease burden. Skin remains the 18<sup>th</sup> leading cause of health burden worldwide (Hay *et al.*, 2014).

Patients of skin disease always experience physical, emotional and socio-economic embarrassment in the society. Psoriasis is one amongst these notorious auto-immune disorder having deep psychological and social impacts (Prashanth *et al.*, 2014). Psoriasis is non- contagious skin disease affecting both sexes equally, and can occur at any age, although it most commonly appears for the first time between the ages of 15 and 25 years (Dilnawaz *et al.*, 2013; Chong *et al.*, 2010). The word ‘psoriasis’ is derived from the Greek word “psora” meaning “itch” + sis "action”. It has been known since ancient times and was originally considered a type of leprosy. It is now considered as immune-mediated chronic inflammatory skin disease that is often associated with systemic manifestations (Al-Shobaili *et al.*, 2013).

In the course of providing their protective function skin cells gets damaged so that they must be replaced. Therefore, our body produces new skin cells deep in the dermal matrix .These cells migrate upward towards the surface as they mature. Some mature skin cells undergo a process called keratinization-the conversion of squamous epithelial cells into keratin or simpler structural proteins. Keratinization eventually leads to cell death leaving a layer of drier, harder organic material (cell bodies) (Nagamani *et al.*, 2015). In a normal healthy adult, the normal skin turnover occurs once in every 30-40 days. Psoriasis accelerates self-renewal due to excessive growth and reproduction of skin cells which leave them to live 4-10 days as a result, migration outside has no time to differentiate and they are not quite functional (Peslyak *et al.*, 2011). Many psoriatic patients tolerate constant pain from cracking and bleeding lesions, and bear the humiliation and discomfort of continually flaking skin. Itching and pain can interfere with basic functions, such as self-care and sleep. Approximately 60% of psoriasis patients missed an average of 26 days of work a year due to their illness (Augustin and Radtke , 2014).

### **1.1.1. Epidemiology of psoriasis**

Worldwide psoriasis is affecting, as presumed, approximately 120–180 million people. The population prevalence of psoriasis has been reported to range from 2% to 3%. Around 150,000 new cases of psoriasis are reported annually. Studies in developed countries have reported higher prevalence rates of on average about 4.6% (Chandran *et al.*, 2010).

A total of 7.2 million US adults had psoriasis in 2010; an estimated 7.4 million US adults were affected in 2013. When stratifying the sample by race among those between ages 20 and 59 years, the psoriasis prevalence was highest in Caucasians at 3.6% followed by African Americans 1.9%, Hispanics 1.6% and others 1.4%. The prevalence of psoriasis among US adults has not changed significantly since 2003 to 2004 (Rachakonda *et al.*, 2014). Population-based surveys from China and Japan have given a similar low prevalence ranging respectively from 0.05 to 1.23% and 0.29 to 1.18%. The same rates were found in India 0.5%-2.3% (Alexis *et al.*, 2014).

Interestingly, clinic and population surveys have shown great variability between different Sub-Saharan African populations. These suggest that West African countries like Nigeria, Ghana, Mali, Senegal, Angola etc, have a lower prevalence of psoriasis 0.05 % to 1% than 3% in the eastern countries like Kenya, Uganda and Tanzania (Diallo *et al.*, 2012).

Information on the prevalence of psoriasis in Ethiopia is extremely limited; however, data on the incidence of psoriasis at a specified clinic or hospital over a defined period of time are more frequently available. While the absolute incidence rate at a clinic may fail to reflect the local prevalence of psoriasis, the relative rates of psoriasis incidence, obtained from clinics were retrieved and reviewed. A total of 2342 dermatologic cases were diagnosed at ALERT pathology laboratory from January 2007 to December 2010 which represent a wide range of inflammatory, infectious, and neoplastic diseases. During this period out of 632 inflammatory dermatoses 43 were diagnosed with psoriasis (Gimbel *et al.*, 2013).

### **1.1.2. Clinical and histological features of psoriasis**

The most frequent clinical type of psoriasis which is affecting 85%–90% of the patients is psoriasis vulgaris (also called plaque psoriasis) (Jiang *et al.*, 2015) characterized by erythematous plaques of the skin with well-defined borders and silvery scales that can be frequently observed on scalp, tips of fingers and toes, palms, soles, umbilicus, gluteus, under the breasts and genitals, elbows, knees, shins and sacrum (Kuchekar *et al.*, 2011) (Figure 1.1A). Less common phenotypes include guttate, inverse, pustular, erythrodermic and palmo-plantar psoriasis.

Histological analysis of psoriasis shows an increased thickening of the epidermis, in conjunction with incomplete keratinocyte differentiation of the upper layers, leading to the retention of nuclei in the stratum corneum (Figure 1.1B). The basal layers of the epidermis are characterized by increased keratinocyte proliferation and elongated epidermal rete ridges. More dermal blood vessels are formed, responsible for the redness of the lesions. Moreover, a massive immune cell infiltrate can be observed in both dermis and epidermis, containing T cells, dendritic cells and others. Very characteristic is the presence of neutrophils in the epidermis and especially in the stratum corneum (Di Meglio *et al.*, 2011; Lowes *et al.*, 2014).

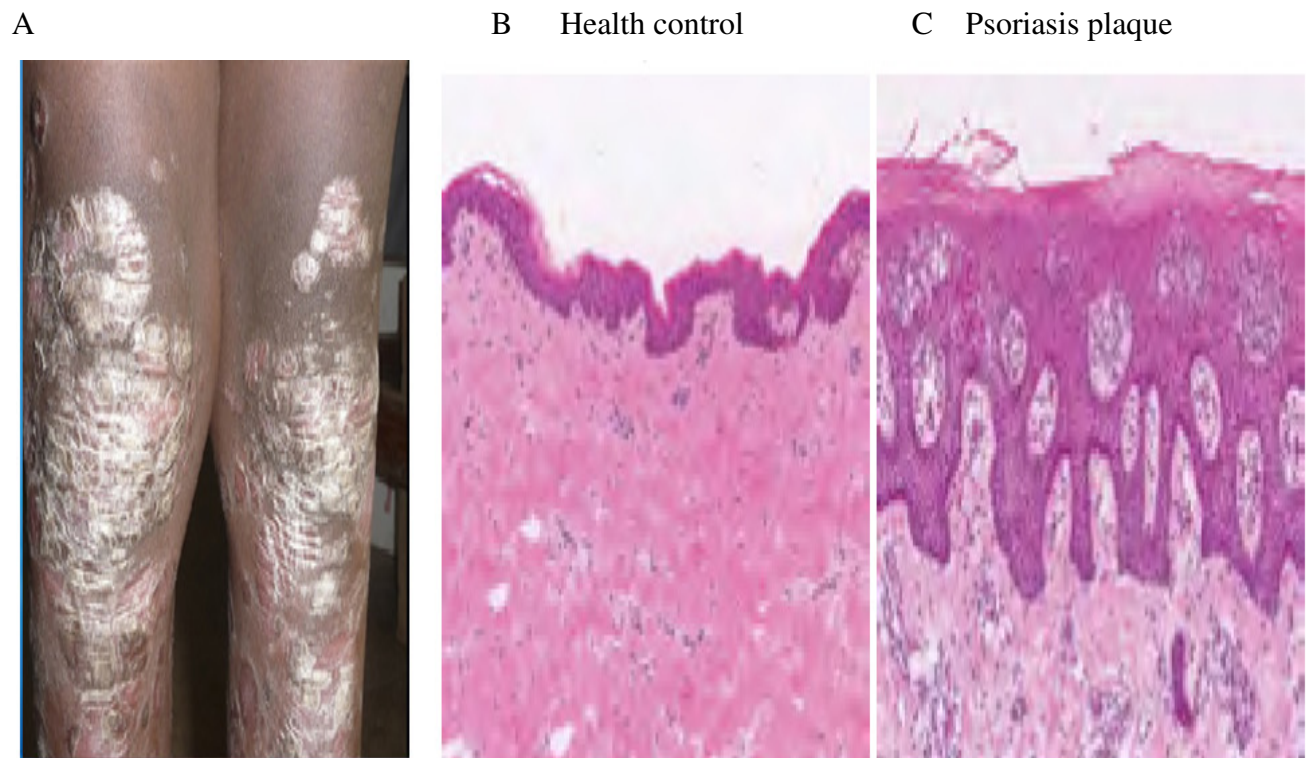


Figure 1.1: Clinical manifestation of plaque type psoriasis. (A) Representative image of psoriatic plaques on the knee of a psoriasis patient. (B) Cross-sections through normal healthy and lesional (C) psoriatic skin (Wohn, 2015).

### **1.1.3. Co-morbidities of psoriasis**

The relationship between psoriasis and other diseases has drawn increasing interest in recent years. Psoriasis is associated with several co-morbidities, including psoriatic arthritis, cardiovascular disease, obesity, diabetes, hypertension, dyslipidemia, metabolic syndrome, nonalcoholic fatty liver disease, cancer, anxiety and depression. Recently, also lymphoma, multiple sclerosis, chronic obstructive pulmonary disease, Chron's disease and inflammatory bowel disease are found at a higher prevalence in psoriatic patients compared to the general population. These disease associations may be due to the systemic inflammatory mediators generated in psoriasis, shared risk factors (i.e., smoking, alcohol consumption), or treatment (Ni and Chiu, 2014).

### **1.1.4. Treatment of psoriasis**

Although treatment options have increased during the past years, no cure is known for psoriasis. Treatment in psoriasis can generally be categorized into topical and systemic therapies. The treatment choice is dictated by the severity, type, and location of psoriasis (Mrowietz *et al.*, 2011). Patients with mild psoriasis can usually be managed with topical agents including: corticosteroids, tar, retinoids and vitamin D derivates. Moderate to severe psoriasis requires phototherapy or systemic therapies such as methotrexate, retinoids, cyclosporine or the biologic immune modifying agents including alefacept, the anti- Tumor Necrosis Factor (TNF) agents or anti-IL12/23 monoclonal antibody (ustekinumab) (Menter *et al.*, 2010).

### **1.1.5. Pathogenesis of psoriasis**

The exact cause and progress of psoriasis remains poorly understood but there is now increasing evidence that psoriasis is a result of a complex interaction between impaired barrier function and innate and adaptive immune systems, which is impacted by environmental and genetic contributions (Balato *et al.*, 2012)

### **1.1.5.1. Genetic factors**

Compared with the general population, a higher incidence of the disease has been identified among first-degree and second-degree relatives of psoriasis. The risk of psoriasis is greater in monozygotic twins than in dizygotic twins, confirming the genetic basis of the disease (Skroza *et al.*, 2013). The mode of inheritance of psoriasis is complex. Several susceptibility loci for psoriasis PSORiasis Susceptibility locus (*PSORS*) have been identified, but the major genetic determinant of psoriasis is *PSORS1*, which is located within the major histocompatibility complex (MHC) on chromosome 6p. Current data suggest that human leukocyte antigen (HLA) Cw6 is the susceptibility allele within *PSORS1*. This association is particularly strong in patients with early onset psoriasis (Chandran *et al.*, 2010). One of the most important features of HLA-C is its capacity to regulate both innate and adaptive responses at the levels of both antigen presentation and natural killer cell regulation. In addition to *PSORS1*, linkage analyses and association studies have highlighted psoriasis loci on several other chromosomes outside of the MHC region, designated *PSORS2-PSORS10* (Sun *et al.*, 2014).

### **1.1.5.2. Environmental triggers**

Psoriasis can be triggered by several environmental stimuli, all of them also known to worsen the ongoing disease. Direct skin trauma can trigger psoriasis (Koebner phenomenon), Streptococcal throat infection, allergies, diet, weather and use of specific medications such as Lithium, anti-malarial medications may also trigger the condition or exacerbate existing psoriasis. Human immunodeficiency virus infection has not been shown to trigger psoriasis, but can exacerbate existing disease. As the infection progresses, psoriasis often worsen (Roberson *et al.*, 2010). Smoking, Obesity and alcohol use and abuse are also associated with psoriasis. These associations may not be causative but patients with psoriasis may be more susceptible to unhealthy behaviors (Weigle *et al.*, 2013).



### 1.1.5.3. Immunopathogenesis of Psoriasis

The innate immune system is activated in the skin in genetically predisposed individuals exposed to environmental triggers like infection or trauma. The initial spark initiating the inflammatory cascade in genetically predisposed individuals has not been known. However, data supports antimicrobial peptide (LL-37) and self-DNA complexes to be a potential explanation of the mechanism through which host DNA is turned into a pro-inflammatory stimulus that breaks immunologic tolerance in psoriasis. The hypothesis is that complexes of the antimicrobial peptide LL-37 cathelicidin and self-DNA released from necrotic cells in the stressed skin activate plasmacytoid dendritic cells and starts of the inflammatory cascade in psoriasis (Al-Shobaili *et al.*, 2013).

It was previously assumed that T-helper 1 (T<sub>H</sub>1) cells played the dominant role in the initiation and maintenance of psoriasis but, in recent years, the view has changed in favor of a T<sub>H</sub>17 mediated disease. Innate immune cells produce key cytokines; tumor necrosis factor alpha (TNF- $\alpha$ ), interferon alpha (IFN- $\alpha$ ), Interferon gamma (IFN- $\gamma$ ), interleukin 1 beta (IL-1 $\beta$ ), and IL-6 that activate dendritic cells (Chu *et al.*, 2011). Activated dendritic cells present antigens and secrete mediators such as IL-12 and IL-23, leading to the differentiation of T<sub>H</sub>1 and T<sub>H</sub>17. IL-23 serves as a key master cytokine regulator. T cells secrete mediators (e.g., IL-17 and IL-22) that activate keratinocytes and induce the production of antimicrobial peptides, proinflammatory cytokines and chemokines. These mediators feed back into the proinflammatory disease cycle and shape the inflammatory infiltrate (DiMeglio *et al.*, 2011).

There is compelling evidence that oxidative stress drives the production of oxidation products, such as 4-hydroxy- 2-nonenal or malonaldehyde which can denature proteins, alter apoptosis, and influence the release of proinflammatory mediators, such as cytokines, which may be critical for the induction of some inflammatory skin diseases. This is also based on the recognition that ROS ( reactive oxygen species) can act as second messengers in the induction of several biological responses, such as the activation of Nuclear factor kappa (NF-kB) or Activator protein 1 (AP-1), the generation of cytokines, the modulation of Pro-inflammatory signaling pathways (Yadav *et al.*, 2013).

Besides direct damaging effects of unregulated ROS production, dysregulation of several pro-inflammatory pathways, like Mitogen-activated protein kinase (MAPK), NF- $\kappa$ B, and Janus kinase-Signal transducer and activator of transcription (JAK-STAT) has been considered to contribute to psoriasis etiology. Several members of the MAPK signaling pathways, like Extracellular-signal-regulated kinase (ERK) 1/2, c-Jun N-terminal kinases (JNK) and MAPK were activated in psoriatic skin, further supporting this notion. The recent demonstration that the peroxisome proliferator-activated receptors, whose natural ligands are polyunsaturated fatty acids and their oxidation products and an increased ROS generation by infiltrated leukocytes into psoriatic has further strengthened the concept that ROS may play a significant role in the pathogenesis of psoriasis (Wagener *et al.*, 2013).

Several studies have investigated the role of oxidants/antioxidants systems in psoriasis with discordant results. Hence, it was decided to investigate the total oxidant- anti oxidant status and oxidative stress index in serum of psoriatic patients.

## 1.2. Literature Review

### 1.2.1. Generation of Reactive Oxygen Species (ROS) in the skin

Free radicals can be defined as reactive chemical species having a single unpaired electron in an outer orbit and are continuously produced by the organism's normal use of oxygen. This unstable configuration creates energy that is released upon reaction with adjacent molecules, such as proteins, lipids, carbohydrates, and nucleic acids. The majority of free radicals that damage biological systems are derived from oxygen and more generally referred to as "reactive oxygen species (Bhattacharya *et al.*, 2015).

ROS can be defined as oxygen-containing molecules, which are more reactive and potent than molecular oxygen itself. ROS can be grouped into oxygen-centered radicals: hydroxyl radical ( $\cdot\text{OH}$ ), peroxy radical ( $\text{ROO}\cdot$ ), alkoxy radical ( $\text{RO}\cdot$ ), Superoxide anion ( $\text{O}_2^{\cdot-}$ ) and oxygen-centered non-radicals: singlet oxygen ( $^1\text{O}_2$ ) and hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) (Oyinloye *et al.*, 2015). Reactive nitrogen species (nitrous oxide, peroxyxynitrite, nitroxyl ion, etc.) are also a class of free radicals derived from nitrogen and considered a subclass of ROS. Reactive nitrogen species (RNS) are generated as a result of sequential reactions that begin with nitric oxide synthase (NOS) mediated conversion of arginine to citrulline. In this reaction, nitric oxide (NO) is generated, which reacts with  $\text{O}_2^{\cdot-}$  to produce peroxyxynitrite ( $\text{ONOO}^-$ ) (Katiyar *et al.*, 2015).

Reactive oxygen species performs beneficial functions in our body and should be maintained. It is the mediator of phagocytosis, apoptosis, detoxification reactions, executioner of precancerous cells and infections, etc. It is beneficially involved in signaling pathways to maintain cellular homeostasis in body. The ROS regulates many metabolic and cellular processes including proliferation, migration, gene expression, and immunity (Klaunig *et al.*, 2010). In the healthy skin, practically all types of skin cells produce ROS and RNS. These free radicals are indispensable effectors in the homeostatic pathways leading to cell proliferation, differentiation, senescence, and death (Pai *et al.*, 2014).

The sources of ROS which are enzymatic as well as non-enzymatic, in the cell are manifold. Enzymes that are ROS producing, on purpose or as a byproduct, include the mitochondrial electron transport chain, nicotinamide adenine dinucleotide phosphate-oxidase (NADPH oxidases) xanthine oxidoreductase (XOR), several peroxisomal oxidases, enzymes of the cytochrome P450 family, cyclooxygenases, and lipoxygenases (Rinnerthaler *et al.*, 2015). Cellular ROS generation also include phagosomes located in specialized cells of the immune system, involved in killing of pathogen; and the endoplasmic reticulum, cell membranes and cytosol (Dickinson *et al.*, 2011).

In human skin, superoxide anion radical  $O_2^{\cdot -}$  is formed by one-electron reduction of molecular oxygen catalyzed by various mitochondrial and cytoplasmic enzymes (Kurahashi, *et al.*, 2015). Mitochondria are the major site for ROS production and also a target of free radicals in the cells. The mitochondrial respiratory chain produces superoxide and  $H_2O_2$  at complex I and III, which can be enhanced by inhibition of the respiratory chain owing to lack of oxygen (Pankotai *et al.*, 2010).

More recent evidence reveals that complex II in the mitochondrial membrane also contributes to the overall  $O_2^{\cdot -}$  Production in human skin cells (Rastogi and Pospisil , 2011). The contribution of mitochondria to the production of ROS in the skin is only substantial in the stem cells, but further on is small compared to other organs, because during the cornification process the keratinocytes degrade all their organelles including the nucleus, mitochondria, peroxisomes, and the endoplasmic reticulum (Anderson *et al.*, 2014).

Apart from the mitochondria,  $O_2^{\cdot -}$  is formed in the cytoplasm of skin keratinocyte cells by a reduction of molecular oxygen catalyzed by heme-flavoprotein NADPH oxidase (NOX) (Bazela *et al.*, 2014). The cytosol has the capacity to produce ROS as a byproduct of the arachidonic acid metabolism. The enzymes cyclooxygenase (COX) and lipoxygenase (LOX) both use arachidonic acid as a substrate to synthesize prostaglandin  $H_2$  and the leukotrienes, respectively. Both enzymes have the capacity to produce superoxide in the presence of NADH or NADPH. The levels of arachidonic acid are relatively low in the skin, but increase in inflammatory skin diseases such as psoriasis, atopic dermatitis, and aging (Rahman *et al.*, 2012).

The skin is at the interface between the body and the environment and is therefore in constant contact with pollutants, xenobiotics, and UV radiation. These exogenous factors represent the main contributor to the formation of ROS in human skin, therefore being very specific for this organ. Additionally, alcohol intake, false nutrition, and physiological and mechanical stress are believed to contribute to this kind of exogenous mediated ROS production. In addition the skin is also one of the very few organs that are in direct contact with atmospheric oxygen (Poljsak *et al.*, 2012).

Inflammation begins when tissues react to a local irritation usually caused by a physical injury, infection or by exposure to toxicant. Fluid and accompanying white blood cells traverse the vascular barrier leading to swelling, erythema, further inflammation and attraction of further white blood cells. During the inflammatory phase, there is a burst of respiration leading to the creation and release of free radicals, including production of both ROS and RNS (Roberts *et al.*, 2010).

Another important pathway that leads to ROS production in skin is formation of prostaglandin E<sub>2</sub> (PGH<sub>2</sub>) from arachidonic acid. The reaction is catalyzed by cyclooxygenase (COX), a prostaglandin-endoperoxide synthetase. The enzyme catalyzes the synthesis of hydroxyl endoperoxides, followed by ROS production (Reuter *et al.*, 2010).

### **1.2.2. Role of Oxidative Stress in the skin**

The skin is one of the major targets of ROS attack since it is exposed to Ultraviolet (UV) radiation and a variety of environmental pollutants, high pressure of molecular oxygen and, in addition, is rich in polyunsaturated fatty acids. After unbalanced ROS exposure, resulting in lipid and protein modifications, the redox status of the intercellular milieu is shifted toward oxidizing conditions (Amaro-Ortiz *et al.*, 2014).

Excess levels of ROS due to overproduction or because of insufficient scavenging generate oxidative stress, leading to injurious effects via: (1) oxidative modification and damage of biomolecules, altering lipid/protein/DNA structure and function; (2) further irreversible oxidation of reactive protein thiol groups which is hallmark of oxidative stress (Mailloux *et al.*, 2012) and (3) dysregulation of cell signaling pathways (Lenaz *et al.*, 2012), triggering

downstream signaling cascades leading to altered cytokine release and exacerbation of inflammation. Combined, excess ROS lead to pathological changes in cells and tissues, as exemplified by inflammatory skin conditions like psoriasis (Garcia-Bailo *et al.*, 2011).

Reactive oxygen species participate in the pathogenesis of many dermatologic diseases both as initiators being primarily involved in their pathology or being the secondary initiating agents generated during the respiratory burst of activated polymorphonuclear leukocytes (Kruk *et al.*, 2014).

Ultraviolet radiation of sunlight has been demonstrated to generate ROS leading to oxidative stress in skin due to depletion of endogenous antioxidant enzymes. ROS generated by UV radiation primarily cause damage to DNA through oxidative modifications and mutations, but also by inducing expression of different genes, such as matrix metalloproteinases and collagenases, thereby affecting collagen integrity and skin aging (Amaro-Ortiz *et al.*, 2014). Additionally, UV-mediated ROS generation also indirectly affects cellular function and survival via its effect on cell signaling pathways. For instance, activation of MAPK proteins occurs after UV exposure, suggesting that it may be responsible for executing the effects of UV-induced oxidative stress (Akasaka *et al.*, 2010).

Reactive oxygen species may participate in the allergic reactions in the skin. It has been reported that nickel triggered allergic reactions in patients are characterized by an elevated level of free iron ions and decreased Glutathione/ glutathione disulfide (GSH/GSSG) ratio in the skin tissue, indicating the presence of oxidative stress (Sicherer *et al.*, 2013).

Reactive oxygen has a paradoxical action on melanocytes because it not only enhances depigmentation, but also increases pigmentation in the skin. An example of melanocyte degeneration induced by oxidative stress is vitiligo, characterized by circumscribed depigmented macules in the skin. The skin of patients with vitiligo vulgaris contains high levels of Superoxide dismutase (SOD) and low levels of catalase (Venza *et al.*, 2015). An imbalance of the ROS scavenging system results in the accumulation of H<sub>2</sub>O<sub>2</sub> in the skin. H<sub>2</sub>O<sub>2</sub> readily crosses the cell membrane and is therefore easily transferred to melanocytes from the keratinocytes. The transfer of H<sub>2</sub>O<sub>2</sub> is thought to be one of the pathogenetic mechanisms of vitiligo.

ROS can also accelerate skin pigmentation. Keratinocytes adjacent to melanocytes intensively contribute to UV-induced skin pigmentation. Among ROS, NO<sup>-</sup> derived from keratinocytes acts to induce melanogenesis by increasing the amount of the melanogenic factors tyrosinase and tyrosinase-related protein (Masaki *et al.*, 2010).

Another important role of ROS is their participation in the skin aging. The free radical theory of aging is supported by finding that oxidative damage to biomolecules accumulates and increases with age. In the skin aging the process oxidative damage involves not only proteins, lipids and DNA but also is linked with alteration of the collagenous extracellular matrix in the dermis (Rinnerthaler *et al.*, 2015).

The involvement of persistent oxidative stress in melanoma and non-melanoma skin cancers (known as the most prevalent cancers in humans) is due to the fact that ROS can activate oncogene by induction of proto-oncogenes, such as NF- $\kappa$ B, c-jun, c-fos and inactivate certain protease inhibitors. Activation of some transcription factors, like NF- $\gamma$ B, AP-1, p53, Hypoxia-inducible factor 1-alpha (HIF-1 $\alpha$ ) can lead to the expression of over 500 genes. The latter activity leads to increased proteases activity and rates of cells migration, thus to tumor progression and metastasis (Reuter *et al.* 2010).

### **1.2.3. Antioxidants in the skin**

It has been known for some time now that the skin possesses an anti-oxidative system for oxidative stress reduction and maintenance of cellular redox balance. In order to cope with constant and excess efflux of damaging reactive metabolites, the skin has developed several lines of defense. In defense against oxidative stress, the skin engages all of these four mechanisms: reparation, prevention, stabilization and, as an important defense mechanism, antioxidant defense where enzymes and scavengers react directly with ROS, preventing them from reaching their biological target (Stojiljković *et al.*, 2014). Their neutralizing capabilities reside in their ability to donate an electron toward off the deleterious effects of the highly reactive radicals or by converting ROS into different, less harmful, molecules (H Kashou *et al.*, 2011).

Antioxidants can be categorized into primary antioxidants and secondary antioxidants. Primary antioxidants are water soluble and lipid soluble. SOD, Catalase (CAT) and Glutathione peroxidase are the primary antioxidant enzymes which inactivate the ROS into intermediates. Ascorbate, glutathione, uric acid etc. are water soluble, and lipids soluble are tocopherols, ubiquinol and carotenoids, etc (Poljsak *et al.*, 2012).

Superoxide dismutase and Catalase are the most important antioxidant systems that protect the epidermis. SOD converts superoxide anions into hydrogen peroxide, whereas CAT catalyzes the decomposition of hydrogen peroxide to water and oxygen (Stojiljković *et al.*, 2014). In addition to catalase, glutathione peroxidase (GPx) also breaks down H<sub>2</sub>O<sub>2</sub> in the presence of the reduced form of glutathione (GSH). GPx also decomposes lipid hydroperoxides into their corresponding alcohols. Thioredoxin, a ubiquitous oxidoreductase enzyme, breaks down H<sub>2</sub>O<sub>2</sub> in a NADPH-dependent reaction within cells. Metallothionein, a heavy metal ion-induced cysteine-rich peptide, also functions as a ROS scavenger (Masaki *et al.*, 2010).

Secondary antioxidant enzymes are Glutathione reductase, Glucose-6-Phosphate dehydrogenase, glutathione-S-transferase and ubiquinone work directly to detoxify ROS by decreasing the peroxides level and continuously supplying the NADPH and glutathione for primary antioxidant enzymes to maintain their proper functioning. Copper, iron, manganese, zinc, selenium enhances the antioxidant enzyme activities (Klaunig *et al.*, 2010).

These Secondary antioxidant enzymes are mainly derived from food and other dietary sources. Several herbs, spices, vitamins, foods, vegetables etc exhibits antioxidant activities. Flavonoids, isoflavones, flavones, anthocyanins, coumarins, lignans, catechins, isocatechins, epicatechin, etc found in natural foods are called phytochemicals. These groups of naturally occurring compounds can scavenge free radicals and thus they are effective as inhibitors of lipid peroxidation (Pandel *et al.*, 2013).



#### **1.2.4. Oxidative stress in the pathogenesis of psoriasis**

Oxidative stress is believed to be a key factor in the pathogenesis of psoriasis. Studies have suggested the involvement of increased ROS levels in psoriasis pathogenesis. Increased ROS generation by infiltrated leukocytes into psoriatic lesions is accompanied by substantial biomolecular damage, like psoriatic skin lesions containing oxidized LDL. Indeed, a relationship between psoriasis severity, lipoprotein levels and oxidative damage has been proposed (Wagener *et al.*, 2013).

Oxidative stress and inflammation are inextricably tied processes. Chronic inflammation is associated with elevated reactive oxygen species levels; anti-inflammatory cascades are linked to diminished reactive oxygen species concentrations. And the converse is true elevated oxidative stress triggers inflammation, whereas redox balance inhibits the cellular response. Thus, oxidative stress and inflammation may be seen as both causes and consequences of cellular pathology (Terlecky *et al.*, 2012).

Reactive oxygen species are generated through different cellular pathways including calcium dependant pathway, protein tyrosine kinase, protein tyrosine phosphatase, serine threonine kinase, phospholipase, mitogen activated protein kinase, NF- $\kappa$ B, cytokines receptors, growth receptor, G-Protein coupled receptor, ion channel receptor and epidermal growth factor (Shafaq *et al.*, 2012).

It is known that ROS, acting as second messengers, influence the cellular signal transduction pathways such as proinflammatory signaling pathways and modulate the expression of numerous genes. Dysregulation of several pro-inflammatory pathways, like MAPK/AP-1, NF- $\kappa$ B, and JAK-STAT signaling pathways have been regarded as early events in inflammatory disorders such as psoriasis (Armstrong *et al.*, 2011). The activation of these signal transduction cascades triggers cell responses to growth factors, cytokines, neurotransmitters, and other intercellular signal molecules, leading to cell proliferation, differentiation, and apoptosis (Zhou *et al.*, 2009).

#### **1.2.4.1. MAPK/AP-1 pathway**

Studies have demonstrated that ROS can induce or mediate the activation of the MAPK pathways. A number of cellular stimuli that induce ROS production also in parallel can activate MAPK pathways in multiple cell types. The prevention of ROS accumulation by antioxidants blocks MAPK activation after cell stimulation with cellular stimuli indicating the involvement of ROS in activation of MAPK pathways (Son *et al.*, 2011).

ERK, p38, and c-Jun N-terminal kinase (JNK) pathways belong to a group collectively termed “MAPK signaling pathways”. ROS are capable of inducing the ERK pathway in a variety of cell types, including epithelial, endothelial, cardio-myocytes, hepatocytes, and T lymphocytes. Compared to non lesional psoriatic skin, lesional psoriatic skin has increased ERK1/2 expression and treatment of psoriasis results in reduced ERK1/2 expression (Jiang *et al.*, 2011).

TNF can induce cellular proliferation through TNF-receptor-associated factor 2 (TRAF2), Mitogen extracellular kinase kinase (MEKK1), and JNK, which results in c-Jun activation. Therefore, it has been hypothesized that, by activating c-Jun, JNK may mediate TNF overproduction and subsequent keratinocyte hyperproliferation in psoriasis. p38MAP Kinase (MAPK) participates in a signaling cascade that controls cellular responses to cytokines and stress. Compared to controls, lesional psoriatic skin had elevated levels of phosphorylated p38 MAPK (Armstrong *et al.*, 2011).

#### **1.2.4.2. NF- $\kappa$ B pathway modulation by ROS**

IL-1, TNF- $\alpha$  and NK- $\kappa$ B are among the first receptors to generate ROS in non-phagocytic cells. TNF- $\alpha$  and oxidants may synergistically activate NK- $\kappa$ B by ROS independent mechanism (Shafaq *et al.*, 2012). NF- $\kappa$ B, another redox-sensitive transcription involved in cellular processes like inflammation, cell proliferation and survival, has recently been demonstrated to be upregulated and active in psoriatic skin. Dysregulation of NF- $\kappa$ B-mediated signaling may further exaggerate disease severity, as NF- $\kappa$ B-mediated upregulation of pro-inflammatory cytokines activates NF- $\kappa$ B via a positive feedback loop.

Importantly, inhibition of NF- $\kappa$ B nuclear translocation and DNA binding activity dampens the inflammatory component of psoriasis (Wagener *et al.*, 2013). Many of the cytokines involved in psoriasis pathology induce keratinocyte proliferation and these signals are via ROS-mediated action transmitted to transcription factors and associated proliferation pathways (Bito *et al.*, 2012).

#### **1.2.4.3. Transactivation between MAPK/AP-1 and NF- $\kappa$ B**

Although ROS involved in the pathogenesis of psoriasis activates MAPK/AP-1 and NF- $\kappa$ B signaling pathways, there is compelling evidence showing the cross talk between these signal transduction pathways. As a downstream effector kinase of ERK1/2 and p38 MAPKs, Mitogen- and- stress-activated protein kinase 1 (MSK1) phosphorylates NF- $\kappa$ B, leading to the expression of multiple NF- $\kappa$ B-dependent genes that are involved in psoriasis. It may be assumed that activation of the transcription factors MAPK/AP- 1 and NF- $\kappa$ B by ROS as well as subsequent cross talk between these signaling transduction cascades triggers and replenishes the pathogenesis of psoriasis (Zhou *et al.*, 2009).

#### **1.2.4.4. JAK–STAT pathway modulation by ROS**

The JAK-STAT pathways are central to a number of inflammatory and immune processes. For example, IL-22 in keratinocytes activates the STAT3 pathway, an important factor in psoriasis inflammation, resulting in increased  $\beta$ -defensin 2/3 expression. Furthermore IFN- $\gamma$  and IL-20 upregulate STAT1 which leads to a cascade of inflammatory mediators that contributes to the formation of psoriasis plaques (Armstrong *et al.*, 2011).

Among the downstream genes of the JAK–STAT cascades, Inducible nitric oxide synthase (iNOS) is the most important one involved in the generation of oxidative stress contributing to the pathogenesis of psoriasis. It has been reported that iNOS is over-expressed in psoriatic lesions compared to uninvolved skin iNOS expression is mediated by the activation of transcriptional factors such as p38 MAPK , NF- $\kappa$ B , and JAK–STAT (Hsu and Armstrong, 2014).

#### **1.2.4.5. Protein kinase pathways**

Protein kinase C (PKC), whose activity is sensitive to the intracellular redox status, is a family of serine–threonine kinases. PKC $\zeta$  is an atypical isoform and regulates multiple cellular functions. PKC $\zeta$  mRNA and protein are increased in psoriatic lesions with membrane translocation of the phosphorylated form of PKC $\zeta$  compared with uninvolved skin. Phospho-PKC $\zeta$  can activate MAPK pathways and NF- $\kappa$ B by phosphorylating Reactive arthritis (RelA) p65 protein, crucial for its DNA binding after translocation into the nucleus to switch on the transcription of a variety of important proinflammatory molecules (Rundhaug *et al.*, 2010). There from, redox-sensitive PKC can trigger the cytokines, chemokines, and adhesion molecules produced in psoriasis. Indeed, ROS stimulate cell proliferation and trigger undifferentiated keratinocytes to form a defective stratum corneum, an event central to the development of psoriasis. Abnormal keratinocyte growth in psoriasis, therefore, may be partially due to the elevated O<sub>2</sub><sup>•-</sup> generated by Polymorphonuclear (PMN) and psoriatic dermal fibroblasts (Zhou *et al.*, 2009).

#### **1.2.5. Evidence of oxidative stress in psoriasis patients**

Malondialdehyde (MDA) is a stable end product of the peroxidation of membrane lipids by ROS, and, thus, it is used as an indicator of increased lipid peroxidation. Interactions between MDA and membrane components result in disturbed structure and function of cell membranes (Avci *et al.*, 2014).

As a potent regulator of keratinocyte growth and differentiation, the multifunctional signaling molecule NO has been considered to be a strong candidate in the pathogenesis of psoriasis. This heat-labile and unstable compound is synthesized in endothelial cells as well as neurons by constitutive NO synthase (cNOS), while iNOS is found in leucocytes, macrophages, and mesengial cells (Singh *et al.*, 2011).

A small amount of NO produced by cNOS in endothelium is responsible for the relaxation of adjacent smooth muscles and prevents adhesion of platelets and leucocytes to the endothelium. This is the anti-inflammatory effect of NO. However, when produced in large amounts NO can destroy tissues and impair immune response (El-Rahman, *et al.*, 2014).

Notably, decreased antioxidant levels have been found together with increased levels of lipid peroxidation markers in blood of psoriasis patients. Also, serum levels of catalase were elevated in psoriatic patients and increased activity of superoxide dismutase (SOD) and expression levels of peroxiredoxin (Prdx) and glutathione peroxidase (GPx) have been found in psoriatic skin lesions. It is tempting to speculate that increased compensatory antioxidant levels counteract the skewed redox balance (Wagener *et al.*, 2013).

In a study conducted with Egyptian population, erythrocyte concentrations of MDA and activity of CAT and SOD in blood samples of 34 patients with psoriasis and 30 healthy subjects. Statistically significant higher levels of plasma MDA and CAT were detected in patients when compared with control subjects. No significant correlations with severity of psoriasis were found. Levels of MDA were positively correlated with levels of CAT in psoriatic patients. Erythrocyte SOD levels were significantly lower in patients than in controls and negatively correlated with severity of the disease but insignificantly correlated with levels of MDA in psoriatic patients (Abdel-Mawla *et al.*, 2013).

An Indian case control study with a total of 50 patients with confirmed diagnosis of psoriasis before starting treatment and 25 healthy controls showed that higher levels of MDA have been observed in Psoriasis patients when compared to controls (Nagamani *et al.*, 2015). In another Indian study, Priya and colleagues also showed a significant role of ROS in pathogenesis of psoriasis where in 40 psoriasis patients between age group of 25 – 60 the levels of serum MDA and NO were significantly increased and SOD and Total Antioxidant Capacity (TAC), were decreased as compared to 40 normal controls subjects (Priya *et al.*, 2013).

In a case-control study from Romania with 34 psoriasis patients in which all had active disease and none of them received any treatment the susceptibility of erythrocyte to lipid peroxidation (expressed as malondialdehyde concentration) was found higher compared to 32 healthy volunteers (Boda *et al.*, 2013).

In a separate Turkish study, the total oxidant and oxidative stress index levels of 40 psoriasis patients was found higher than the corresponding parameters in 47 healthy control groups. The total antioxidant level was significantly lower (Sürücü, *et al.*, 2015). In another Turkish case-control study, evaluating 23 patients with active psoriasis found a significantly higher Plasma levels of NO and MDA levels than those in controls (Şikar *et al.*, 2011).

Multiple measures of ROS were affected in a cross-sectional case-control study, showing increased levels of serum MDA and significantly decreased serum vitamin E as well as erythrocyte catalase activity in a total 90 psoriasis patients without any drug therapy for preceding two months as compared to 90 matched healthy controls (Pujari *et al.*, 2014).

### **1.3. Statement of the problem**

Skin problems are generally the most common diseases seen in primary care setting all over the globe and its prevalence ranges from 20-50% in developing countries. Compared with other diseases, skin diseases have a lower mortality rate but can affect the wellbeing, quality of life and health conditions (Baur *et al.*, 2013). In Ethiopia, according to some studies, up to 80% of people have one or more skin diseases. It was found that skin diseases represented the sixth most frequent cause of outpatient visits to health care facilities nationwide. Many elements influence the pattern and burden of skin diseases in resource-poor settings, including poverty, environmental, and climatic factors (Gimbel *et al.*, 2013). Psoriasis is one such type of a disease among various skin diseases affecting more than 125 million people, or nearly 3% of the world's population (IFPA 2015). Unfortunately, Ethiopian specific information regarding the true epidemiology of psoriasis is sadly lacking. This suggests that we are inadequately informed to manage a silent epidemic with considerable societal impact, both socially and economically.

Certainly, evidence has shown that psoriasis can affect a patient's quality of life to a level comparable with other chronic conditions, including myocardial infarction and some cancers. It has even been independently associated with suicidal ideation. Though it has long been considered that, with psoriasis, "torture is skin deep", evidence has been mounting that it is a systemic inflammatory disease with an increased risk of mortality when severe (Oliveira *et al.*, 2015). The pathogenesis of psoriasis is not fully understood but evidence suggests that there is a strong genetic component which is mediated by abnormal T lymphocyte function. Recently studies have suggested the involvement of increased reactive oxygen species levels in pathogenesis of psoriasis (Mantovani *et al.*, 2016).

As far as the investigator is aware, no previous study exists in Ethiopia about the status of oxidative stress among psoriatic patients. Therefore, this study was designed to evaluate the total oxidant-antioxidant capacity and oxidative stress index among newly diagnosed psoriatic patients in Alert Hospital Addis Ababa.

#### **1.4. Significance of the study**

Although psoriasis is rarely life threatening, its morbidity and associated co-morbidities have a severe negative impact on the quality of life of the patients and also confer a certain socioeconomic burden. Especially in developing countries, people with psoriasis have to face severe problems with stigmatization, embarrassment, discrimination, low self-esteem and negative attitudes in general among the public, and often bear the brunt of public rejection.

Until recently, it was difficult to treat psoriasis, due to incomplete understanding of the factors behind pathogenesis of psoriasis which points out that there may be some gap in our understanding of etiopathology of psoriasis. Oxidative stress is believed to be a key factor in the pathogenesis of psoriasis as studies have suggested the involvement of increased ROS levels in psoriasis pathogenesis however it is questionable whether the observed abnormalities are responsible for the onset of psoriasis, or resultant from ongoing pathologic process. Therefore, the hypothesis of an imbalance between oxidants and antioxidants in psoriasis and its role in the pathogenesis of the disease still remain to be elucidated.

Based on increasing awareness of the importance of oxidative stress in skin diseases and the limited knowledge of the Pathophysiology , the research was conducted to understand the status of oxidative stress in a cross sectional participants diagnosed with psoriasis. This is based on an assumption that oxidative stress as one of the important factor in pathogenesis of psoriasis. Evaluation of oxidative stress therefore will provide early indication of the disease progression and institution of possible treatment for psoriatic patients to prevent the development of the associated morbidity and mortality. Besides, it can also serve as baseline information to undertake further studies on similar settings in the future.

#### **1.5. Hypothesis**

The level of oxidative stress is not associated with severity of psoriasis.



## **2. OBJECTIVE**

### **2.1. General objective**

To evaluate the status of Total Oxidant - Antioxidant Capacity and Oxidative stress index in Psoriatic patients.

#### **2.1.1. Specific objectives**

1. To assess the status of Total Oxidant Capacity in Psoriatic patients.
2. To assess the status of Total Antioxidant Capacity in Psoriatic patients.
3. To estimate Oxidative stress index in Psoriatic patients.
4. To assess the correlation of Total Oxidant - Antioxidant Capacity and Oxidative stress index with severity of Psoriatic patients.
5. To investigate the correlation between Total Oxidant and Antioxidant Capacity and Oxidative stress index in Psoriatic patients.
6. To investigate the correlation between Total Oxidant and Antioxidant Capacity and Oxidative stress index with severity of Psoriatic patients.
7. To investigate the correlation between psoriasis disease duration and the status of Total Oxidant - Antioxidant Capacity and Oxidative stress index in Psoriatic patients.

### **3. MATERIAL AND METHODS**

#### **3.1. Study design and period**

This study was a comparative Cross sectional study conducted during the period of September 2015 to January 2016.

#### **3.2. Study area and study subjects**

The study was conducted in ALERT Hospital which is located in kolfe keranio kiefle ketema and the analysis of data collected was done at the Department of Biochemistry, College of Health Sciences Addis Ababa, Ethiopia. A total of 90 participants comprising 45 newly diagnosed psoriatic patients in ALERT Hospital and who had given informed written consent and 45 healthy controls were enrolled in the study.

#### **3.3. Inclusion criteria**

Subjects who fulfilled the following criteria were involved in the study

1. Age between 18-65 years.
2. All patients who signed written informed consent
3. Newly diagnosed Patients confirmed with psoriasis

#### **3.4. Exclusion criteria**

Patients with the following conditions were excluded from the study.

1. Patients who had received any systemic or any local steroid medication or any phototherapy treat
2. Patients who don't signed written informed consent
3. Patients with diabetes, hypertension, family history of hyperlipidemia, renal and liver failure, endocrine disorders
4. Smokers and alcoholism (characterized by: a prolonged period of frequent, heavy alcohol use).

### **3.5. Sample size determination**

A convenience sampling technique was used to determine the sample size. When calculating the sample size requirements for study, a number of factors are taken into consideration including effect size, cooperation and attrition, practical constraints such as time, subject availability and finance, subgroup analysis and sensitivity of the measurement used (Suen *et al.*, 2014)

Many studies are based on relatively small sample size due to practical constraints such as time and subject availability, which often limits sample size. Examining the sample size of other comparable studies carried out internationally assessing oxidant-antioxidant levels in psoriasis sample size varies from 23 to 50 in these studies. However, power analysis and sample size calculation was not reported in any of these studies.

Based on the above reasons the present study recruited 45 psoriatic patients and 45 age-sexes matched healthy controls from Alert Hospital, Addis Ababa, Ethiopia. Psoriasis patients were graded and further divided into three groups on the basis of Psoriasis area and Severity index (PASI). Therefore, they were grouped as mild, moderate and severe.

### **3.6. Study variables**

#### **3.6.1. Dependent variables**

- Total antioxidant capacity (TAC)
- Total oxidant capacity (TOC)
- Oxidative stress index (OSI)

#### **3.6.2. Independent variable**

- Age
- Sex
- BMI
- PASI
- Psoriasis disease duration

### **3.7. Ethical Consideration**

All participants in study had information on study. A consent form was prepared with detailed explanation of objectives, risks, and benefits to the study subjects and the assurance of confidentiality of responses were given to participants. Data were collected after obtaining informed consent and agreement from the patients under study. Sample collection was performed by trained health professionals following ethical steps and procedures.

Only patients who had signed the informed consent were enrolled in the study. Strict confidentiality of all information collected was maintained and all protocols of safety were followed throughout the study. Assurances were given to the participants on confidentiality of collected data. Ethical approval was given for research from ethical committee of the Department of Biochemistry (DRERC) with a protocol number of 04/15 DRERC, and ALERT/AHRI ethics review committee /AAERC/ with a protocol number P011/15.

### **3.8. Data and Specimen collection handling and storage**

Data was collected by professional laboratory technologists, nurses and or physicians with involvement of principal investigator. Under preceding instructions psoriatic patients and healthy controls were checked regarding fasting by interview on the morning of the examination by the physician or Nurse. Pertinent information was obtained from standardized clinical files designed for the program and extracted from it using a questionnaire and other demographic details were subsequently collected (See appendix 1).

5 ml of fasting venous blood samples were obtained by vein puncture from vein of the arm of each participant using sterile syringes and transported into anti-coagulant free blood collection vacutainer tubes and allowed to stand for 30 min to coagulate. After centrifugation at 3000 rpm for 10 minutes the serum was separated by sterile by sterile pipette and transferred to 3ml epyndrophs tests tube and was incubated at  $-80^{\circ}\text{C}$  until analyzed. The serum was used for the analysis of Total Antioxidant capacity (TAC), Total Oxidant capacity (TOC) and Oxidative stress index (OSI).

### **3.9. Biochemical assays and Laboratory analysis**

#### **3.9.1. Determination of Total Oxidant Capacity (TOC)**

##### **Principle**

Absolute oxidant status of plasma was measured by robotized colorimetric estimation way for Total oxidant capacity (TOC). In this process, oxidants present in the plasma were determined by oxidation of ferrous particle by o-dianisidine complex to ferric complexes. The oxidation reaction is upgraded by glycerol molecules, which are richly present in the reaction medium. A colored compound is formed when the ferric ion reacts with xylenol orange in an acidic medium. The color strength, which was measured by ELISA microplate reader LT 4000, was correlated to the total quantity of oxidant molecules present in the plasma. The assay was calibrated with hydrogen peroxide, and the results were expressed as mmol H<sub>2</sub>O<sub>2</sub> Equiv./l. (Erel, 2005).

##### **Procedure**

Reagent 1 and 2 were prepared. Reagent 1: [114mg of xylenol orange and 8.18gm of NaCl] were dissolved in 900mL of H<sub>2</sub>SO<sub>4</sub> solution 25mM. The final reagent was composed of 150mM xylenol orange, 140mM NaCl, and 1.35M glycerol, pH 1.75; Reagent 2: [1.96 gm of ferrous ammonium sulfate and 3.17 gm of o-dianisidine dihydrochloride] were dissolved in 1000mL of H<sub>2</sub>SO<sub>4</sub> solution 25mM. TOC of serum was measured by adding 225μL reagent 1 into 35μL serum sample and then 11μL of reagent 2 was added. Finally absorbance was measured by ELISA microplate reader LT 4000 at wavelength 560 nm.

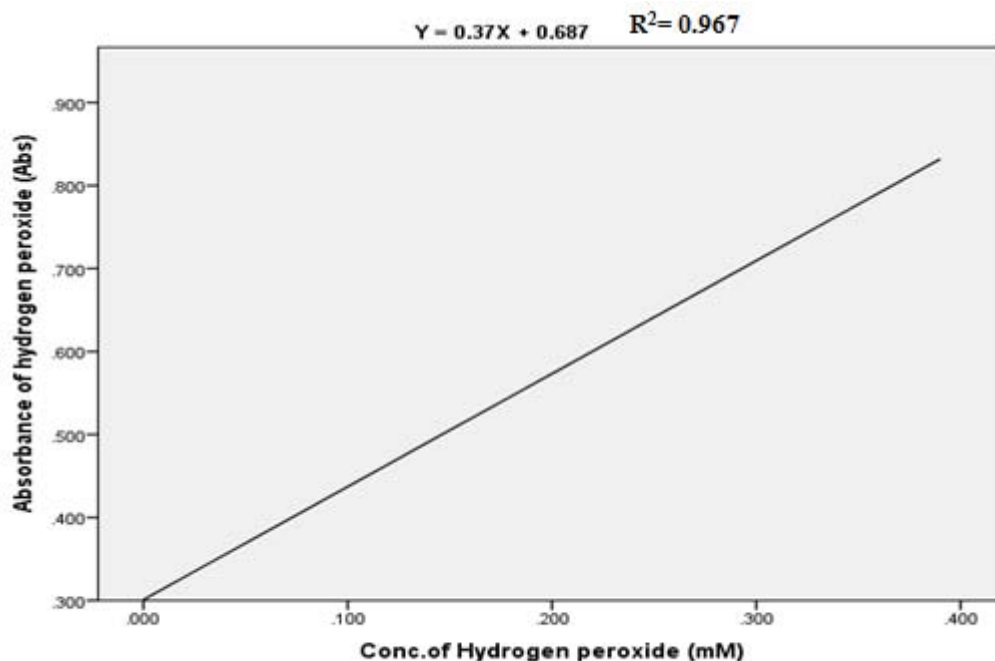


Figure 3.1: Standard curve for estimation of Total oxidant Capacity (TOC)

### 3.9.2. Determination of Total Antioxidant Capacity (TAC)

#### Principle

Total antioxidant capacity of plasma was measured by robotized colorimetric estimation system for Total antioxidant capacity (TAC). In this technique, the hydroxyl radical, the most powerful natural radical, was generated by the Fenton reaction and it responds with the colorless substrate O-dianisidine to create the dianisyl radical, which is splendid yellowish-brown in color. Upon the addition of a plasma sample, the oxidative responses started by the hydroxyl radicals present in the reaction are scavenge by the antioxidant agents of the plasma, keeping the color change and consequently giving a viable estimation of TAC. The assay was calibrated with Trolox, and test results were expressed as mmol TroloxEq/l (Koracevic *et al.*, 2001).

## Procedure

For measurement of total antioxidant capacity reagent 1 and 2 were prepared. Reagent 1: 3.17gm of orthodiansidine dichloride and 0.01764gm of  $\text{Fe}(\text{NH}_4)_2(\text{SO}_4)_2 \cdot 6\text{H}_2\text{O}$  were dissolved in 1000mL of clark and Lubs solution. Reagent 2: 0.641mL of  $\text{H}_2\text{O}_2$  solution (35%) was diluted in 1000ml with clark and lubs solution. TAC of serum was measured by adding 200  $\mu\text{L}$  reagent 1 to 5  $\mu\text{L}$  serum sample and then 20  $\mu\text{L}$  of reagent 2 was added. Finally absorbance was measured by ELISA microplate reader LT 4000 at wavelength 660 nm.

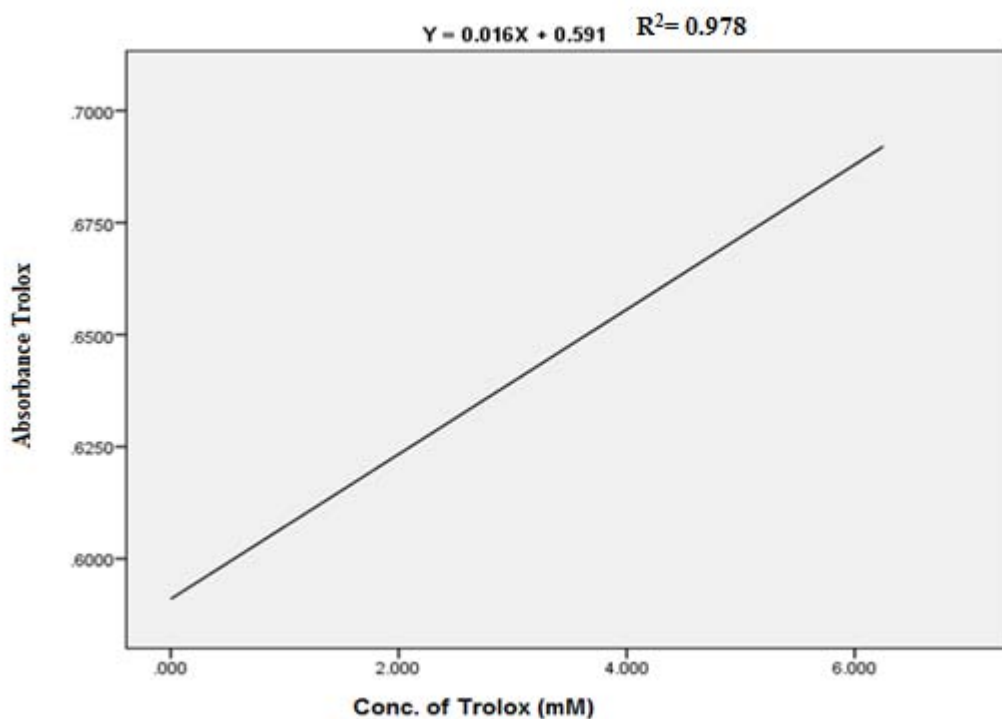


Figure 3.2: Standard curve for estimation of Total antioxidant capacity (TAC)

### 3.9.3. Measurement of oxidative stress index (OSI)

The TOC/TAC ratio provides the OSI, which is an indicator of the degree of oxidative stress. OSI value was calculated using the following formula:  $\text{OSI (arbitrary unit)} = \text{TOC (mmol H}_2\text{O}_2 \text{ equivalent/l)} / \text{TAC (mmol Trolox equivalent/l)}$  (Erel et al., 2005).

### **3.10. Statistical Analysis**

All data was expressed as mean  $\pm$ SD. The statistical significance was evaluated by one way ANOVAs. All the data were entered to Epi Info (version 3.5.1, 2008) and statistical analysis was performed by using SPSS (version 20.0, 2007, America). While post hoc was used to assess statistical comparisons between groups. Correlations between variables were calculated by Pearson's correlation test. A P-value of less than  $<0.05$  was considered statistically significant.

### **3.11. Dissemination of results**

Clinical findings of the study will be submitted and presented to Department of Biochemistry, Addis Ababa University. Based on the findings conclusions and recommendations will also forwarded to Executive bodies and organizations for further action.

### **3.12. Quality control**

The English version of the questionnaire was translated into the local language, Amharic. The data collectors were professional nurses and they were trained about data collection procedure and during the data collection the patient history and other information from the patients' medical record cards were cross checked. The questionnaire was pre-tested to check consistency a week before the actual data collection was done.



## 4. RESULTS

### 4.1. Demographic characterization

Among 90 participants 45 of them were psoriatic patients (24 males and 21 females) with mean age  $39.69 \pm 12.67$  and BMI of  $21.84 \pm 2.63$ . The rest of 45 were age-and sex- matched healthy subjects as a control group (26 males and 19 females) with mean age  $38.53 \pm 12.05$  and BMI of  $23.38 \pm 2.79$ . The clinical severity was determined according to the Psoriasis Area and Severity Index (PASI) with mean score  $20.08 \pm 16.94$ . Psoriasis patients were further graded according to the PASI as, mild (22 patients), moderate (15 patients) and severe (8 patients). Disease duration of the patients ranged from 3 weeks to 5 years with mean  $2.39 \pm 1.61$  (Table 4.1).

Table 4.1: Demographic characteristics and clinical features of psoriatic patients and healthy Controls

	Healthy Controls(n= 45)	Psoriatic Patients (n=45)
Sex [M/F]	26/19	24/21
Age [years]	$38.53 \pm 12.05$	$39.69 \pm 12.67$
Height[m]	$1.60 \pm 0.09$	$1.62 \pm 0.08$
Weight[kg]	$60.13 \pm 8.86$	$57.53 \pm 9.36$
Body mass index [ $\text{kg}/\text{m}^2$ ]	$23.38 \pm 2.79$	$21.84 \pm 2.63$
PASI		$20.08 \pm 16.94$
Mild		22(48%)
Moderate		15(33%)
Sever		8(17%)
Type of lesion		
Plaque		38(84%)
Erythema		1(2%)
Papules		3(6%)
Pustule		3(6%)
Psoriasis duration [years]		$2.39 \pm 1.61$

Results presented as mean  $\pm$  SD and frequency (%) (Except male: female ratio)

## 4.2. Estimated biochemical parameters

### 4.2.1. Total oxidant- antioxidant between psoriasis and health control

Total oxidant capacity (TOC) and oxidative stress index (OSI) ( $13.94 \pm 2.28$ ,  $1.32 \pm 0.59$ ) were significantly ( $p < 0.01$ ) higher in psoriatic patients compared to control subjects ( $11.60 \pm 1.43$ ). On the other hand, the levels of Total Antioxidant capacity (TAC) were significantly decreased ( $p < 0.01$ ) in psoriatic patients ( $1.15 \pm 0.21$ ) compared to normal healthy controls. ( $1.36 \pm 0.16$ ) (Table 4.2).

Table 4.2: TOC, TAC and OSI levels in patients versus controls

Parameter	Healthy Contorts (n=45)	Psoriatic Patients (n=45)	p-value
TOC [mmol H <sub>2</sub> O <sub>2</sub> eqv./l]	$11.60 \pm 1.43$	$13.94 \pm 2.28$	$< 0.01$
TAC [mmol Trolox eqv./l]	$1.36 \pm 0.16$	$1.15 \pm 0.21$	$< 0.01$
OSI [AU]	$0.86 \pm 0.15$	$1.32 \pm 0.59$	$< 0.01$

Results presented: as mean  $\pm$  SD, TAC – Total Antioxidant capacity, TOC – Total Oxidant capacity, OSI – Oxidative Stress Index.

#### 4.2.2. Total oxidant- antioxidant status and severity of disease

Present study shows that, TOC and OSI levels were significantly ( $P<0.01$ ) increased in moderate psoriatic patients than the patients presenting with mild psoriasis, whereas, severe psoriatic patients exhibited significant increase in TOC and OSI ( $P<0.001$ ) as compared to moderate psoriatic patients. Furthermore, TAC activity in moderate psoriatic patients were significantly ( $P<0.01$ ) lower than the patients presenting with mild psoriasis, whereas, severe psoriatic patients exhibited significant decrease in TAC ( $P<0.01$ ) as compared to moderate psoriatic patients (Table 4.3).

Table 4.3: Changes in levels of TOC, TAC and OSI within different groups of psoriasis patients

Parameters	Control(n=45)	Psoriatic patients ( n=45)		
		Mild(n=22)	Moderate(n=15)	Sever(n=8)
TOC [mmol H <sub>2</sub> O <sub>2</sub> eqv./l]	11.60 ±1.43	12.47±1.11 <sup>b1c2</sup>	14.03±1.52 <sup>a2c2</sup>	17.81±0.62 <sup>a2</sup>
TAC[mmol Trolox eqv./l]	1.36±0.16	1.29±0.52 <sup>b1c2</sup>	1.14±0.04 <sup>a2c2</sup>	0.76±0.16 <sup>a2</sup>
OSI [AU]	0.86±0.15	0.96±0.10 <sup>b1c2</sup>	1.26±0.16 <sup>a2c2</sup>	2.43±0.57 <sup>a2</sup>

Results presented as mean ± SD, TAC – Total Antioxidant capacity, TOC – Total Oxidant capacity , OSI – Oxidative stress index ; <sup>a</sup>compared to control, <sup>b</sup>compared to moderate, <sup>c</sup>compared to severe; <sup>1</sup> $p<0.01$ , <sup>2</sup> $p<0.001$

### 4.2.3. Psoriasis disease duration and changes in Total oxidant- antioxidant status

Disease duration of psoriatic patients were ranged from 3 weeks to 5 years with mean  $2.39 \pm 1.61$  (Table 4.1) and further categorized into <1, 1-3 and > 3 years. Psoriatic patients with a year duration greater than 3 years have showed a significant increase in TOC and OSI ( $P < 0.001$ ) with a decreased TAC levels when compared to patients with a year duration less than a year and patients between 1 to 3 years (Table 4.4)

Table 4.4: Psoriasis disease duration and changes in levels of TOC, TAC and OSI in psoriatic patients.

Parameters	Psoriasis disease duration [ years]		
	<1	1-3	>3
TOC [mmol H <sub>2</sub> O <sub>2</sub> eqv./l]	12.41±1.23 <sup>c1</sup>	12.39±0.87 <sup>c1</sup>	15.86±1.89 <sup>[ba]1</sup>
TAC [mmol Trolox eqv./l]	1.29±0.05 <sup>c1</sup>	1.25±0.08 <sup>c1</sup>	0.98±0.21 <sup>[ba]1</sup>
OSI [AU]	0.96±0.11 <sup>c1</sup>	0.1±0.08 <sup>c1</sup>	1.75±0.68 <sup>[ba]1</sup>

Results presented as mean  $\pm$  SD, TAC – Total Antioxidant capacity, TOC – Total Oxidant capacity, OSI – Oxidative stress index; <sup>a</sup>compared to <1, <sup>b</sup>compared to 1-3, <sup>c</sup>compared to >3; <sup>1</sup> $p < 0.001$

#### 4.2.4. Correlations between Total oxidant –antioxidant in psoriatic patients

The levels of TOC with TAC showed a negative correlation with significant ( $p < 0.001$ ) association. On the other hand TOC showed a positive correlation with levels of OSI with significant ( $p < 0.001$ ) association in psoriatic patients (Table 4.5).

Table 4.5: Correlations between TOC levels and TAC, OSI levels in psoriatic patients

Parameter	r	P-value
TOC and TAC	-0.81	<0.001
TOC and OSI	0.86	<0.001

Total Antioxidant capacity, TOC – Total Oxidant, OSI – Oxidative Stress Index,  $**p < 0.001$ , Results were expressed in pearson correlation coefficient (-) negative and (+) Positives correlation

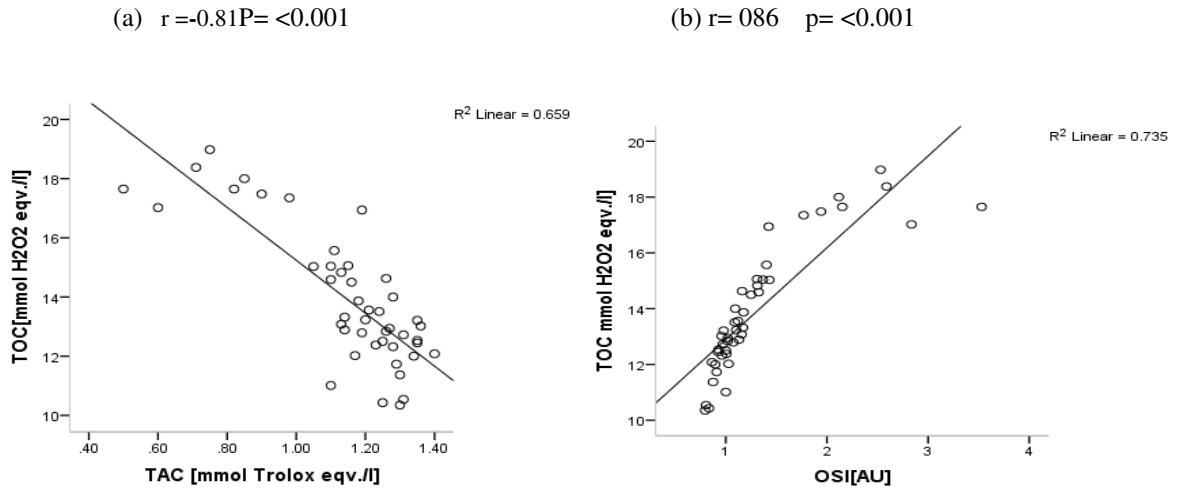


Figure 4.1: Regression fit of (a) TOC Vs TAC and (b) TOC Vs OSI

#### 4.2.5. Correlations between Total oxidant –Antioxidant and severity of disease

It was found a negative correlation between TOC and TAC but it was not significant ( $p > 0.05$ ) among mild, moderate and severe psoriatic patients. Conversely a positive correlation was found between TOC and OSI among the three groups with significant correlation in mild and moderate ( $p < 0.001$ ) whereas, severe psoriatic patients exhibited insignificant ( $P > 0.05$ ) correlation (Table 4.6).

Table 4.6: Correlations between TOC levels and TAC, OSI levels among the different groups of psoriasis patients

	Mild	Moderate	Severe
TOC and TAC	-0.22 <sup>a</sup>	-0.047 <sup>a</sup>	-0.018 <sup>a</sup>
TOC and OSI	+0.927 <sup>b</sup>	+0.949 <sup>b</sup>	+0.065 <sup>a</sup>

Results were expressed in Pearson correlation coefficient (-) negative and (+) Positive correlation among variables a - $p > 0.05$ , b  $p < 0.001$

## 5. DISCUSSION

Skin cells are a major target of oxidative stress mainly due to ROS originating from the environment and skin metabolism itself. Although endogenous antioxidants attenuate the harmful effects of ROS, increased or prolonged presence of free radicals can override ROS defense mechanisms and mediate numerous cellular responses that contribute to the development of a variety of skin disorders, including psoriasis (Zhou *et al.*, 2009).

The histological studies of epidermis of psoriasis patients revealed that polymorphonuclear leukocytes get infiltrated in it. These polymorphonuclear leukocytes might be releasing cytokines like TNF- $\alpha$  and interleukins in the surrounding tissues, for which ROS are known to be the mediators of their action. This in turn might trigger the inflammatory reactions that are the basis of psoriasis (Balato *et al.*, 2012).

Generation of ROS from neutrophils, keratinocytes and fibroblasts can contribute to neutrophil activation which may play an important role in psoriatic process. ROS can act as second messengers in the induction of several biological responses such as the activation of NF-kB or AP-1, the generation of cytokines, the modulation of signaling pathways and the activation of peroxisome proliferator-activated receptors (shafaq *et al.*, 2012). It is now known that psoriasis can occur due to abnormalities in essential fatty acid metabolism, lymphokine secretion, free radical generation, lipid peroxidation and eicosanoid metabolism. Toxic effects of free oxygen radicals induce through production of lipid peroxidation and the reduction of natural antioxidants. An impaired antioxidant skin barrier may result in an increase of free oxygen radicals in psoriatic plaques (Ala *et al.*, 2013).

Although there have been extensive studies on the roles of serum oxidants and antioxidants levels in psoriasis, their importance in the etiology or in the enhancement of the disease remains controversial (Gurupadappa *et al.*, 2011).

The present work investigated the possible role of oxidative stress in the pathogenesis of psoriasis through determination of serum TOC, TAC and OSI and their correlations with disease severity as expressed by PASI score in 45 psoriatic patients and age and sex matched 45 healthy individuals.

Oxidant generation is part of normal human metabolism and host defense mechanism, however when produced in excess oxidants and their products are known to play an important role in the pathogenesis of chronic inflammatory disorders (Bouayed and Bohn, 2010). Many free radicals are highly reactive and can either donate an electron to or extract an electron from other molecules therefore behaving as oxidants or reductants. The most important free radicals in many diseases are oxygen derivatives, particularly superoxide anion and the hydroxyl radical. The hydroxyl radical or a closely related species is probably the final mediator of most free radical induced cellular damage (Rahman *et al.*, 2012).

It is believed that in addition to genetic predisposition, ROS and mediated oxidative stress may play a role in the pathogenesis of inflammatory skin diseases, such as psoriasis. It is also believed that the ROS produced by keratinocytes, fibroblasts and endothelial cells causes chemotaxis and accumulation of neutrophils, as a result there is a production of superoxide in phagocytic reactions in psoriatic lesions (Güven *et al.*, 2013). Overproduction of ROS during the inflammatory process in psoriasis may result in oxidative stress leading to damage of cellular structures such as lipids, proteins and DNA and interference with production of cytokines such as IL-1 and TNF- $\alpha$ . It is also responsible for a decrease in the cAMP/cGMP ratio leading to epidermal hyperproliferation (Nassiri *et al.*, 2009).

The present work shows a significantly ( $p < 0.001$ ) increased concentration of TOC was observed in psoriatic patients ( $13.94 \pm 2.28$ ) compared to healthy controls ( $11.60 \pm 1.43$ ) (Table 4.2). This significantly elevated TOC could be due to high free radical load and insufficient antioxidant which consequently indicates a high oxidative stress condition in psoriatic patients. In concordance with this study, Sürücü *et al.*, (2015), Karababa *et al.*, (2013), Gabr *et al.*, (2012) and Hashemi *et al.*, (2010) have reported increased TOC levels in psoriatic patients as compared to healthy controls.

There are compelling evidences which indicate that oxidative stress drives the production of oxidation products, such as MDA, which can denature proteins, alter apoptosis, and influence the release of proinflammatory mediators, such as cytokines, which may be critical for the induction of some inflammatory skin diseases (Aly and shahin, 2010).



This argument is substantiated by reports sikar *et al.*, (2012), Mohamed (2014), Pujari *et al.*, (2010), Attwa and Swelam, (2011) which showed that psoriatic patients are normally in oxidative imbalance as a consequence of elevated peroxidation products like MDA.

The infiltrated and activated leukocytes might lead to release of ROS via processes like respiratory burst. Polymorphonuclear (PMN) leukocytes have the potential to damage surrounding tissue by releasing superoxide anion radical produced via NADPH oxidase/myeloperoxidase which further gives rise to other activated oxygen species which all together were known to induce lipid peroxidation (Andrade *et al.*, 2012). ROS in turn also stimulates PMN recruitment by increasing PMN adhesion to endothelium. The increased generation of ROS, by increased infiltration and activation of PMNs might target cellular polyunsaturated fatty acids for lipid peroxidation, which might be the indicative factor for increased concentration of MDA in serum of psoriatic patients (Kiran *et al.*, 2015).

Nitric oxide is a gaseous free radical that is released by the family of NO synthetase enzymes. It is a potent vasodilator, thus contributing considerably to the cardinal signs of inflammation. It is also known to exhibit cytotoxic effects in human skin (Aly and shahin, 2010). As a potential regulator of keratinocyte growth and differentiation, the multifunctional signaling molecule NO<sup>•</sup> has been considered to be a strong candidate in the pathogenesis of psoriasis. NO<sup>•</sup> is an important marker of inflammation. Expression of iNOS is involved in the pathogenesis of cutaneous inflammation in psoriasis which is confirmed by increase in mRNA expression of iNOS in skin lesions as compared to uninvolved skin as a result nitrite levels and nitrite–nitrate ratios appear to be good indicators for the increased NO<sup>•</sup> production in psoriatic patients( Kadam *et al.*, 2010). This concept is supported by Sikar *et al.*, (2012) and Kadam *et al.*, (2010) who demonstrated significantly elevated serum NO levels in different types of psoriatic patients when compared to normal individuals indicating deranged level of oxidative stress in psoriatic patients.

Results of the present study showed that the values of serum TOC were significantly higher ( $p < 0.001$ ) in all psoriasis patients than normal healthy controls. It was further found that the concentrations of serum TOC were increased with severity of the psoriasis (Table 4.3). The values of TOC were significantly higher in, moderate psoriatic patients than mild psoriatic patients ( $p < 0.001$ ) as well as severe psoriatic patients than moderate psoriatic patients ( $p < 0.001$ ) respectively and also the research considered increased TOC levels as psoriasis year duration increases ( $P < 0.001$ ) thereby confirming that the load was decidedly higher with the progression of psoriasis (Table 4.4). In very early phase of developing psoriasis lesions, macrophages were seen within the epidermis followed by lymphocytes. During subsequent development neutrophils began to appear between the upper layers forming pockets (Micro-abscesses). Neutrophil migration into the epidermis was most pronounced in active disease and occurred in a rhythmic pattern (Pujari *et al.*, 2014). These presumptions were well supported by findings of Attwa and Swelam, (2011) and Gurupadappa *et al.*, (2011).

Skin cells are equipped with both enzymatic [superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GP)] and non-enzymatic mechanisms (vitamin C, vitamin E, carotenoids, thiol antioxidants, flavinoids, selenium and others) to deal with the harmful effects of ROS (Şikar *et al.*, 2012). TAC is a biochemical parameter suitable for evaluating known and unknown antioxidants and their synergistic interaction and consumption by normal or increased levels of ROS production therefore assessed giving an insight into the delicate balance between oxidants and antioxidants in living cells (Bansal and Bilaspuri, 2010). Its value expresses the number of antioxidant molecules present in serum. An individual antioxidant level or activity indicates the antioxidant characteristics of only one antioxidant, whereas TAC may represent the total antioxidant characteristics of all antioxidants found in the serum (Kayar *et al.*, 2015).

Antioxidants can protect the epidermis from the events that contribute to epidermal toxicity and diseases. Deficiencies in any of the antioxidant defense system can cause a reduction in the total antioxidant status of an individual (Wagener *et al.*, 2013).

The results of this study showed statistically significant ( $p < 0.001$ ) differences in TAC values ( $1.15 \pm 0.21$ ) between psoriatic patients and healthy controls ( $1.36 \pm 0.16$ ) (Table 4.2). This is in concordance with the studies of Sürücü *et al.*, (2015), Karababa *et al.*, (2013), Gabr *et al.*, (2012) and Hashemi *et al.*, (2010) which suggest that antioxidant defense mechanisms are exhausted in psoriatic patients. The decreased TAC may be possibly due to depressed state of antioxidant system or due to the exaggerated inflammatory processes and oxidative stress in these patients. Antioxidants prevent oxidative injury of structural lipids and proteins contributing to barrier integrity, which is essential for healthy skin condition. This suggests that cellular redox environment plays a pivotal role in skin homeostasis and that skin disease could result from an imbalance between pro-oxidant and antioxidant stimuli (Galassetti *et al.*, 2012). However, in a study performed by Nassiri *et al.*, (2009) and his colleagues found no significant difference in serum TAC levels between cases and controls. They explained this by the younger age of Iranian patients and also because Iranian regular diet which contains more fruit and vegetable and less saturated fatty acid in comparison with western diets.

One important line of defense is a system of enzymes, including glutathione peroxidase, superoxide dismutase (SOD) and catalase (CAT) which decreases the concentration of the most harmful oxidants. During oxidative stress there can be the overload of superoxide radicals in the psoriasis (Benedetti *et al.*, 2012). Since, the natural antioxidant defense system SOD limits the development of inflammatory and immune processes, the tendency to decreased SOD activity in psoriasis patients is one of the reasons for the aggravated dermatological status. This over formation of the superoxide radical also was proposed to inhibit the activity of catalase (Shamsi *et al.*, 2010). This might be one of the reasons for significantly low serum vitamin E, CAT and SOD activities in psoriatic patients compared to healthy controls which are explained by the work of Reshma *et al.*, (2011), Mohammed *et al.*, (2014) and Pujari *et al.*, (2014).

The present study serum TAC levels were decreased significantly in; moderate psoriasis patients than mild psoriasis patients ( $p < 0.001$ ) whereas, severe psoriatic patients exhibited significant decrease in TAC activity ( $P < 0.001$ ) as compared to moderate psoriatic patients (Table 4.3).

Moreover a decreased TAC levels was observed as psoriasis year duration increases ( $P < 0.001$ ) (Table 4.4). The low TAC level can be explained with excessive depletion of the antioxidant capacity caused by free radical elimination to reduce the detrimental effects of ROS and their products in order to maintain pro-oxidant-antioxidant homeostasis. Pujari *et al* (2011) also found positive correlation between increased MDA, NO in serum of psoriatic patients with severity of psoriasis whereas negative correlation between decreased TAC with severity of psoriasis. The reduced level of TAC may reflect a decrease in the antioxidant capacity.

When the oxidant/antioxidant balance is tilted towards oxidants and oxidative stress arises, there is a significant negative correlation between the TAC and TOC values (Kayar *et al.*, 2015). In the present investigation a negative correlation was demonstrated between the TAC and TOC, which is one of the scales indicating the severity of psoriasis. This corresponds to previously reported findings (Sürücü *et al.*, (2015) and Karababa *et al* (2013). These findings may indicate the importance of the anti-oxidant system in the pathogenesis and severity of psoriasis.

Oxidative stress index (OSI), a proportional value between TAC and TOC, is a recently investigated parameter that is affected directly by oxidant and antioxidant status and that can present the degree of oxidative stress more clearly. Because OSI serves as an element of balance, its use is likely to be more important than TAC and TOC values in interpreting the study results. Therefore, OSI may be a useful and practical parameter for evaluating oxidative injury in psoriatic patients (Kose *et al.*, 2014).

The OSI estimated in this study was significantly ( $p < 0.001$ ) higher in psoriatic patients ( $1.32 \pm 0.59$ ) than in healthy controls ( $0.86 \pm 0.15$ ) (Table 4.2). This indicates the exact degrees of imbalance of Oxidative stress towards the oxidant status. This finding is in keeping with most previously published findings as Gabr and Al-Ghadir (2012) and Hashemi *et al.*, (2010) have reported high OSI values in psoriasis patients compared with control groups. On the basis of these results, we can deduce that a compromised antioxidant defense mechanism, accompanied by increased oxidant levels and OSI values in psoriasis patients, might play an important role in the pathogenesis of this disease.

Present study showed that there was statistically significant increase in OSI value in mild ( $P < 0.01$ ), moderate ( $P < 0.01$ ) and severe ( $P < 0.01$ ) psoriasis patients when compared to healthy controls. In addition, OSI in moderate psoriatic patients were significantly ( $P < 0.01$ ) lower than the patients presenting with mild psoriasis, whereas, severe psoriatic patients exhibited significant decrease in OSI ( $P < 0.01$ ) as compared to moderate psoriatic patients (Table 4.3) and also the research considered increased OSI as psoriasis year duration increases ( $P < 0.001$ ) (Table 4.4). Similar results were reported by Sürücü *et al.*, (2015) Gabr and Al-Ghadir (2012). This might be due to the prolonged release of excess ROS in the skin cells which can aggravate inflammatory injury and promote chronic inflammation. The decrease in TAC and increase in TOC was more pronounced in severe patients comparing to mild and moderate indicating that the anti-oxidants were nearly completely utilized to scavenge the superoxide free radicals.

## **6. CONCLUSION**

The presented study concluded oxidative stress in psoriatic patients, indicated by increased TOC as well as OSI and decreased TAC levels when compared to healthy controls. Furthermore the study found worsened oxidant and antioxidant status according to the severity and duration of disease which may be the crucial point in the pathogenesis of psoriasis and need further elucidations. These findings suggest that a weakened antioxidant defense system may play a significant role in the increased oxidative stress found in psoriatic patients. Therefore, it is possible to conclude that increase of oxidative stress in psoriatic patients as severity and duration of the disease increases. Moreover the study also supports the possibility of involvement of oxidative stress in pathogenesis of psoriasis.

## **7. RECOMMENDATION**

- Ethiopian specific information regarding the true epidemiology of psoriasis has to be studied adequately to manage the health, societal and economic impact of psoriasis nationwide.
- Psoriatic patients have to elucidate their feeding habit with high intake of fruit and vegetables, legumes, grains and cereals, fish and antioxidant levels with their therapy.
- Further research should continue to enhance the understanding of the genetics and causes of psoriasis in Ethiopia and result in improvements in diagnosing and treating this disease.
- Estimating DNA metabolites and important enzymatic antioxidants (superoxide dismutase and catalase activities) needed to be considered in subsequent studies in the future.
- ROS lie downstream as inflammatory driver therefore an effective biomarker of ROS may have even greater potential.
- Finally, of considerable interest is the possibility of using this information to develop novel strategies for diagnosis, prognosis and treatment of psoriasis patients are warranted.

## **8. LIMITATION**

Small patient numbers: Invariably, this is only a cross sectional study with small number of patients. However, a cohort study with larger number of patients needs to be attempted in Ethiopia. Another limitation of this study is that we did not use an objective method to assess stress reactions with reference to nutrition.



## 9. REFERENCES

- Abdel-Mawla, M.Y., Nofal, E., Khalifa, N., Abdel-Shakoor, R. and Nasr, M., (2011). *Role of Oxidative Stress in Psoriasis: An Evaluation Study*.
- Akasaka, E., Takekoshi, S., Horikoshi, Y., Toriumi, K., Ikoma, N., Mabuchi, T., Tamiya, S., Matsuyama, T. and Ozawa, A., (2010) *Protein oxidative damage and heme oxygenase in sunlight-exposed human skin: Roles of MAPK responses to oxidative stress*. Tokai J Exp Clin Med, 35(4), pp.152-164.
- Ala, S., Shokrzadeh, M., Golpour, M., Salehifar, E., Alami, M. and Ahmadi, A., (2013). *Zinc and copper levels in Iranian patients with psoriasis: a case control study*. Biological trace element research, 153(1-3), pp.22-27.
- Alexis, A.F. and Blackcloud, P., (2014) *Psoriasis in skin of color: epidemiology, genetics, clinical presentation, and treatment nuances*. The Journal of clinical and aesthetic dermatology, 7(11), p.16.
- Al-Shobaili, H.A. and Qureshi, M.G., (2013). *Pathophysiology of Psoriasis: Current Concepts*. INTECH Open Access Publisher.
- Aly, D.G. and Shahin, R.S., (2010) *Oxidative stress in lichen planus*. Acta Dermatoven APA, 19(1), pp.3-11.
- Amaro-Ortiz, A., Yan, B. and D'Orazio, J.A., (2014) *Ultraviolet radiation, aging and the skin: prevention of damage by topical cAMP manipulation*. *Molecules*, 19(5), pp.6202-6219.
- Anderson, A., Bowman, A., Boulton, S.J., Manning, P. and Birch-Machin, M.A., (2014) *A role for human mitochondrial complex II in the production of reactive oxygen species in human skin*. Redox biology, 2, pp.1016-1022.
- Andrade, M.F., Kabeya, L.M., Azzolini, A.E.C., Santos, E.O., Figueiredo-Rinhel, A.S., Paris, M.R., Emery, F.S., Pupo, M.T. and Lucisano-Valim, Y.M., (2013) *3-Phenylcoumarin derivatives selectively modulate different steps of reactive oxygen species production by immune complex-stimulated human neutrophils*. International immunopharmacology, 15(2), pp.387-394.
- Armstrong, A.W., Voyles, S.V., Armstrong, E.J., Fuller, E.N. and Rutledge, J.C., (2011). *Angiogenesis and oxidative stress: common mechanisms linking psoriasis with atherosclerosis*. Journal of dermatological science, 63(1), pp.1-9.

- Attwa, E. and Swelam, E., (2011). *Relationship between smoking-induced oxidative stress and the clinical severity of psoriasis*. Journal of the European Academy of Dermatology and Venereology, 25(7), pp.782-787.
- Augustin, M. and Radtke, M.A., (2014). *Quality of life in psoriasis patients*. Expert review of pharmacoeconomics & outcomes research, 14(4), pp.559-568.
- Avci, E., Akarlan, Z.Z., Erten, H. and Coskun-Cevher, S., (2014). *Oxidative stress and cellular immunity in patients with recurrent aphthous ulcers*. Brazilian Journal of Medical and Biological Research, 47(5), pp.355-360.
- Balato, A., Ayala, F., Schiattarella, M., Megna, M., Balato, N. and Lembo, S., (2012). *Pathogenesis of psoriasis: the role of pro-inflammatory cytokines produced by Keratinocytes*. INTECH Open Access Publisher.
- Bansal, A.K. and Bilaspuri, G.S., (2010). *Impacts of oxidative stress and antioxidants on semen functions*. Veterinary medicine international
- Baur, B., Sarkar, J., Manna, N. and Bandyopadhyay, L., (2013). *The pattern of dermatological disorders among patients attending the skin OPD of a tertiary care hospital in Kolkata, India*. IOSR Journal of Dental and Medical Sciences (JDMS), 4(3), pp.04-09.
- Bazela, K., Solyga-Zurek, A., Debowska, R., Rogiewicz, K., Bartnik, E. and Eris, I., (2014) *L-Ergothioneine protects skin cells against UV-induced damage—a preliminary study*. Cosmetics, 1(1), pp.51-60.
- Benedetti, S., Tagliamonte, M.C., Catalani, S., Primiterra, M., Canestrari, F., De Stefani, S., Palini, S. and Bulletti, C., (2012). *Differences in blood and semen oxidative status in fertile and infertile men, and their relationship with sperm quality*. Reproductive biomedicine online, 25(3), pp.300-306.
- Bhattacharya, S., (2015). *Reactive Oxygen Species and Cellular Defense System*. In Free Radicals in Human Health and Disease (pp. 17-29). Springer India.
- Bito, T. and Nishigori, C., (2012). *Impact of reactive oxygen species on keratinocyte signaling pathways*. Journal of dermatological science, 68(1), pp.3-8.
- Boda, D., Negrei, C., Toderescu, C.D. and Nicolescu, F., (2013). *evaluation of certain oxidative stress parameters in patients with psoriasis and psoriatic arthritis*. Studia Universitatis" Vasile Goldis" Arad. Seria Stiintele Vietii (Life Sciences Series), 23(2), p.243.

- Bouayed, J. and Bohn, T., (2010). *Exogenous antioxidants—double-edged swords in cellular redox state: health beneficial effects at physiologic doses versus deleterious effects at high doses*. *Oxidative medicine and cellular longevity*, 3(4), pp.228-237.
- Chandran, V. and Raychaudhuri, S.P., (2010). *Geoepidemiology and environmental factors of psoriasis and psoriatic arthritis*. *Journal of autoimmunity*, 34(3), pp.J314-J321.
- Chandran, V., (2010). *Genetics of psoriasis and psoriatic arthritis*. *Indian journal of dermatology*, 55(2), p.151.
- Chong, L., (2010) *Is Psoriasis a Cutaneous Disease or Systemic Disease?*. *Medical Bulletin*, 15(11).
- Chu, C.C., Di Meglio, P. and Nestle, F.O., (2011). *Harnessing dendritic cells in inflammatory skin diseases*. In *Seminars in immunology*(Vol. 23, No. 1, pp. 28-41). Academic Press.
- Denat, L., Kadarkar, A.L., Marrot, L., Leachman, S.A. and Abdel-Malek, Z.A., (2014). *Melanocytes as instigators and victims of oxidative stress*. *Journal of Investigative Dermatology*, 134(6), pp.1512-1518.
- Di Meglio, P., Perera, G.K. and Nestle, F.O., (2011) *The multitasking organ: recent insights into skin immune function*. *Immunity*, 35(6), pp.857-869.
- Diallo, M., (2012). *Psoriasis Epidemiology*. *Journal of Clinical Case Reports*,2012.
- Dickinson, B.C. and Chang, C.J., (2011) *Chemistry and biology of reactive oxygen species in signaling or stress responses*. *Nature chemical biology*,7(8), pp.504-511.
- Dilnawaz, M., Sadiq, S., Shaikh, Z.I., Aziz, H., Khan, S.A. and Jawad, B., (2013). *Clinical audit: baseline Psoriasis Area and Severity Index (PASI) and Dermatology Life Quality Index (DLQI) assessment of psoriasis patients*. *Journal of Pakistan Association of Dermatologists*, 23(4), pp.407-411.
- EL-RAHMAN, S.H.A. and EL-REFAEI, A.M., (2014). *Measurement of Serum Nitric Oxide in Different Types of Psoriasis*.
- Erel, O., (2005) *A new automated colorimetric method for measuring total oxidant status*. *Clinical biochemistry*, 38(12), pp.1103-1111.
- Gabr, S.A. and Al-Ghadir, A.H., (2012). *Role of cellular oxidative stress and cytochrome c in the pathogenesis of psoriasis*. *Archives of dermatological research*, 304(6), pp.451-457.

- Galassetti, P., (2012). *Inflammation and oxidative stress in obesity, metabolic syndrome, and diabetes*. Experimental diabetes research.
- Garcia-Bailo, B., El-Soheby, A., Haddad, P.S., Arora, P., BenZaied, F., Karmali, M. and Badawi, A., (2011). *Vitamins D, C, and E in the prevention of type 2 diabetes mellitus: modulation of inflammation and oxidative stress*. Biologics, 5(1), pp.7-19.
- Gimbel, D.C. and Legesse, T.B., (2013). *Dermatopathology practice in Ethiopia*. Archives of Pathology and Laboratory Medicine, 137(6), pp.798-804.
- Guyen, B., Can, M., Genc, M. and Koca, R., (2013). *Serum prolidase activity in psoriasis patients*. Archives of dermatological research, 305(6), pp.473-476.
- H Kashou, A. and Agarwal, A., (2011). *Oxidants and antioxidants in the pathogenesis of HIV/AIDS*. The open reproductive science journal, 3(1).
- Hashemi, M., Mehrabifar, H., Daliri, M. and Ghavami, S., (2010). *Adenosine deaminase activity, trypsin inhibitory capacity and total antioxidant capacity in psoriasis*. Journal of the European Academy of Dermatology and Venereology, 24(3), pp.329-334.
- Hay, R.J., Johns, N.E., Williams, H.C., Bolliger, I.W., Dellavalle, R.P., Margolis, D.J., Marks, R., Naldi, L., Weinstock, M.A., Wulf, S.K. and Michaud, C., 2014. *The global burden of skin disease in (2010): an analysis of the prevalence and impact of skin conditions*. Journal of Investigative Dermatology, 134(6), pp.1527-1534.
- Hsu, L. and Armstrong, A.W., (2014). *JAK inhibitors: treatment efficacy and safety profile in patients with psoriasis*. Journal of immunology research, 2014.
- IFPA *International Federation of Psoriasis Associations 2015*
- Jean, J. and Pouliot, R., (2010). *In vivo and in vitro models of psoriasis*. INTECH Open Access Publisher.
- Jiang, F., Zhang, Y. and Dusting, G.J., (2011). *NADPH oxidase-mediated redox signaling: roles in cellular stress response, stress tolerance, and tissue repair*. Pharmacological reviews, 63(1), pp.218-242.
- Jiang, S., Hinchliffe, T.E. and Wu, T., (2015). *Biomarkers of an autoimmune skin disease—psoriasis*. Genomics, proteomics & bioinformatics, 13(4), pp.224-233.
- Kadam, D.P., Suryakar, A.N., Ankush, R.D., Kadam, C.Y. and Deshpande, K.H., (2010). *Role of oxidative stress in various stages of psoriasis*. Indian journal of clinical biochemistry, 25(4), pp.388-392.

- Karababa, F., Yesilova, Y., Turan, E., Selek, S., Altun, H. and Selek, S., (2013). *Impact of depressive symptoms on oxidative stress in patients with psoriasis*. Redox Report, 18(2), pp.51-55.
- katiyar .s, saify .k, singh .s.k, rai . (2015) *oxidative stress verses skin hyperpigmentation*. International journal of current innovation research, vol. 1, issue 1, pp 9-12
- Kayar, A., Dokuzeylul, B., Kandemir, F.M., Kirbas, A., Bayrakal, A. and Or, M.E., (2015). *Total oxidant and antioxidant capacities, nitric oxide and malondialdehyde levels in cats seropositive for the feline coronavirus*. Veterinarni Medicina, 60(5), pp.274-281.
- Kiran, P.U. and Deedi, M.K. (2015). *Study of MDA and SGPT levels in patients with psoriasis*.
- Klaunig, J.E., Kamendulis, L.M. and Hocevar, B.A., (2010). *Oxidative stress and oxidative damage in carcinogenesis*. Toxicologic pathology, 38(1), pp.96-109.
- Kruk, J. and Duchnik, E., (2014). *Oxidative stress and skin diseases: possible role of physical activity*. Asian Pac. J. Cancer Prev, 15, pp.561-568.
- Koracevic D., Koracevic G., Djordjevic V., Andrejevic S. And Cosic V. (2001). *Method for the measurement of antioxidant activity in human fluids*. J. Clin Pathol, 54, 356-361.
- Kose, O., Canakci, V., Canakci, C.F., Yildirim, A., Kermen, E., Arabaci, T. and Gungor, A., (2014). *The effect of obesity on total antioxidant/oxidant status and oxidative stress index in patients with chronic periodontitis*. Oxidants and Antioxidants in Medical Science, 3(2), pp.153-159.
- Gurupadappa, J.R.G. and KS K., (2011). *Psoriasis: An oxidative stress condition*.
- Kuchekar, A.B., Pujari, R.R., Kuchekar, S.B., Dhole, S.N. and Mule, P.M., (2011). *international journal of pharmacy & life sciences*. Int. J. of Pharm. & Life Sci.(IJPLS), 2(6), pp.857-877.
- Kurahashi, T. and Fujii, J., (2015). *Roles of antioxidative enzymes in wound healing*. Journal of Developmental Biology, 3(2), pp.57-70.
- Lenaz, G., (2012). *Mitochondria and reactive oxygen species. Which role in physiology and pathology?*. In Advances in mitochondrial medicine (pp. 93-136). Springer Netherlands.
- Lowes, M.A., Suárez-Fariñas, M. and Krueger, J.G., (2014). *Immunology of psoriasis*. Annual review of immunology, 32, p.227.

- Mailloux, R.J. and Harper, M.E., (2012). *Mitochondrial proticity and ROS signaling: lessons from the uncoupling proteins*. Trends in Endocrinology & Metabolism, 23(9), pp.451-458.
- Mantovani, A., Gisondi, P., Lonardo, A. and Targher, G., (2016). *Relationship between Non-Alcoholic Fatty Liver Disease and Psoriasis: A Novel Hepato-Dermal Axis?*. International journal of molecular sciences, 17(2), p.217.
- Masaki, H., (2010). *Role of antioxidants in the skin: anti-aging effects*. Journal of dermatological science, 58(2), pp.85-90.
- Menter, A., Korman, N.J., Elmets, C.A., Feldman, S.R., Gelfand, J.M., Gordon, K.B., Gottlieb, A., Koo, J.Y., Lebwohl, M., Leonardi, C.L. and Lim, H.W., (2011). *Guidelines of care for the management of psoriasis and psoriatic arthritis: section 6. Guidelines of care for the treatment of psoriasis and psoriatic arthritis: case-based presentations and evidence-based conclusions*. Journal of the American Academy of Dermatology, 65(1), pp.137-174.
- Mohammed .I. hamzah (2014) *Oxidative stress markers and Proinflammatory Cytokine in Iraqi patients with Psoriasis vulgaris*
- Mrowietz, U., Kragballe, K., Reich, K., Spuls, P., Griffiths, C.E.M., Nast, A., Franke, J., Antoniou, C., Arenberger, P., Balieva, F. and Bylaite, M., (2011). *Definition of treatment goals for moderate to severe psoriasis: a European consensus*. Archives of dermatological research, 303(1), pp.1-10.
- Nagamani, M., Prahalladu, P. and Vijayababu, P.V.S.S.,(2015) *Lipid Peroxidation Product As A Marker Of Oxidative Stress In Psoriasis-A Case Control Study In North Coastal Andhra Pradesh*.
- Nassiri, S., Malekzad, F., Sarlak, M., Saeedi, M., Hedayati, M.A.H.D.I. and Qaisari, M., (2009). *Interplay among antioxidants and oxidants in psoriasis*. Iranian Journal of Dermatology, 12(2), pp.56-59.
- Ng, K.W. and Lau, W.M., (2015) *Skin deep: the basics of human skin structure and drug penetration*. In Percutaneous Penetration Enhancers Chemical Methods in Penetration Enhancement (pp. 3-11). Springer Berlin Heidelberg.
- Ni, C. and Chiu, M.W., (2014) *Psoriasis and comorbidities: links and risks*. Clinical, cosmetic and investigational dermatology, 7, p.119.

- Ojeh, N., Pastar, I., Tomic-Canic, M. and Stojadinovic, O., (2015). *Stem Cells in Skin Regeneration, Wound Healing, and Their Clinical Applications*. International journal of molecular sciences, 16(10), pp.25476-25501.
- Oliveira, M.D.F.S.P., Rocha, B.D.O. and Duarte, G.V., (2015). *Psoriasis: classical and emerging comorbidities*. Anais brasileiros de dermatologia, 90(1), pp.9-20.
- Oyinloye, B.E., Adenowo, A.F. and Kappo, A.P., (2015). *Reactive oxygen species, apoptosis, antimicrobial peptides and human inflammatory diseases*. Pharmaceuticals, 8(2), pp.151-175.
- Pai, V.V., Shukla, P. and Kikkeri, N.N., (2014) *Antioxidants in dermatology*. Indian dermatology online journal, 5(2), p.210.
- Pandel, R., Poljšak, B., Godic, A. and Dahmane, R., (2013). *Skin photoaging and the role of antioxidants in its prevention*. ISRN dermatology
- Pandey, M.K., Mitra, P. and Maheshwari, P.K., (2012). *The lipid peroxidation product as a marker of oxidative stress in epilepsy*. J. Clin. Diagn. Res, 6(4), pp.590-2.
- Pankotai, E., László Virág, M.D. and István Szokodi, M.D., (2010). *The role of mitochondria in reactive nitrogen species production and in restoring energy levels of oxidatively injured cells*.
- Peslyak, M., (2011). *Model of pathogenesis of psoriasis. Part 1. Systemic psoriatic process*. Mikhail Peslyak.
- Poljšak, B. and Dahmane, R., (2012). *Free radicals and extrinsic skin aging*. Dermatology research and practice, 2012.
- Poljsak, B., Dahmane, R. and Godic, A., (2013). *Skin and antioxidants*. Journal of Cosmetic and Laser Therapy, 15(2), pp.107-113.
- Poljsak, B., Pesti, M., Jamnik, P. and Raspor, P., (2011). *Impact of environmental pollutants on oxidation-reduction processes in the cell environment*. Encyclopedia of Environmental Health. Elsevier.
- Prasanth, D., Masram, P., Paul, R. and Anup, T., (2014) *a systemic analysis on psoriasis WSR TO EKAKUSHTA*.
- Priya, R., Kumar, U., Saran, A., Kumari, R. and Kishore, C., (2014). *Oxidative stress in psoriasis*. Biomedical Research (0970-938X), 25(1).

- Pujari, V.M., Ireddy, S., Itagi, I. and Kumar, S., (2014). *The Serum Levels of Malondialdehyde, Vitamin E and Erythrocyte Catalase Activity in Psoriasis Patients*. Journal of clinical and diagnostic research: JCDR, 8(11), p.CC14.
- Pujari, V.M., Suryakar, A.N. and Ireddy, S., (2010). *Oxidants and antioxidant status in psoriasis patients*. Biomed Res, 21(2), pp.221-3.
- Rachakonda, T.D., Schupp, C.W. and Armstrong, A.W., (2014). *Psoriasis prevalence among adults in the United States*. Journal of the American Academy of Dermatology, 70(3), pp.512-516.
- Rahman, T., Hosen, I., Islam, M.T. and Shekhar, H.U., (2012). *Oxidative stress and human health*. Advances in Bioscience and Biotechnology, 3(7A), p.997.
- Rastogi, A. and Pospíšil, P., (2011). *Spontaneous ultraweak photon emission imaging of oxidative metabolic processes in human skin: effect of molecular oxygen and antioxidant defense system*. Journal of biomedical optics, 16(9), pp.096005-096005.
- Reshma, S., Vasudha, K.C. and Prasad, A.S., (2011). *High-sensitivity C-reactive protein, lipid profile, malondialdehyde and total antioxidant capacity in psoriasis*. International Journal of Biological and Chemical Sciences, 5(4), pp.1394-1402.
- Reuter, S., Gupta, S.C., Chaturvedi, M.M. and Aggarwal, B.B., (2010). *Oxidative stress, inflammation, and cancer: how are they linked?*. Free Radical Biology and Medicine, 49(11), pp.1603-1616.
- Rinnerthaler, M., Bischof, J., Streubel, M.K., Trost, A. and Richter, K., (2015). *Oxidative stress in aging human skin*. Biomolecules, 5(2), pp.545-589.
- Roberson, E.D. and Bowcock, A.M., (2010). *Psoriasis genetics: breaking the barrier*. Trends in Genetics, 26(9), pp.415-423.
- Roberts, R.A., Smith, R.A., Safe, S., Szabo, C., Tjalkens, R.B. and Robertson, F.M., (2010). *Toxicological and pathophysiological roles of reactive oxygen and nitrogen species*. Toxicology, 276(2), pp.85-94.
- Rundhaug, J.E. and Fischer, S.M., (2010). *Molecular mechanisms of mouse skin tumor promotion*. Cancers, 2(2), pp.436-482.
- Shafaq, N., (2012). *An overview of oxidative stress and antioxidant defensive system*.



- Shamsi, M.B., Venkatesh, S., Kumar, R., Gupta, N.P., Malhotra, N., Singh, N., Mittal, S., Arora, S., Arya, D.S., Talwar, P. and Sharma, R.K., (2010). *Antioxidant levels in blood and seminal plasma and their impact on sperm parameters in infertile men*. Indian journal of biochemistry & biophysics, 47(1), p.38.
- Sicherer, S.H. and Leung, D.Y., (2014). *Advances in allergic skin disease, anaphylaxis, and hypersensitivity reactions to foods, drugs, and insects in 2013*. Journal of Allergy and Clinical Immunology, 133(2), pp.324-334.
- Şikar Aktürk, A., Özdoğan, H.K., Bayramgürler, D., Çekmen, M.B., Bilen, N. and Kıran, R., (2012). *Nitric oxide and malondialdehyde levels in plasma and tissue of psoriasis patients*. Journal of the European Academy of Dermatology and Venereology, 26(7), pp.833-837.
- Silveira, S., Pedroso, J.E. and Myaki Pedroso, D.M., (2014). *UV light and skin aging*. Reviews on environmental health, 29(3), pp.243-254.
- Singh, A.K., Pandita, S., Huozha, R., Kushwaha, R., Chandra, V.S.G. and Vaidya, M.M., (2011). *role of nitric oxide in immunity—A REVIEW*.
- Skroza, N., Proietti, I., Pampera, R., La Viola, G., Bernardini, N., Nicolucci, F., Tolino, E., Zuber, S., Soccodato, V. and Potenza, C., (2013). *Correlations between psoriasis and inflammatory bowel diseases*. BioMed research international.
- Son, Y., Cheong, Y.K., Kim, N.H., Chung, H.T., Kang, D.G. and Pae, H.O., (2011). *Mitogen-activated protein kinases and reactive oxygen species: how can ROS activate MAPK pathways?* Journal of signal transduction.
- Stojiljković, D., Pavlović, D. and Arsić, I., (2014) *Oxidative Stress, Skin Aging and Antioxidant Therapy/Oksidacioni Stres, Starenje Kože I Antioksidaciona Terapija*. Acta Facultatis Medicae Naissensis, 31(4), pp.207-217.
- Suen, L.J.W., Huang, H.M. and Lee, H.H., (2014). *A comparison of convenience sampling and purposive sampling*. Hu Li Za Zhi, 61(3), p.105.
- Sun, L. and Zhang, X., (2014) *The immunological and genetic aspects in psoriasis*. In Applied Informatics (Vol. 1, No. 1, pp. 1-21). Springer Berlin Heidelberg.
- Sürücü, H.A., Aksoy, N., Özgöztas, O., Sezen, H., Yesilova, Y. and Turan, E., (2015). *Prolidase activity in chronic plaque psoriasis patients*. Advances in Dermatology and Allergology/Postępy Dermatologii i Alergologii, 32(2), p.82.
- Terlecky, S.R., Terlecky, L.J. and Giordano, C.R., (2012). *Peroxisomes, oxidative stress, and inflammation*. World J Biol Chem, 3(5), pp.93-97.

- Venza, M., Visalli, M., Beninati, C., De Gaetano, G.V., Teti, D. and Venza, I., (2015). *Cellular mechanisms of oxidative stress and action in melanoma*. Oxidative medicine and cellular longevity, 2015.
- Wagener, F.A., Carels, C.E. and Lundvig, D., (2013). *Targeting the redox balance in inflammatory skin conditions*. International journal of molecular sciences, 14(5), pp.9126-9167.
- Weigle, N., (2013) *Risk Factors and Etiology*.
- Wohn, C., (2015). *Mechanisms of Psoriatic Plaque Formation in Mice*.
- Yadav, U. and Ramana, K.V., (2013). *Regulation of NF-B-induced inflammatory signaling by lipid peroxidation-derived aldehydes*. Oxidative medicine and cellular longevity.
- Zhou, Q., Mrowietz, U. and Rostami-Yazdi, M., (2009). *Oxidative stress in the pathogenesis of psoriasis*. Free Radical Biology and Medicine, 47(7), pp.891-905.

## Appendix I: Questionnaire

Thank you for your willingness to participate; your cooperation is very important to the success of the study. This is a questionnaire you are asked to fill out. Please answer the questions as frankly and accurately as possible. All information obtained in the study will be kept confidential.

1. Personal identification: a. Full name of the subject : \_\_\_\_\_  
b. Subject code number: \_\_\_\_\_
2. Demographic detail
  - a. Age : \_\_\_\_\_
  - b. Gender : Male  Female
  - c. Region : \_\_\_\_\_ Residence area \_\_\_\_\_
  - d. Height : \_\_\_\_\_
  - e. Weight : \_\_\_\_\_
  - f. Marital status: Single  Married  Widowed
  - g. Education level: Illiterate  High School or less  College or above
  - h. Occupation : \_\_\_\_\_
3. Group: Study  Control
4. Date of diagnosis : \_\_\_\_\_
5. Age of onset : \_\_\_\_\_
6. Disease duration : \_\_\_\_\_
7. **Dermatology History**

	Yes	No
a. Prior skin cancers	<input type="checkbox"/>	<input type="checkbox"/>
b. Has an immediate family member had skin cancer or skin problems?		
c. Trauma	<input type="checkbox"/>	<input type="checkbox"/>
d. Operation wound	<input type="checkbox"/>	<input type="checkbox"/>
e. Vaccination	<input type="checkbox"/>	<input type="checkbox"/>
f. Insect or animal bite	<input type="checkbox"/>	<input type="checkbox"/>
g. Sun burn	<input type="checkbox"/>	<input type="checkbox"/>
h. Obesity	<input type="checkbox"/>	<input type="checkbox"/>
i. Stress	<input type="checkbox"/>	<input type="checkbox"/>
j. Seasonal variation	<input type="checkbox"/>	<input type="checkbox"/>
k. Are you currently pregnant or breast feeding?	<input type="checkbox"/>	<input type="checkbox"/>
l. Any other health problems _____		

<b>8. Treatment history -Medications and Allergies</b>	<b>yes</b>	<b>No</b>
a. Tropical	<input type="checkbox"/>	<input type="checkbox"/>
b. Phototherapy	<input type="checkbox"/>	<input type="checkbox"/>
c. systemic	<input type="checkbox"/>	<input type="checkbox"/>
d. None	<input type="checkbox"/>	
e. Other : _____		

**9. Social history**

a. Do you smoke?	<input type="checkbox"/>	<input type="checkbox"/>
b. How often do you drink alcohol?		
None	<input type="checkbox"/>	1-3
	<input type="checkbox"/>	4-6
	<input type="checkbox"/>	11-15
	<input type="checkbox"/>	21+
	<input type="checkbox"/>	
c. What is your occupation or former occupation? _____		
d. Number of people living in household? _____		

**10. Medical History**

Do you have the following medical conditions?	Yes	No
1. Artificial joints	<input type="checkbox"/>	<input type="checkbox"/>
2. Cancer	<input type="checkbox"/>	<input type="checkbox"/>
3. Asthma	<input type="checkbox"/>	<input type="checkbox"/>
4. Depression psychiatric diseases/ disorder	<input type="checkbox"/>	<input type="checkbox"/>
5. Stomach ulcers	<input type="checkbox"/>	<input type="checkbox"/>
6. Tuberculosis	<input type="checkbox"/>	<input type="checkbox"/>
7. hay fever	<input type="checkbox"/>	<input type="checkbox"/>
8. Bleeding disorders	<input type="checkbox"/>	<input type="checkbox"/>
Other _____		

**For physician or clinician use only**

11. General physical examinations

a. Severity of psoriasis and percent of body surface affected

- Mild up to 3%: \_\_\_\_\_
- Moderate 3%-10%: \_\_\_\_\_
- Severe > 10%: \_\_\_\_\_

b. Types of lesion

- Papules : \_\_\_\_\_
- Pustule : \_\_\_\_\_
- Plaque : \_\_\_\_\_
- Erythema : \_\_\_\_\_

## **Appendix II: Consent form**

### **PART I: patient information**

You are invited to take part in a research study. Before you decide, it is important for you to understand why the research is being done and what it will involve. Please take a time to read the following information carefully and discuss it with others if you wish.

#### **General description of the disease: psoriasis**

Psoriasis is a long lasting disease characterized by patches of abnormal skin. These skin patches are typically red, itchy, and scaly. They may vary in severity from small and localized to complete body coverage. Psoriasis is generally considered a genetic disease which is triggered by environmental factors. Psoriasis is associated with an increased risk of psoriatic arthritis, lymphomas, cardiovascular disease, Crohn's disease and depression. Psoriasis is not contagious. There is no cure for psoriasis. However, various treatments can help control the symptoms.

#### **Purpose of the study**

The aim of the study is to evaluate levels of some chemical components among patients of psoriasis at Alert Hospital, Addis Ababa, Ethiopia. This will be important in the monitoring of the risk of developing of cardiovascular diseases.

#### **Procedure**

If you are volunteers to participate in this study, we will ask you the following

- you will be asked to come the hospital after fasting overnight and compliance regarding fasting will be checked by on the Moring of the examination
- you are required to reply to the questions raised by physician or nurse
- you need to allow withdrawal of a 5ml of a serum blood from your hand

**Potential risk**

The only risk is that may result is mild pain associated with the withdrawal procedure. This is just the result of the routine work which is experienced by all patients. There is nothing else which may make the patient feel discomfort.

**Benefits**

This study is designed to determine the correlation between psoriasis and oxidative stress, total antioxidant capacity and levels of lipid profile which have a strong effect on developing cardiovascular diseases. Thus the patients involved in the study will have got information about their status and they may take measures to control the disease, before it is going worse. This will be done without any payment.

**Confidentially**

Any information that is obtained from the result and that can be identified with you will be remained confidential and will be disclosed only with you permission. We will not use your name in any of the information we get from the study or in any of the research report.

**Sharing the result**

At the end of the study, the study finding will be published and it will be disseminated to all responsible bodies for taking actions and other purpose.

**The right to refuse**

Please know that your participation in this study is entirely voluntary and you are free withdrawn at any time from the study. Taking of your blood sample to be used for this research is completely voluntary. Your decision will not affect your right to take medication or any other health service facility now or in future

### **Appendix III -Informed consent**

I volunteer to participate in a research project conducted Asrat Endrias from Addis Ababa University.

I have been informed about a study that plans to measure the status of oxidative stress in psoriasis. For this purpose, blood needs to be taken. The aims of the study and the possible risks, include mild pain during blood collection were explained to me.

I am also informed that all the information contained within the questionnaire is to be kept confidential. Moreover, I have also been well informed of my right to kept hold of information, decline to cooperate and make myself withdraw from the study.

It's therefore, with full understanding of the situation that I gave the informed consent voluntarily to the researcher to use the blood taken for the investigation. Moreover, I have had the opportunity to ask questions about it and received clarification to my satisfaction. I have also been informed that the nature of the questionnaire is private.

I have been given a copy of this consent form and I have read and understood the explanation provided to me. I have had all my questions answered to my satisfaction, and I voluntarily agree to participate in this study.

Name of participant \_\_\_\_\_ Signature \_\_\_\_\_ Date \_\_\_\_\_

Name of investigator \_\_\_\_\_ Signature \_\_\_\_\_ Date \_\_\_\_\_

For participants who cannot read and write

- Witness name \_\_\_\_\_
- Signature \_\_\_\_\_
- Date \_\_\_\_\_



## Appendix IV-Amharic Version

አዲስ አበባ ዩ.ንቨርሲቲ፡ የሕክምና ፋካልቲ፡ ባዮኬሚስትሪ ት/ክፍል

**የጥናት ተሳታፊዎች የመረጃና የስምምነት ቅፅ**

**መግቢያ፡**

በዚህ ጥናታዊ ዕውቅና እንዲሳተፉ እየተጋበዙ ነው። በጥናቱ ላይ ለመሳተፍ ከመወሰንን በፊት ጥናቱ ለምን እንደሚካሄድና ምንም ዓይነት ነገሮች እንደሚያጋጥሙት ማወቅ ነው። ስለዚህ ጥቂት ጊዜ ይወስዱና የሚከተለውን ስለጥናቱ በተመለከተ መረጃ ይመልከቱ አስፈላጊ ከሆነም ከሌሎች ሰዎች ጋር ይወያዩ። ማንኛውም ግልጽ ያልሆነ ነገር ካለ ወይም ተጨማሪ መረጃ ከፈለጉ የጥናቱ ባለቤት መጠየቅ ይችላሉ።

**የጥናቱ ዋና ተመራማሪ ፡ አስራት አንድርያስ** ኢ-ሜይል - [endasrias@gmail.com](mailto:endasrias@gmail.com)

አድራሻ፡- ሞባይል +251 924437711

**የበአለርት/ አህሪሆስፒታል የምርምር ድጋፍ ሰጭ ኮሚቴ ስልክ ቁጥር፡** 0113-48-12-89

### ሶርያሊስ

ሶርያሊስ የማይደን በዙር የሚተላለፍ ለረጅም ጊዜ አብሮ የሚኖር መድሃኒት ያልተገኘለት የቆዳ በሽታ ነው። በሽታው ከትንሻ የሰውነት ክፍል እስከ ሙሉ የሰውነት አካል የሚሸፍን እና ቆዳን የማሳከክ እንዲሁም የመቆጣት ባህሪ ያለው ሲሆን በተለያዩ መድሃኒቶች በሽታው ወደ ሌላ የሰውነት ክፍል እንዳይሰፋፋ መከላከል ይቻላል።

በሽታውምከ ልብ ህመም ለየመገጣጠሚያ ችግሮች ለስካር በሽታ ለአንጀት መቆጣት ለአእምሮ ጭንቀት ያጋልጣል። ስለዚህም አሁን የምናደርገው ጥናት ከላይ የተጠቀሱትን በሶርያሊስ በሽታ ምክንያት ሊከሰቱ የሚችሉትን ችግሮች ለመቆጣጠር ይረዳል ብለን እናስባለን።

### የጥናቱ አላማ፡

የዚህ ጥናት ዋና አላማ በአለርት ሆስፒታል ውስጥ በተለያዩ መረጃ ለሚገኙ ሶርያሊስ የቆዳ ሁሙማን በህመሙ ምክንያት ሊመጣሉ ተብለው የሚታሰቡ ተያያዥ ችግሮች ምን እንደሚመስል ጥናት ማድረግ ነው።

### የተሳታፊዎች ሁኔታ፡-

በጥናቱ ሊዳተፉ ከተስማሙ

1. ሀኪሙ ለሚጠይቁት ጥያቄዎች መልስ መስጠት ለምሳሌ፡- ዕድሜ፣ በሽታው መች አንደተያዙ/ ከዚህ በፊት ምን ዓይነት መድሃኒቶች እንደደተተቀሙ.....
2. ለምርመራው በመርፌ ከአጅ ላይ ትንሽ ናሙና 5ml የደምመጠን፣ለወሲድ መስማማት፡፡

**ሊከሰቱ ስለሚችሉ ስጋቶች እና የምችት መጋደሎች**

ለጥናቱ በሚወሰደው ደም ምክንያት የተለየ ችግር አይከሰትም። ምክንያቱም የጥናቱና ሕመሙና አወሳሰድ ከወትሮው በሽተኛው ለራሱ ብሎ ከሚሰጠው የተለየ አይደለም። እንዲያውም ለበሽተኛው ያለበትን ሁኔታ እንዲያውቅ እና እርምጃ እንዲወስድ ያደርገዋል። ከዚህ ጥናት ጋር በተያያዘ በጤናም ሆነ በሚያገኙት ተገቢ ህክምና ምንም አይነት ጉዳት ስለማያስከትል አይስጉ። ሊከሰት የሚችለው ችግር በደም አወሳሰድ ግዜ ከመርፌ መወጋት የተያያዘ ህመም ብቻ ነው። ይህ ደግሞ ከተለመደው የተለየ አይደለም።

**ጥቅሞች**

የምርመራው ውጤት ለህሞማኖቹ እንዲሁም አጠቃላይ ለሶርያሲስ ታማሚዎች በተደጋጋሚ ላሚያጋጥማቸው ከበሽታው እና ጋር ለተያያዙ እና ሌሎችም ችግሮች ከመከሰታቸው በፊት በቅድሚያ እርምጃ እንዲወስዱ እንዲሁም ጥንቃቄ እንዲያደርጉ መረጃ ይሰጣል። ሆስፒታሉም በዚህ ምክንያት የሚመጣውን ወጪ እንዲሁም የሰው ሃይል እና ግዜን ይቆጥባል።

**ምስጢር ስለመጠበቅ**

ለሁሉም ጥያቄዎች የሰጡንን መልሶች የሚስጥረር እንደምንይዝ ሊያውቁ ይገባል። የተወሰደው ናሙና ለጥናት ብቻ የሚውል ነው። ስለሚወሰደው ማንኛውም መረጃዎች ሆነ የጥናት ውጤት ለማሰራጨት በስምሳይ ሆነ በሚስጥር (ኮድ) የሚመዘገብ ይሆናል።

**የጥናቱ ያለመሳተፍ ወይም ራስም የማግለል መብት**

በዚህ ጥናት መሳተፍ ሙሉ በሙሉ በርሶፍ ቃድ የተመሰረተ መሆኑን ልናሳስብ እንወዳለን። በጥናቱ ውስጥ ማሳተፍ አለመፈለግዎ በጤና እንካብካቤዎ ላይ ምንም ተፅዕኖ አይኖረውም ። በመሆኑም በማንኛውም ግዜ ምንም ዓይነት ምክንያት ሳይሰጡ ከጥናቱ ራስን የማግለል መብት የተጠበቀ ነው። የህንን ከወሰኑም ማንም ምክንያቱን እንዲገልፁ ሊያስገድደዎት አይችልም። የሰጡት ደም ለዚህ ጥናት እንደሚውል ማድረግ በእርሶሙ ሉፈቃድ ብቻ ሲሆን በጥናቱ ላለመሳተፍ መወሰን ወይም አለመወሰን መድሐኒት ወይም ሌላ የጤና አገልግሎት የማግኘት መብት አሁንም ሆነ ለወፊቱ ምንም አይነት ተፅዕኖ አያሳድርብትም።

**መረጃ ስለማግኘት**

ይህ ጥናት በአዲስ አበባ የኒቨርስቲ ሜዲካል ፋኩልቲ የባዮኬሚስትሪ ትምህርት ክፍል እና በአለርት ሆስፒታል የምርምር ድጋፍ ሰጭ ኮሚቴ ድጋፍ አግኝቷል። ማንኛውም አይነት ጥያቄዎች ካለዎት እርግጠኛ ሆነው ለጥናት ለማሳተፍ እስኪወስኑ ድረስ ተገቢ የሆነውን ማንኛውንም መረጃ የመጠየቅ መብት አለዎት።

**ክፍልሁለት:-የስምምነትቅፅ**

የዚህጥናት መሰረታዊ ዓላማ እና ሌሎች መረጃን በሚገባ ተገንዚቤለሁ።ተሳትፎዬ በፍቃደኝነት ላይ ብቻ የተመለከዘ እደሆነም ተረድቻለሁ። ማንኛውም ሰብዓዊም ሆነ ህጋዊ መብት ሳይነካ ከጥናቱ ራሴን ማግለል እደምችልም እንደሆነ። ስለጥናቱ ዝርዝር ጉድይ በግልፅ ከተረዳሁት ባሻገር በተጨማሪ ማብራሪያ ብፊልግ መጠየቅ እደምችል ተረድቻለሁ። በመሆኑም በፈቃዴ የዚህ ጥናት አካል እንድሆን ስፈልግ የምጠብቅብኝ ሁሉ ለምድረግ በመወሰን መሆኑን በፈርማዬ አረጋግጣለሁ።

የተሳታፊ ስም : \_\_\_\_\_

- ቀን: \_\_\_\_\_
- ፊርማ: \_\_\_\_\_

የጥናቱ ዋና ተመራመሪ ስም \_\_\_\_\_

- ቀን \_\_\_\_\_
- ፊርማ \_\_\_\_\_

ማንበብ እና መፃፍ ለማይችሉ የጥናቱ ተሳታፊዎች

- የምስክር ስም \_\_\_\_\_
- ፊርማ \_\_\_\_\_
- ቀን \_\_\_\_\_

**መጠይቅ**

**አዲስ አበባ ዩንቨርሲቲ- የሕክምና ፋካልቲ -ባዮኬሚስትሪ ት/ክፍል**

ይህን መጠይቅ ለመሙላት ፍቃደኛ በመሆንዎ ከልብ እናመሰግናለን :: የእርስዎ ትብብር ለምርምሩ ስኬት አስፈላጊ እንደመሆኑ መጠን ይህን መጠይቅ በተቻለ መጠን በግልፅ እና በትክክል እንዲሞሉት በአክብሮት እጠይቃለሁ:: ማንኛውም እርስዎን የተመለከተ መረጃ ሚስጥራዊነቱ የተጠበቀ ነው::

**የማንነት መለያ**

1. የሚስጢር ቁጥር: \_\_\_\_\_ የካርድ ቁጥር: \_\_\_\_\_

**ማህበራዊ ና ስነ ሀዘብ መራጃ**

1. ዕድሜ: \_\_\_\_\_
2. ፆታ: \_\_\_\_\_ ወንድ  ሴት
3. ክልል አድራሻ ከተማ: \_\_\_\_\_
4. ቁመት: \_\_\_\_\_ ከብደት: \_\_\_\_\_
5. የጋብቻ ሁኔታ: ያገባ  ላጤ  የፈታ
6. የትምህርት ደረጃ: አንደኛ ደረጃ  ሁለተኛ ደረጃ  ኮሌጅ
7. የስራ ዓይነት: \_\_\_\_\_
8. ምድብ: \_\_\_\_\_ 1. የምርምር  2. ቆጥጥር
9. የታከሙበት ቀን: \_\_\_\_\_
10. በሽው የታየበት ዕድሜ: \_\_\_\_\_
11. በሽታው የቆየበት ጊዜ: \_\_\_\_\_

**የቆዳ ህመም መንገዶች**

አዎ አይ

1. የቆዳ ካንሰር	<input type="checkbox"/>	<input type="checkbox"/>
2. አደጋ	<input type="checkbox"/>	<input type="checkbox"/>
3. በተመሳሳይ ህመም የተጠቁ የቤተሰብ አባል	<input type="checkbox"/>	<input type="checkbox"/>
4. የቆዶ ጥገና ቁስል	<input type="checkbox"/>	<input type="checkbox"/>
5. ክትባት	<input type="checkbox"/>	<input type="checkbox"/>
6. የተባይ እና የእንስሳት ንክሻ	<input type="checkbox"/>	<input type="checkbox"/>
7. የወቅትመቀያየር	<input type="checkbox"/>	<input type="checkbox"/>
8. እርግዝና ወይም ጡት ማጥትባት	<input type="checkbox"/>	<input type="checkbox"/>
9. መድሃኒት	<input type="checkbox"/>	<input type="checkbox"/>
10. ከፍተኛ የፀሃይ ብርሃን	<input type="checkbox"/>	<input type="checkbox"/>
11. ከልክ ያለፈ ውፍረት	<input type="checkbox"/>	<input type="checkbox"/>
12. ጭንቀት	<input type="checkbox"/>	<input type="checkbox"/>

ሌላ \_\_\_\_\_

**ከዚህ በፊት የወሰዱት ወይም የተጠቀሙት የህክምና አይነት**

- |                              |                      |                      |
|------------------------------|----------------------|----------------------|
| 1. በቆዳ የሚቀቡ የቅባት /ክሬም መድሃኒቶች | <input type="text"/> | <input type="text"/> |
| 2. የብርሃን ህክምና(ፎቶ ቴራቢ)        | <input type="text"/> | <input type="text"/> |
| 3. መዲሃኒቶች(የእንክብል፣ ሽሮፕ/ፈሳሽ)   | <input type="text"/> | <input type="text"/> |
| (ለየተኛው አይነት በሽታ ጥቀስ ) _____  |                      |                      |
| 4. የለም                       | _____                |                      |
| 5. ሌላ ካለ ጥቀስ                 | _____                |                      |

**ማህበራዊ-ኒታዎ**

አዎ

አይ

- |                                 |                      |                      |
|---------------------------------|----------------------|----------------------|
| 1. የማጨስ ልምድ አልዎት                | <input type="text"/> | <input type="text"/> |
| 2. የመጠጥ ልምድ አልዎት                | <input type="text"/> | <input type="text"/> |
| 3. የስራዎ አይነት ምንድን ነው ወይም ምን ነበር | _____                |                      |
| 4. የቤተሰብ ብዛት ቁጥር                | _____                |                      |

**ከዚ በታች ከተዘረዘሩት የጤና ችግሮች ያለባችን ምልክት ያርጉ።**

አዎ

አይ

- |                        |                      |                      |
|------------------------|----------------------|----------------------|
| 1. ሰው ሰራሽ አካል          | <input type="text"/> | <input type="text"/> |
| 2. ካንሰር                | <input type="text"/> | <input type="text"/> |
| 3. አዝማ                 | <input type="text"/> | <input type="text"/> |
| 4. የ አእምሮ ጭንቀት/ችግር     | <input type="text"/> | <input type="text"/> |
| 5. የጨጓራ ቁስትል           | <input type="text"/> | <input type="text"/> |
| 6. ቲቢ                  | <input type="text"/> | <input type="text"/> |
| 7. የብናኝ አለርጂ/hay fever | <input type="text"/> | <input type="text"/> |
| 8. የደም መፍሰስ ችግር        | <input type="text"/> | <input type="text"/> |

ሌላ \_\_\_\_\_

# Appendix VII –PASI

## PSORIASIS AREA AND SEVERITY INDEX (PASI) WORKSHEET

HOSPITAL NO.: .....

PATIENT NAME: .....

DATE OF VISIT: .....

The Psoriasis Area and Severity Index (PASI) is a quantitative rating score for measuring the severity of psoriatic lesions based on area coverage and plaque appearance.

Plaque characteristic	Lesion score	Head	Upper Limbs	Trunk	Lower Limbs
<b>Erythema</b>	0 = None				
<b>Induration/Thickness</b>	1 = Slight				
	2 = Moderate				
<b>Scaling</b>	3 = Severe				
	4 = Very severe				
Add together each of the 3 scores for each body region to give 4 separate sums (A).					
<b>Lesion Score Sum (A)</b>					

Percentage area affected	Area score	Head	Upper Limbs	Trunk	Lower Limbs
<b>Area Score (B)</b> <i>Degree of involvement as a percentage for each body region affected (score each region with score between 0-6)</i>	0 = 0%				
	1 = 1% - 9%				
	2 = 10% - 29%				
	3 = 30% - 49%				
	4 = 50% - 69%				
	5 = 70% - 89%				
	6 = 90% - 100%				
Multiply Lesion Score Sum (A) by Area Score (B), for each body region, to give 4 individual subtotals (C).					
<b>Subtotals (C)</b>					
Multiply each of the Subtotals (C) by amount of body surface area represented by that region, i.e. x 0.1 for head, x 0.2 for upper body, x 0.3 for trunk, and x 0.4 for lower limbs.					
<b>Body Surface Area</b>		x 0.1	x 0.2	x 0.3	x 0.4
<b>Totals (D)</b>					
Add together each of the scores for each body region to give the final PASI Score.					

PASI Score =