



**ECOLOGICAL, FLORISTIC AND ETHNOBOTANICAL STUDIES IN AND AROUND
WEJIG-MAHGO-WAREN MASSIF FOREST PATCHES IN SOUTHERN TIGRAY,
ETHIOPIA**

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Addis Ababa University

Addis Ababa, Ethiopia

May 2019



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Mebrahtu Hishe Gidey

A Dissertation Submitted to

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(Biology: Botanical Science)

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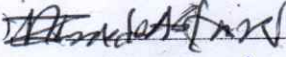
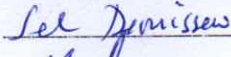

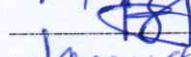
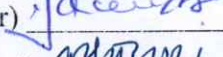
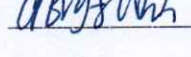
Ecological, Floristic and Ethnobotanical studies
in and around Wejig-Mahgo-Waren Massif
Forest Patches in Southern Tigray, Ethiopia

By

Mebrahtu Hishe Gidey

A Thesis Presented to the Graduate Programmes of the Addis Ababa University in Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy in Plant Biology & Biodiversity Management

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Abstract

Ecological, Floristic and Ethnobotanical Studies in and around Wejig-Mahgo-Waren Massif Forest Patches in Southern Tigray, Ethiopia

Mebrahtu Hishe Gidey, PhD Dissertation

Addis Ababa University, 2019

*There is a declining trend of biodiversity in general and plant diversity in particular. This trend is leading to the loss of the associated indigenous and local botanical knowledge in Ethiopia, which is more severe in the northern highlands than elsewhere in the country. The purpose of this research was to study vegetation composition, soil seed bank and to document associated ethnobotanical and management practices applied by the forest fringe communities in and around Wejig-Mahgo-Waren Massif Forest area. Vegetation data were collected from a sample of 150 quadrats (each 20 m x 20 m) placed in ten transect lines, which were systematically laid. All vascular plant species were recorded. Diameter at breast height was measured and seedlings and saplings were counted and recorded. Height and percentage cover abundance were estimated. Soil samples were collected from 75 quadrats down to 0-5 cm and 5 -10 cm depth. Stratified random sampling of 309 informants was performed for collection of data on indigenous knowledge, of social classes associated with the forest and the resources in it. Thirty key informants were purposively selected with the help of local administrators, elders and other community members for collection of ranking exercises. For collection of ethnobotanical data, semi-structured interview, guided field walks and focus group discussions were applied. Basal area, density, frequency, importance value index, Shannon-wiener diversity index, cluster analysis and ordination were computed on ecological data. Density, composition, depth distribution and Sorensen coefficient of similarity were computed for soil seed bank analysis. Ethnobotanical analytical tools, including preference ranking, informant consensus factor, fidelity level, direct matrix ranking and cultural significance index were employed to describe the reciprocal relationship between the forest vegetation and forest fringe communities. Vegetation study revealed a total of 264 plant species belonging to 162 genera and 82 families. Woody (45.45%) species were higher than herbs (42.04%). Asteraceae (27 species, 32.93%), Poaceae (24 species, 29.27%) and Fabaceae (23 species, 20.05%) were the dominant families in terms of number of species. Five plant communities were identified in the forest vegetation, namely *Cordia alliodora* – *Carissa spinarum*, *Olea europaea* subsp. *cuspidata* – *Juniperus procera*, *Dodonaea angustifolia* – *Acacia abyssinica*, *Erica arborea* – *Myrsine africana* and *Acacia etbaica* – *Acacia tortilis*. The highest (3.87) and the least (3.21) Shannon-Wiener diversity indices were found in communities two and five, respectively. Altitude, slope, livestock grazing and human impacts were the factors influencing species distribution. From soil samples, the total number of species recorded was 54, representing 42 genera and 23 families. The number of viable seeds in the soil samples corresponded to a seed bank density down to 10 cm was 1115 m⁻². Herbs (85.16%) and woody species (14.84%) were the plants counted from the germination assay. Soil Seed bank and standing vegetation shared 36 species and their similarity was 23 %. The informants viewed the forest as an important entity for attraction of rain and for wildlife habitation followed by beekeeping and foraging as well as for sourcing straw and fuelwood. Of 79 plant species differentiated as useful to the community, 52 (66%) were used in traditional medicine (human and livestock), 28 (35%) as livestock fodder/ forage and 27 (34%) were recognized as honeybee forage. The highest informant consensus factor values were calculated for dermatological ailments (0.98,) followed by external injuries, bleeding and snakebites (0.92). The highest fidelity level (96.15%) was recorded for *Verbascum sinaiticum* followed by *Withania somnifera* (89.47%). Deforestation, agricultural expansion, fuelwood collection, grazing and settlement in decreasing order were considered responsible for forest resources depletion. Both the associated indigenous knowledge and the forest resources were under pressure. Thus, to restore the forest in the shortest possible time, restoration strategy that combines planting of seedlings of indigenous species and natural restoration techniques need to be applied.*

Keywords/phrases: Multiuse, Plant communities, Restoration, Tigray, Wejig-Mahgo-Waren Forest

DEDICATION

This dissertation is dedicated to the local people of the study area who maintained the forest and their indigenous botanical knowledge, even at the worst drought times.

“A nation that destroys its soils destroys itself. Forests are the lungs of our land, purifying the air and giving fresh strength to our people.”

(Franklin D. Roosevelt, 29 Jan 1935)

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List of acronyms and abbreviations

| | |
|------------|-----------------------------------|
| AAU | Addis Ababa University |
| AR | Arsi Floristic region |
| BA | Bale Floristic region |
| BA | Basal Area |
| CCA | Canonical Correspondence Analysis |
| CF | Correction Factor |
| CSI | Cultural Significance Index |
| CSA | Central Statistical Agency |

| | |
|----------------|--|
| DAF | Dry Afromontane Forest |
| DBH | Diameter at Breast Height |
| DCA | Detrended Correspondence Analysis |
| DCR-DNH | Department of Conservation and Recreation/Division of Natural Heritage |
| DF | Degree of Freedom |
| DMR | Direct Matrix Ranking |
| EFAP | Ethiopian Forestry Action Program |
| ETH | Ethiopia |
| EW | Eritrea West Floristic region |
| EWNHS | Ethiopian Wildlife and Natural History Society |
| EVI | Ethno-ecological importance value index |
| FAO | Food and Agriculture organization |
| FEE | Flora of Ethiopia and Eritrea |
| FL | Fidelity Level |
| GD | Gondar Floristic region |
| GG | Gamo-Gofa Floristic region |
| GIS | Geographic Information System |
| GJ | Gojjam Floristic region |
| GPS | Geographic Position System |
| HA | Harerge Floristic region |
| ha | hectare |
| ICF | Informant Consensus Factor |
| IL | Illubabor Floristic region |

| | |
|--------------------------|--|
| IVI | Important Value Index |
| IUCN | International Union for the Conservation of Nature |
| IUFRO | International Union of Forest Research Organizations |
| MEF | Ministry of Environment and Forest |
| KF | Kefa Floristic region |
| MEFCC | Ministry of Environment, Forest and Climate Change |
| m.a.s.l | Meter Above Sea Level |
| NGO | Non-governmental organization |
| NMSA | National Metrological Service Agency |
| RCI | Relative Cultural Importance |
| REDD⁺⁺ | Reduced Emission from Degradation and Deforestation |
| RDO | Relative Dominance |
| SD | Standard Deviation |
| SD | Sidamo Floristic region |
| SPSS | Statistical Package for Social sciences |
| SR | Similarity Ratio |
| SS | Sorensen Coefficient of Similarity |
| SU | Shewa Floristic region |
| TFAP | Tigray Forestry Action Program |
| TU | Tigray Floristic region |
| WCMC | World Conservation Monitoring Centre |
| WG | Welega Floristic region |
| WU | Welo Floristic region |

CHAPTER ONE

1. INTRODUCTION

1.1 Background

Ethiopia is located in the horn of Africa stretching from 3° N to 15° N latitude and from 33° E to 48° E longitude with an area coverage of 1,104,300 km². It is a country of great geographic diversity with wide altitudinal and physiographic variations. The altitude ranges from 125 meters below sea level in the Danakil Depression in Afar National Regional State to the highest peak of 4,553 meters above sea level (m.a.s.l.) on Mount Ras Dashen in Amhara National Regional State. The Great Rift Valley divides the Western and Southeastern highlands and the highlands on each side give way to vast semi-arid lowland areas in the East and West, especially in the southern part of the country (Friis *et al.*, 2010).

Biodiversity is understood as a key factor for the sustainability of life and its loss is one of the greatest environmental crises. The growing human population and the demand for natural resources have put great pressure on the biodiversity wealth of the world through deforestation, habitat fragmentation and overexploitation of species (Terborgh and van Schaik, 1997; Noss, 1999). Habitat loss and change, over-harvesting and climate change have been the direct causes of global biodiversity loss (Wood *et al.*, 2000), while population growth, changes in economic activities, socio-political factors, cultural factors and technological change are indirect drivers (Ministry of Agriculture, 2003). Beside these global factors, lack of technical knowledge and awareness and political instability has exacerbated the problem in many developing countries (Ayyad, 2003).

Ethiopia is one of the top 25 biodiversity-rich or megadiverse countries in the world where two of the world's 34 biodiversity hotspots (Brooks *et al.*, 2002), namely the Eastern Afromontane and the Horn of Africa hotspots (WCMC, 1994) trespass through its highland and lowland areas. It is also among the countries in the Horn of Africa regarded as the major center of diversity and endemism for several plants and animals. It is assumed that its diverse topography that gave rise to a wide range of altitude variations and other environmental gradients endowed the country with diversity of life. This has resulted in wide variations in rainfall, humidity and temperature because of which the country comprises ten ecosystems that range from Afro-alpine at the highest elevations to desert and semi-desert ecosystems at the lowest elevations. Present assessment of the flora of Ethiopia (Ensermu Kelbessa and Sebsebe Demissew, 2014) shows that the country is home to about 5,757 species of vascular plants. Nearly 10.58% (544 species) of which are known to be endemic to Ethiopia.

The definition of forest given in important studies of East-African vegetation (Greenway, 1973; White, 1983; Friis, 1992) has been adopted: "Forest is a continuous stand of woody individuals, at least 5 m in height, with crowns touching or intermingling." However, Ethiopia adopted a new forest definition as follows: "Land covering at least 0.5 hectare covered by (trees and bamboo), attaining a height of at least 2 meters and a canopy cover of at least 20%, or trees with the potential to reach these thresholds *in situ* in due course" (MEFCC, 2016). This forest definition differs from the definition used for international reporting to the Global Forest Resources Assessment (FAO, 2011) and from the forest definition used in the National Forest Inventory that applied the FAO forest definition with the thresholds of 10% canopy cover, a 0.5 ha area and a 5 m height. The reason for Ethiopia to change its national forest definition is to better capture

dry and lowland-moist vegetation resources. The reason for lowering the tree height from five to two meters is to capture *Combretum - Terminalia* dense woodlands found in Gambella and Benishangul Gumuz Regional States (MEFCC, 2016).

Currently, natural forests in Ethiopia mainly occur in the southwestern part of the country, while the forests that originally existed in central and northern Ethiopia have almost disappeared (EFAP, 1994; Feoli *et al.*, 2002; Melaku Bekele, 2003). Accelerated deforestation and habitat fragmentation that arises largely due to the conversion of forests to agricultural land-use types and the overutilization of forest resources to satisfy food and energy requirements of the increasing population are major environmental concerns in Ethiopia (Friis *and* Sebsebe Demissew, 2001; Gessesse Dessie and Kleman, 2007).

According to Zerihun Woldu *et al.* (2002) improving the management of the natural resources while providing ecological services and immediate economic needs are the major research and development challenges for the degraded areas of dry lands of East Africa in general and northern Ethiopia in particular. Due to fast population growth, overgrazing and deforestation for agricultural activities, fuelwood and construction material and the natural forest cover had decreased to a level of 2.5 percent in 1999 (Reusing and Kasberger, 2000). The combination of high endemism and fast habitat degradation in Ethiopia leads to a great risk of species extinction (Nyssen, 2001).

Tigray is one of the most environmentally degraded regions in Ethiopia, which has been left with relatively low remnant natural vegetation. According to pollen and charcoal studies in northern Ethiopia, forest disturbance has a 3000-year history (Darbyshire *et al.*, 2003) and soil erosion following vegetation clearance in Tigray occurred in the middle Holocene (Bard *et al.*, 2000).

Around 500 BC, *Podocarpus-Juniperus* forest that was common in the area was converted into a secondary vegetation of *Dodoniaea* scrub and grasslands that dominated northern Ethiopia for 1800 years while *Juniperus*, *Olea* and *Celtis* spread around AD 1400 to 1700 (Darbyshire *et al.*, 2003).

Ecological degradation, as evidenced by a declining forest area and several erosion phenomena, is most dramatic in the northern highlands of Tigray with the prevalence of recurrent drought spells and war events in the recent decennia (Janzen, 1988; Muys *et al.*, 2004). At present, the original vegetation is confined to around religious and worship sites where cutting trees and removal of plants are forbidden and in limited other isolated and protected areas (Feoli, 1996). In this region, remains of the original climax forest now only occur as sparse forest patches of just a few hectares, either surrounding churches, or in inaccessible places, like cliffs and mountain ridges (Kindeya Gebrehiwot, 1997). These small remnants of forest are called forest relics and are classified as dry Afromontane forest ecosystems. Mitiku Haile and Kindeya Gebrehiwot (2000) stated that the misuse of natural resources has resulted in very serious land degradation in most places. It can be said that environmental degradation, drought and socioeconomic instability are common in contemporary Tigray.

In 2003, the natural forest cover in Tigray was only 0.2 % of the total land mass of the region (Ministry of Agriculture, 2003), indicating the severity of forest degradation in the region. Currently, the western escarpment of the Great Rift Valley is the only site with an intact Afromontane forest cover in northern Ethiopia, such as Hugumburda, Dess'a and *Wejig-Mahgo-Waren* massif forests. *Wejig-Mahgo-Waren* massif forest faces a great danger from agricultural

expansion, settlement, overgrazing, incidence of tree falling, fuelwood and building material collection by the local community.

Local communities, government and NGOs are tackling the problem of soil and vegetation degradation through building stone bunds on slopes, thus creating terraces and construction of check dams in erosion valleys (Bossuyt and Hermy, 2001). In addition to biological techniques that make use of plantation of seedlings and natural restoration are receiving more and more attention nowadays.

These techniques combine both soil stabilization and reforestation. One widely used option here is the creation of closed areas, also called exclosures, i.e., demarcated areas under strict conservation management, often controlled by a local community (Nyssen *et al.*, 2004). In these areas, cultivation, collection of fuelwood and grazing are forbidden, whereas harvesting of grass is strictly controlled in order to allow natural forest restoration from soil seed bank (Reubens *et al.*, 2007). Dormant seeds (viable seeds that would germinate when favorable conditions set in) in the soil, collectively known as the seed bank, play a crucial role in the restoration of vegetation (Hirsch *et al.*, 2012). Seed dormancy or longevity is an important parameter for the existence of a seed bank. Seed banks are formed by seeds, either born and produced on site or carried to the site by dispersal agents and accumulated in the soil and form seed banks (Hirsch *et al.*, 2012).

According to published reports, e.g., Mirutse Giday and Gobena Ameni (2003), Gidey Yirga (2010a), Gidey Yirga (2010b), Girmay Zenebe *et al.* (2012), Abraha Teklay *et al.* (2013), Gebremedhin Gebreegzabher *et al.* (2013), Kalayu Mesfin *et al.* (2013), Moravec *et al.* (2014), Abraha Teklay (2015), Meaza Gidey *et al.* (2015), Fitsumbirhan Tewelde *et al.* (2017), studies

on medicinal and other uses of plants in Tigray is increasing. Furthermore, there are unpublished MSc. and PhD thesis on ethnobotanical studies conducted by Gebremedhin Hadera (2000), Abrha Tesfay (2008), Nurya Abdurhman (2010), Genet Atsbeha (2012) and Tadesse Beyene (2015). However, only few of the above-mentioned researches, have tried to assess the interaction of the local communities with respective nearby remnant forests (Gebremedhin Hadera, 2000; Abrha Tesfay, 2008) and their contribution to the livelihood of local communities.

It is known that traditional knowledge on conservation of forests and their uses have been transmitted orally from generation to generation. *Wejig-Mahgo-Waren* forest is one of the remnant dry Afromontane forests in northern Ethiopia which plays a crucial role in the livelihoods of local communities. However, because of the informal nature of oral transmission, there is a need to investigate and document the traditional use of plant species and management of the forest resources by the local people for sustainable resource utilization for the present as well as the future generations. Thus, accommodating new conservation approaches, such as participatory forest management could contribute significantly to mitigate the problem of forest destruction (Zerihun Woldu *et al.*, 2002).

In response to the loss of forests, the Ministry of Environment and Forest of Ethiopia (MEF) designated some Pilot REDD+ Project Sites Visit Report (Ministry of Environment and Forestry, 2015). Likewise, the Ministry of Agriculture (Ministry of Agriculture, 2015) and the Ethiopian Wildlife and Natural History Society (EWNHS, undated) independently made assessments on potential forest areas in Tigray. And all of these institutions identified *Wejig-Mahgo-Waren* massif forest as a potential area for restoration. Tigray region has six state forests, namely *Wejig-Mahgo-Waren* (Southern zone), *Hugumburda-Gratkahassu* (Southern zone), *Hirmi* (North

Western zone), *Waldiba* (North Western zone), *Asimba* (Eastern zone) and *Dess'a* (Eastern zone) (Ministry of Agriculture, 2015). Among these, the vegetation ecology of *Hugumburda-Gratkahsu* (Leul Kidane, 2015; Ermias Aynekulu (2011), and *Dess'a* (Ermias Aynekulu, 2011) have been studied. However, no work has been conducted to date on the vegetation ecology, population structure, regeneration status, soil seed bank, cognitive domain, use and management of *Wejig-Mahgo-Waren* massif forest. It is, therefore, imperative and urgent to assess the standing vegetation, soil seed bank of the forest and indigenous botanical knowledge of the forest fringe communities.

1.2 Statement of the research problem

The vascular plant species of *Wejig-Mahgo-Waren* massif forest have not been investigated previously; the plant communities that exist in the forest have not been described; the regeneration status of the forest has not been known; the species present in the soil seed bank and the restoration potential of the forest have not been quantitatively assessed. Information on how the surrounding communities look upon, use and manage the forest resources has not been documented. The research ideas presented below were developed to address the gaps identified by applying a combination of relevant methods from the fields of plant taxonomy (floristics), plant ecology, and ethnobotany.

1.3 Research questions, hypotheses and objectives

1.3.1 Research questions

1. What is the vascular plant species composition in *Wejig-Mahgo-Waren* massif forest?
2. What type of plant communities exist in *Wejig-Mahgo-Waren* massif forest?
3. What is the regeneration status of trees/shrubs in *Wejig-Mahgo-Waren* massif forest?

4. What is the soil seed bank of *Wejig-Mahgo-Waren* massif forest?
5. What are the correlations between the standing vegetation and species richness in the soil seed bank?
6. What is the interaction between *Wejig-Mahgo-Waren* massif forest and the communities living around the forest?

1.3.2 Research hypotheses

H₀: The fringe communities around *Wejig-Mahgo-Waren* massif forest exert high pressure on the forest through extraction of its resources for several use values and deforestation leading to less floristic diversity and community types with very low restoration potential from soil seed bank.

H_a: The fringe communities around *Wejig-Mahgo-Waren* massif forest exert no pressure on the forest making the forest high in floristic diversity and community types with very good restoration potential from soil seed bank.

1.3.3 Objectives

1.3.3.1 Major objective

- ✓ To study the vegetation composition, population structure, soil seed bank and ethnobotanical knowledge of fringe communities in and around *Wejig-Mahgo-Waren* massif forest in Southern Tigray, Ethiopia.

1.3.3.2 Specific objectives

- ✓ To record the vascular plant species composition in *Wejig-Mahgo-Waren* massif forest.
- ✓ To describe and classify plant community types found in the study forest.
- ✓ To assess the population structure and regeneration status of the study forest.
- ✓ To assess the soil seed bank of the forest compared to the standing vegetation.
- ✓ To assess the interaction of forest fringe communities with forest vegetation in and around *Wejig-Mahgo-Waren* massif forest.

CHAPTER TWO

2. LITERATURE REVIEW

2.1 Vegetation types of Ethiopia

A large proportion of the Eastern Afromontane and Horn of Africa biodiversity hotspots lie in Ethiopia and the country is one of the biodiversity centers of the world (Conservation International, 2007). Ethiopia has a complex relief and a variety of climates and thus diverse habitats with rich flora and fauna. After completion of the writing of the modern flora of Ethiopia, an article published by Ensermu Kelbessa and Sebsebe Demissew (2014), showed that the total number of vascular plant species is 5,757 with 10.58% endemism. The Ethiopian vegetation is highly influenced by climate, which is associated with elevation (Dugdale, 1964). Southwestern Ethiopia receives more precipitation than other parts of the country due to the humid air coming from the Congo basin (Dugdale, 1964). This westerly wind, however, cannot penetrate further than 30°E and rarely influences the Horn region because the Ethiopian highlands act as barriers. The flora of southern Ethiopia is more similar to that of Kenya and Uganda than the flora of northern Ethiopia (Dugdale, 1964). The vegetation on the Afromontane belt (900-3200 m) of Ethiopia has been under tremendous pressure from human activities and over grazing, which has led to the replacement of the evergreen forests by grasslands (Tewolde Berhan Gebre Egziabher, 1988).

The most frequently cited studies on the vegetation of Ethiopia include that of Pichi-Sermolli (1957), Breitenbach (1963), White (1983) and Friis (1992). The extent and delineation of the vegetation maps of Pichi-Sermolli (1957) and Breitenbach (1963) are very similar, but differ in the descriptions and terminology of the mapping units (Friis and Sebsebe Demissew, 2001).

According to Pichi-Sermolli (1957), the vegetation of northern Ethiopia is classified as montane evergreen thicket and savanna. The common species in this vegetation type include *Juniperus procera*, *Olea europaea* subsp. *cuspidata*, *Acokanthera schimperi*, *Carissa spinarum* and species of *Euclea*, *Rhamnus*, *Rhus* and *Maytenus* (Friis and Sebsebe Demissew, 2001).

Breitenbach (1963) mapped the vegetation of Ethiopia and Eritrea into seven basic categories using a physiognomic approach. The seven categories are Lowland savannah, Lowland savannahs, Lowland woodlands, Highland forests, Mountain woodlands, Mountain savannahs and Mountain savannah. The sub-categories are based on local rainfall (arid, semi-arid, semi-humid and humid) and stages of succession, which range from pioneer to climax types. According to Breitenbach's (1963) vegetation map, the three dominating vegetation types in Ethiopia in a decreasing order are Lowland steppes, Lowland woodlands and Mountain savannahs.

White (1983) classified the vegetation of Africa into 81 vegetation groups using physiognomy and floristic composition. Accordingly, the dominant vegetation types in Ethiopia are Forest transitions and mosaics, Woodland, Woodland mosaics and transitions, Bushland and thicket, Semi-desert vegetation, Grassland, Edaphic grassland mosaics, Altimontane vegetation, Azonal vegetation and Desert. The vegetation on the lower escarpment is classified as bushland and thicket and the vegetation on the Tigrean plateau and the upper escarpment as forest transition and mosaics.

Friis (1992) classified the forests of the Horn of Africa into nine vegetation types. He followed the classification and terminologies of White (1983) and added the lowland semi-deciduous forest (dry peripheral semi-deciduous Guineo-Congolian forest) and the altitudinally transitional

forest (transitional rain forest), which were identified after White's vegetation map had been published. According to the classification of the forests and forest trees of northeast tropical Africa (Friis, 1992), *Wejig-Mahgo-Waren* massif forest is broadly categorized as a dry single-dominant Afromontane forest. It is characterized by a dry climate annual precipitation less than 1000 mm, and with *Juniperus procera* and *Olea europaea* subsp. *cuspidata* as the dominant species. This dry single-dominant Afromontane forest of the escarpment and transition between single-dominant Afromontane forest and East African evergreen and semi-evergreen bush land that occurs between 1500 and 2400 characterizes the escarpments in northern Ethiopia.

The vegetation map of Ethiopia was later simplified into eight major vegetation types by Sebsebe Demissew *et al.* (1996) as Afroalpine and sub-Afroalpine zone, dry evergreen mountain forest and grassland, moist evergreen mountain forest, evergreen scrub, *Combretum-Terminalia* and savanna, *Acacia-Commiphora* woodland, lowland (semi-) evergreen forest, desert and semi-desert scrubland and coastal vegetation. According to Sebsebe Demissew *et al.* (1996), dry evergreen mountain forest, grassland and evergreen scrub characterize the forest remnants in northern Ethiopia. The most recent revision of the vegetation map of Ethiopia was by Friis *et al.* (2010). In this work twelve vegetation types are recognized Desert and semi-desert scrubland, *Acacia-Commiphora* woodland and bushland, Wooded grassland of the western Gambela region, *Combretum-Terminalia* woodland and wooded grassland, Dry evergreen Afromontane forest and grassland complex, Moist evergreen Afromontane forest, Transitional rainforest, Ericaceous belt, Afro alpine belt, Riverine vegetation, Fresh-water lakes, lake shores, marsh and floodplain vegetation, Salt-water lakes, salt-lake shores, marsh and pan vegetation. The classification was mainly based on altitude and/or climatic variables.

2.2 Determinants of biodiversity

Latitude and elevation are important drivers of species distribution patterns (Gaston, 2000; Lomolino, 2001). However, the effect of these two factors varies with local, regional and global scales, as well as with historical factors (ter Steege and Zagt, 2002). A palaeobotanic study by Bobe (2006), for instance, indicates that the differentiation of moist forests and drier *Acacia* woodlands in Africa occurred during the early Cenozoic period. Pollen studies in Kenya (Lamb *et al.*, 2003) and northern Ethiopia (Darbyshire *et al.*, 2003) indicate that the flora of East Africa had experienced several changes due to climate change and anthropogenic disturbances.

At a global scale, diversity decreases with increasing distance from the equator (Gaston, 2000; Mutke and Barthlott, 2005; Kreft and Jetz, 2007). However, the general species-richness pattern along latitudinal gradients is sometimes reduced due to other factors, like longitude, elevation, topography and aridity (Gaston, 2000). There are several reasons for the higher species diversity in the tropics compared to other regions, one of which is the larger area of the tropics than of any other zone (Rosenzweig, 1995).

The other reason for a large number of species in the tropics is the low-temperature variability in space and time that increases the probability of similar habitats (Rosenzweig, 1995). Constant and higher temperatures in the tropics also contribute to higher species diversity by increasing the rate of metabolism, which speeds up the passing of generations and increases the rate of mutation that again leads to new species (Tokeshi, 1999).

Elevation plays a major role in plant species diversity and floristic formations, especially in mountainous areas (Gaston 2000; Kreft and Jetz, 2007). Compared to species-latitude and

species-area relationships, the species-elevation relationship is less studied (Lomolino, 2001). Elevation determines species distribution in montane systems by influencing area, climate, geographical isolation of montane communities and feedback among zonal communities (Kreft and Jetz, 2007). Generally, habitat diversity is a widely accepted determinant of species diversity (Mutke and Barthlott, 2005). The theory of spatial heterogeneity (Pianka, 1966) stated that more heterogeneous and complex physical environments support more diverse plant and animal communities.

Apart from environmental drivers of species diversity, anthropogenic disturbance affects species diversity through habitat loss and fragmentation (Brooks *et al.*, 2002). Habitat loss is the leading cause of species extinction (Pimm and Raven, 2000). Biodiversity loss is more serious than other environmental threats because it is irreversible (Mittermeier *et al.*, 1998). Habitat fragmentation affects biodiversity by reducing the area of a habitat, which reduces the persistence of a species and through the negative edge effect that increases mortality while decreasing reproduction (Farhig, 2003).

Identifying diversity hotspots can support conservation planning (Myers *et al.*, 2000). Conservation International (2007), for instance, used the world biodiversity hotspots to prioritize biodiversity conservation areas. The hotspot concept is based on species richness, endemism and threat (Myers *et al.*, 2000). Studies at global scale have documented less overlap among the three criteria (Orme *et al.*, 2005). Nearly half of the world's vascular plant species are endemic to 25 hotspots (Myers *et al.*, 2000), of which 17 are in tropical forests each having at least 1500 endemic plant species (Brooks *et al.*, 2002). However, many plant species have a too small population sizes to be viable and this may increase the rate of extinction in the long term

(Whitemore, 1997). The human population growth rate in the hotspots is higher than the average rate worldwide, suggesting that human-induced habitat loss is still a major threat (Cincotta *et al.*, 2000). Mckee *et al.* (2004) found a strong relationship between population growth and number of threatened species.

Recently, nine new areas were included as part of the world biodiversity hotspots (Conservation International, 2007). Among these are the Horn of Africa and eastern Afrotropical hotspots that include large parts of the lowlands and highlands of Ethiopia, respectively. However, Africa has fewer endemic species and is less rich in tropical species, which is probably due to the slow rate of geographical speciation (Rosenzweig, 1995). Large-scale vegetation changes in Africa occurred due to changes in the atmospheric moisture content driven by tropical sea surface temperature changes in the mid-Pleistocene (Schefus *et al.*, 2003).

2.3 Plant communities

A plant community is defined as the collection of plant species growing together in a particular location that show a definite association or affinity with each other (Kent and Coker, 1992). Plant community study is a useful approach in conservation planning (Ferrier *et al.*, 2009). The concept of plant community, for instance, provides useful information on the underlying environmental drivers of species distribution, as plants that live together have similar environmental requirements for their existence. The principles of plant communities are based either on the continuum or on the individualistic theories. Clements (1916) in his plant succession theory considered plant communities as one big organ (superorganismic concept) composed of various species, which repeats itself with regularity over a given region. In his holistic approach, forests, scrubs and grassland are considered as major groups of vegetation,

which can develop to climax communities with sufficient time and long-term stability. Gleason (1926), on the other hand, considered plant species distribution as a continuum. Gleason takes environmental factors and tolerance ranges of species as determinants of the existence and abundance of a species in a given region. This viewpoint is known as the individualistic concept of the plant community.

Although differences still exist among ecologists on the concepts of plant communities, the Gleason's individualistic theory has been widely used (Callaway, 1997). Plant ecologists who favor vegetation classification follow the approach of Clements and group species into communities. Those ecologists who do not believe in classification follow the continuum theory and arrange species along environmental gradients as continua, using ordination methods (Kent and Coker, 1992).

2.4 Multivariate analysis

Multivariate analysis of community data is very challenging due to various reasons (Green, 1979), namely noise, redundancy, relationship and outliers and large data matrix. To facilitate communication among ecologists, there is a need for the selection of only a modest number of preferred methods for the common usage (Pielou, 1977). In order to meet these challenges, ecologists have developed, applied and tested a number of multivariate methods, among others, the two are cluster analysis and ordination.

2.4.1 Cluster analysis

For ecological data, cluster analysis classifies sites, species or variables. Cluster analysis is an explicit way of identifying groups in raw data and helps us to find structure in the data. There are

several types of cluster analysis, based on different ideas about the cluster concept. A major distinction can be made between divisive and agglomerative methods. A second way of distinguishing methods is to classify them by hierarchical and non-hierarchical methods. Hierarchical methods start from the idea that the groups can be arranged in a hierarchical system.

2.4.1.1 Hierarchical agglomerative methods

Agglomerative methods start with individual objects, which are combined into groups by collection of objects or groups into larger groups. Agglomerative methods require a similarity or dissimilarity matrix (site by site). In many sciences, agglomerative techniques are employed much more frequently than divisive techniques. The historical reason for this is the inefficient way early polythetic divisive techniques used computer resources, while the agglomerative ones were more efficient (van Tongeren, 1995).

The first step in agglomerative clustering is to choose a distance or dissimilarity measure, such as euclidean, manhattan or similarity ratio. Hierarchical clustering method produces a dendrograms, which allow the user to read off the “height” at which items, or clusters or both are combined which together would form a new and larger cluster (Zerihun Woldu, 2017). Clustering algorithms, which link different groups with each other, are principally of three types: single linkage, complete linkage and average linkage. Average linkage is most often used in ecology (average linkage includes also popular ward method (weighted average linkage) and beta flexible).

Ward's method, also known as error sum of squares clustering, is in some respects similar to average-linkage clustering and centroid clustering. Between-cluster distance can either be computed as a squared distance between all pairs of sites in a cluster weighted by cluster size

(resembling average-linkage clustering) or as an increment in squared distances towards the cluster centroid when two clusters are fused (resembling centroid clustering). Penalty by squared distance and cluster size makes the clusters tighter than those in centroid clustering and average linkage and more like those obtained in complete linkage (van Tongeren, 1995).

2.4.2 Ordination analysis

Ordination is the collective term for multivariate techniques that arrange sites along axes on the basis of data on species composition. The use of Principal Component Analysis (PCA), Redundancy Analysis (RA), Canonical correspondence Analysis (CCA) or Canonical Analysis (CA), therefore, depends on whether the relationship between species data and environmental data is linear or unimodal. Species respond linearly along environmental gradient, which could be true for rather homogeneous ecological data, where ecological gradients are not too long, while species respond unimodally along gradient is having its optima at certain gradient position. This model is more close to reality of ecological data and is more suitable for heterogeneous datasets structured by strong or long ecological gradients, with high species turnover and many zeroes in the species matrix (Zerihun Woldu, 2017).

Lepš and Šmilauer (2001) proposed a simple rule to decide whether to use unimodal or linear method based on the length of the first DCA axis. If the length is > 4 , data are heterogeneous and unimodal methods should be used. If the length is < 3 , data are homogeneous and linear methods should be used.

Legendre and Legendre (1998) and Zerihun Woldu (2017) recommend the following conditions while choosing the approaches to fit our specific purposes based on the nature of the input data:

PCA and RDA should be used to analyze species data if the relations along the gradients are linear. CA and CCA can be used to analyze unimodal relationships between species and environmental variables.

2.5 Measures of species diversity

Whittaker (1972) stated that species diversity can be interpreted in terms of species richness and evenness and identifies alpha, beta and gamma types of species diversity. Alpha diversity (species richness) is the number of species per standard size or community. Beta diversity is the difference in species diversity between areas or communities. It is sometimes called habitat diversity because it represents differences in species composition between different areas or environments (Whittaker, 1972; Kent and Coker, 1992). Since communities and habitats are often difficult to delineate, beta-diversity is measured among study quadrats (Mark, 2001). The total or gamma diversity of a landscape or geographic area is a product of the alpha diversity of its communities and the degree of beta differentiation among them (Whittaker, 1972). The difference between alpha and gamma diversity is a matter of scale, which is often subjectively defined (Peet, 1974).

A large number of diversity indices have been used to measure species diversity. Magurran (2004) provided an in-depth review of concepts and measurements of diversity. Species richness, meaning a count of the number of plant species in a quadrat, area or community, is often equated with diversity. However, as Magurran states, most methods used in measuring diversity actually consist of two components.

The first is species richness and the second is the relative abundance (evenness or unevenness) of species within a sample or community. Whittaker (1972) considers species richness as a strong measure of species diversity. However, using species richness *per se* as a measure of diversity is criticized, because species richness is just one component of species diversity (Sanjit and Bhatt, 2005). The Shannon index and Simpson's index of diversity, which combine species richness with relative abundance, are widely used in species diversity studies (Kent and Coker, 1992). The Shannon index expresses the relative evenness or equitability of species, while Simpson's index (Simpson, 1949) gives weight to dominant species (Whittaker, 1972). The diversity indices are biased either on species richness or species evenness, which makes it difficult to obtain one robust index of diversity measurement (Magurran, 2004). The Shannon index is insensitive to rare species (Sanjit and Bhatt, 2005).

2.6 Plant population structure and regeneration status

Studies on population structure and density of major canopy tree species help to understand the status of regeneration of species and management history and ecology of the forest. Plant population structure shows whether or not the population has a stable distribution that allows continuous regeneration to take place. Size class distribution as diameter at breast height (DBH) and height are used to evaluate structure and regeneration of species. The structure of the species may have inverted J-shape pattern, which is an indicator of stable population structure and is a reflection of healthy regeneration potential of the species exhibited in a natural forest where disturbance is minimum (Harper, 1977; Getachew Tesfaye *et al.*, 2010). Any population pattern that differed from inverted J-shaped, have been attributed to disturbed forest (Poorter *et al.*, 1996), e.g. bell shape, J-shape and U-shape.

The variation in relative abundance of size classes and population structure of species is the result of past and present disturbance as well as management history of the forest. Based on intensity of disturbance species show variation in population structure pattern reflected through difference in the abundance of different size classes (Tamrat Bekele, 1993). Size class distribution or population structure gives good indication of the impact of disturbance and forest successional trends.

The population structure, characterized by the presence of sufficient population of seedlings, saplings and adults, indicates successful regeneration of forest species. Seedling densities in forest understory's are dynamic and rates may vary among species and in gap and shade environments. Information on tree seedling ecology can provide options for forest development through improvement in recruitment, establishment and growth of the desired species. Regeneration studies have significant implication on the conservation and restoration of degraded natural forests (Getachew Tesfaye *et al.*, 2010). Thus, regeneration status of woody species can be predicted by the age structure of their populations. The study of regeneration of forest trees has important implications for the management of natural forests and in one of the thrust areas of forestry. Research in this field contributes to planning, conservation and decision making in forest resources management programs (Pokhriyal *et al.*, 2010).

2.7 Soil seed bank and its importance in vegetation study

The soil seed bank refers to all viable seeds and fruits present in the soil. Soil seed banks can be either transient, with seeds that germinate within a year of initial dispersal, or persistent, with seeds that remain in the soil for more than one year (Simpson *et al.*, 1989). They exhibit

variations in space as well as time and display both horizontal and vertical dispersion, reflecting initial dispersal onto the soil and subsequent movement (Simpson *et al.*, 1989).

Soil seed banks reflect partly the history of the vegetation and can play an important role in its restoration after disturbances. They have been exploited in two contexts: to manage the composition and structure of existing vegetation and to restore or establish native vegetation (van der Valk and Pederson, 1989).

The bank of long-lived seeds in the soil stores over many decades and contributes as a source of propagules that ensure continual occupation of a site after disturbances while serving as a gene pool by buffering genetic changes in the population (Hill and Morris, 1992). Soil seed banks play a crucial role in the dynamics of plant populations. In forest management, natural seed banks play a vital role in restoration after disturbances, for example, tree felling. The fact that many economically important trees are canopy species whose seeds have little dormancy makes it important to leave some individuals of the species to act as local seed sources. Knowledge of which species are not represented in the persistent soil seed bank can be just as important as knowing which species are represented. This is especially true in the management of vegetation for conservation. The dynamics of a soil seed bank include recruitment into the dormant seed bank population through seed rain, losses from the dormant seed bank through seed predation or death and change into the active seed bank to germinate and form a seedling bank, through a stimulus.

2.7.1 Seed longevity in the soil

Depending on internal and external environmental factors as well as species to species interactions, the period of seed viability in the soil varies. The longevity of seeds in soil commonly depends on the conditions through which they pass; for example, some seeds can stay long in the soil if kept in dry atmosphere. Viability is also affected by ageing; with increase in age viability of seeds decreases until it stops completely. The common reasons for loss of viability are denaturation and inactivation of proteins and enzymes, over drying and exhaustion of reserve foods of dormant seeds due to respiration (Rajan, 2000).

The longevity of seeds in the soil seed bank associated with the principle of dormancy caused by either seed coat, condition of embryo, light sensitivity, or chemical inhibitors. The dormancy generally helps the embryo to pass adverse environmental conditions (Rajan, 2000).

Evidences of longevity in seeds come from archeological sites, dated herbarium sheets, or experiments on shelf-stored seeds or seeds buried in the field. However, only viability tests of seeds, which have been subjected to the moisture, temperature and gaseous compositions of soil under field condition provide results of ecological importance (Demel Teketay and Granström, 1997).

2.7.2 Significance of soil seed banks

The seed banks found in different environments represent a record of past as well as present vegetation growing on the area and nearby. If an existing vegetation stand is destroyed by various causes, the seed bank will immediately serve as a source from which new vegetation

arises (Harper, 1977). In addition, their significance in restoration of lost vegetation, seed banks are also essential in rehabilitation of a degraded land.

2.7.3 Spatial variation

The vertical distribution of the soil seed bank and number of species was consistent at all sites with the highest densities in the upper three centimeters of soil and then gradually decreasing densities with increasing depth (Demel Teketay and Granström, 1997). There was considerable variation among species in depth distribution. Some species, e.g. *Juniperus procera*, *Clematis hirsuta*, *Girardinia diversifolia* and *Pilea tetraphylla* were almost entirely confined to the litter layer. Some other species, e.g. *Laggera crispata*, *Lobelia giberroa*, *Crassula alsinoides*, *Veronica abyssinica* and *Poa leptoclada*, had almost uniform depth distribution (Demel Teketay and Granström, 1997).

Another set of species, e.g. *Indigofera rothii*, *Solanum nigrum* and *Eragrostis schweinfurthii* had their seeds distributed in the deeper layers. In general, seeds of herbs, grasses and sedges were more deeply distributed in the soil than those of trees, shrubs and climbers. Herbs, grasses and sedges have small seeds compared with many woody species and species with small seeds have a better chance of becoming buried in deeper layers of the soil (Thompson, 1987). Considerable intra-site differences of horizontal distribution of seeds in the soil were also apparent among species. Some species, such as *Crassula alsinoides* and *Dichrocephala integrifolia*, showed a relatively even distribution while most of the woody species and some herbaceous species had patchy or clumped distribution.

2.7.4 Seedling emergence and seed extraction techniques

Several methods have been used to estimate the density and composition of soil seed banks and there has been much debate on the utility of the different approaches (Bernhardt *et al.*, 2008). The ‘seedling emergence method’, which involves spreading a thin layer of soil over a sterilized medium in a glasshouse and identifying and counting emerging seedlings, is commonly used (Ter Heerd *et al.*, 1999). This method gives a good indication of the more germinable seed bank, but does not provide a complete assessment of the seed bank flora unless the soil sample is maintained for extended periods at simulated habitat conditions (Thompson and Grime, 1979).

Indeed, the seedling emergence method can dramatically underestimate the density of the seed bank due to errors associated with seed dormancy and specific environmental requirements for germination (Bernhardt *et al.*, 2008; Wright and Clarke, 2009).

A second method, regarded as providing a better estimate of total seed bank densities, is seed extraction from the soil (Brown, 1992). This can be achieved through ‘rinsing’, flotation in a salt solution or other high specific gravity liquid, or a combination of both (Bernhardt *et al.*, 2008).

Seeds are then hand sorted under a microscope and identified. Seed extraction is not commonly used as it is time consuming, ineffective at finding small seeded species and may overestimate the viable seed bank by including non-viable seeds (Baskin and Baskin, 1998).

Besides, depending on when samples are collected, seeds may lose viability before the next germination season, i.e., if one determine the seed content several months before the next germination season, one do not know if these seeds would have survived until the germination season.

Studies comparing these two methods have found contrasting results (Johnson and Anderson, 1986; Brown, 1992). Seed extraction methods detected greater seed densities and species richness of a forest soil seed bank compared with seedling emergence techniques (Brown, 1992). Johnson and Anderson (1986) reported similar estimates for seed density, but greater species richness of the soil seed bank with the emergence method. Bernhardt *et al.* (2008) found that the seedling emergence method failed to detect nearly 90% of all most germinable seeds found via the 'rinsing method'. Despite large differences between the two approaches, seedling emergence is the most commonly used method (Capon and Brock, 2006; Robertson and James, 2006; Poorter *et al.*, 2007).

2.8 Ethnobotany and its relevance

2.8.1 The concept of ethnobotany

Ethnobotany is a multidisciplinary science encompassing botany, anthropology, economics and linguistics, few among others, which studies the way in which a society relates to its environment and particularly to the plant world. These relationships can be social, economic, symbolic, religious, commercial and artistic (Aumeeruddy-Thomas and Pei, 2003). Ethnobotany as an interdisciplinary science is in a position to contribute much to plant conservation. Ethnobotany was originally based largely on qualitative methods, such as inventories of plants and their uses, with a major focus on the economic importance of plants. In addition, it focuses on documenting, analyzing and using of indigenous knowledge, beliefs and practices related to plant resources (Martin 1995; Balick and Cox, 1996; Zemedu Asfaw, 1997). Consequently, applied ethnobotany strives to bridge the gap between traditional knowledge and scientific

knowledge and to understand the relationships between local practices and knowledge systems and policies, rules and economic trends at the national and international level (Aumeeruddy-Thomas and Pei, 2003). The contribution of applied ethnobotany is not limited to pure science, but has an important role to play in understanding the dynamic relationships between biological diversity and social and cultural systems and their development.

2.8.2 Relevance of scientific documentation in indigenous botanical knowledge

Ethnobotany is an indispensable tool to identify and document plant species that have been underutilized by human beings for centuries for various reasons (Martin, 1995; Balick and Cox, 1996; Tilahun Teklehaymanot and Mirutse Giday, 2007).

Traditional people all over the world, through their indigenous knowledge know which plant species are threatened and need priority management (Given, 1994; Aumeeruddy-Thomas and Pei, 2003). Based on this knowledge, ethnobotanists need to study the traditional diverse use of plants and the indigenous knowledge on management in a scientific way. This ethnobotanical information will further enable to set priority management for those areas and species useful for human consumption (i.e., culturally and economically useful plants) and areas of high species diversity (Given, 1994) as well as areas and plant species threatened due to habitat change and overuse.

Moreover, ethnobotany becomes useful tool in the introduction of alternative resource management systems that involve local people—the key generators, custodians and promoters of local biodiversity (Rastogi *et al.*, 1998), which helps in the effective joint-management of resources between government and local people.

Therefore, rigorous ethnobotanical study is indispensable to get the information from the indigenous societies using different approaches and methods in a scientific way before its death with the knowledgeable elderly people (Martin, 1995; Cotton, 1996; Furze *et al.*, 1997; Zemedu Asfaw, 1997; Mushongi, 2001). Ethnobotany faces challenges from loss of biodiversity and the low regard to indigenous knowledge (Martin, 1995; Balick and Cox, 1996), which needs immediate attention by all concerned bodies for the benefit and continued existence of all living things.

2.8.3 Recent methodological advances in ethnobotanical studies

2.8.3.1 Perspectives of ethnobotanical data collection

Ethnobotanical study refers to the broad range of data scientifically collected by researchers on the interactions between people and plants. Data are recorded in many different forms, collection of plants, recorded by interviews, laboratory analyses, photographs, market surveys and so on (Martin, 1995). When an informant describes the use of a plant, he/she is clearly or indirectly denoting to the concepts and categories drawing from his/her cultural experience that may or may not correspond to the concepts and categories of the ethnobotanist (Alexiades 1996).

Ethnobotanists follow definite sets of procedures to retrieve reliable ethnobotanical data set. The most important and first step in collecting ethnobotanical dataset is to delimit the cultural domain or subject of interest. For example, the cultural domain chosen may be plants used as stomachaches or as edible fruits. The next important step is to define how to categorize and measure local knowledge and management of the environment. Either types of cognitive systems or behavioral study approaches may be followed when collecting data on the local use categories

or domains of the cultural significance of plants and their relationships with the environment and community. An emic point of view corresponds to the perceptions, nomenclature, classifications, knowledge, beliefs, rules, and ethics of the local plant world as defined by a native of the local cultural community. Emic knowledge allows a native person to behave in culturally appropriate and meaningful ways in different cultural contexts. An etic denotes the conceptual categories and organization of the ethnobotanical environment according to the researcher, who often is alien to the local culture and whose conceptual system ideally derives from the language and rules of science (Zent, 1996).

2.8.3.2 Perspectives of ethnobotanical data analysis

Since Harshberger coined the term 'ethnobotany', the field of study has long been criticized for being too qualitative (Albuquerque, 2009) until efforts were made to apply quantitative techniques to the direct analysis of field-collected plant use data. There has been increasing trend in using quantitative techniques in the analysis of ethnobotanical data (Prance *et al.*, 1987; Martin, 1995). Ethnobotanical methods are increasing from qualitative to quantitative analysis. The methods of ethnobotany started with the qualitative approach and gradually added more quantitative methods. Phillip and Gentry (1993) defined quantitative ethnobotany as the direct application of quantitative techniques to the analysis of contemporary plant use data. Progress made in the field of quantitative ethnobotany allowed to employ reproducible methods, statistical measures of variation and hypothesis testing in ethnobotanical research (Höft *et al.*, 1999). Most recently, data collected from ethnobotany can be quantitatively analyzed through three major categories, namely informant consensus, subjective allocation and use totaled.

In the method of informant consensus factor, the relative importance of each use is calculated directly from the degree of consensus in informants' responses. The importance of different plants or uses is assessed by the proportion of informants who independently report knowledge of a given use or who claim to have used a plant in a specific way (Phillips *et al.*, 1994). In the subjective allocation method, the relative importance of each use is subjectively assigned by the researcher. The importance of different plants or uses is estimated by the researcher on the basis of his or her assessment of the cultural significance of each plant or use (Stoffle *et al.*, 1990). In the uses totaled method, no attempt is made to quantify the relative importance of each use. The number of uses are simply totaled, by category of plant use, plant taxon, or vegetation type. Not surprisingly, this has been the most popular approach, since it is the fastest and most straightforward way to quantify ethnobotanical data according to some researchers (Bennett, 1992).

Höft *et al.* (1999) presented elaborated techniques on the application of multivariate statistical analyses in ethnobotany. Castaneda and Stepp (2007) developed the ethno-ecological importance value index (EVI) by combining the cultural and ecological data to evaluate the cultural importance of vegetation types. Depending on the aim of the research, it is possible to make comparisons between different cultural groups, plant species and vegetation categories from ethnobotanical perspectives. This includes identification of differences in the relative cultural importance of species or determination of the magnitude of ethnobotanical knowledge held by different demographic groups or communities. Relative Cultural Importance (RCI) indices are quantitative measures designed to transform the complex, multidimensional concept of “importance” into standardized and comparable numerical scales or values. Per-taxon plant use citation data from ethnographic plant interviews is applied to RCI formulas to derive values.

A range of cultural significance indices (CSI) has been developed to measure the cultural importance of a species or a family of plants provided to a community (Hoffman and Gallaher, 2007). The Cultural Significance Index (CSI), an anthropological approach presented by Turner (1988) and modified by Stoffle *et al.* (1990) and Silva *et al.* (2006), calculates importance through researcher- determined weighted ranking of multiple factors. Turner (1988) assigned scores on a five-point scale to the variables of quality and intensity of use and assigned a score of 2, 1, or 0.5 for the exclusivity or preference of use. To reduce the subjectivity of this approach, Silva *et al.* (2006) revised the CSI with a two-point scale for the variables. They also incorporated a consensus method called a correction factor to reduce the sensitivity of this method to sampling intensity. The ethnographic, qualitative approach of the CSI method requires considerable experience and rapport with a cultural group for meaningful results.

2.8.4 Status of forest resources and threats on indigenous botanical knowledge

An increasing demand for arable land and forest products due to the rapidly growing population and poverty of the rural people have also been mentioned as major threats to the survival of many of the Ethiopian plant species (Ensermu Kelbessa *et al.*, 1992). If present trends in population growth continue, deterioration of natural resources will be more rapid in the future and will result in a great loss of biodiversity (Ensermu Kelbessa *et al.*, 1992; Zerihun Woldu, 1999).

Traditional plant use is of extremely high importance in many societies and is prevalent in African communities who lived in harmony with the natural resources for centuries without bringing any detrimental effect on the survival of the biodiversity (Thrupp, 1997; Bussmann, 2006). According to Plotkin (1995), indigenous people living in forests best know, use and

protect biodiversity. However, the survival and existence of indigenous people and their long-term accumulated knowledge faces challenges, because of modernization, genetic erosion on plant and animal resources, low recognition to their knowledge and varied culture, loss of biodiversity (Martin, 1995; Balick and Cox, 1996; Thrupp, 1997; Almaz Negash, 2001; Bussmann, 2006).

Indigenous knowledge face challenges from genetic erosion because of the destruction of trees and bush land, including land clearance for agricultural lands, the collection of fuelwood and the burning of forests and bush to provide pastureland (Almaz Negash, 2001). In addition to these, indigenous people are also suffering from the displacement and restriction of using their local ecosystems and resources because of the establishment of strict protected areas (Furze *et al.*, 1997), which is believed to be one factor for the encroachment and destruction of most of the world-protected areas.

CHAPTER THREE

3. MATERIALS AND METHODS

3.1 Study area description

3.1.1 Location of the *Wejig-Mahgo-Waren* massif forest

Wejig-Mahgo-Waren massif forest is an elongated chain of natural forest that is located in Southern Tigray at about 630 km North of Addis Ababa, the capital of Ethiopia, and 120 km south of Mekelle town, the capital of Tigray National Regional State. It is located between 12° 47'-12° 58' North latitude and 39 ° 30'-39 ° 48' East longitudes with altitudinal variation from 1627 to 2970 m.a.s.l., covering a total area of 8,772 ha (MEF, 2015).

According to MEF (2015), *Wejig-Mahgo-Waren* massif forest is partly degraded with remnant of some intact forest patches. The patches of remnant natural forests belong to the Dry Afromontane forest types with dominant trees, including *Juniperus procera*, *Olea europaea* subsp. *cuspidata* and *Acacia abyssinica*. The vegetation type in most of the area is of the secondary forest type.

The lower part of *Wejig-Mahgo-Waren* massif forest seen adjacent to the highway from *Mekoni* to *Mekelle* is mainly *Acacia* woodland. The forest area falls within five sub-districts of three districts found in and near the forest: *Alaje* (Ayba), *Hintalo-Wajirat* (Tsehafti) and *Raya-Azebo* (Tsigea, Ebo) (Figure 1).

3.1.2 Topography

Wejig-Mahgo-Waren massif forest is within the domain of the northern highlands of Ethiopia in the western escarpment of the rift valley. The topography includes hill areas, flat lands, mountains and valleys.

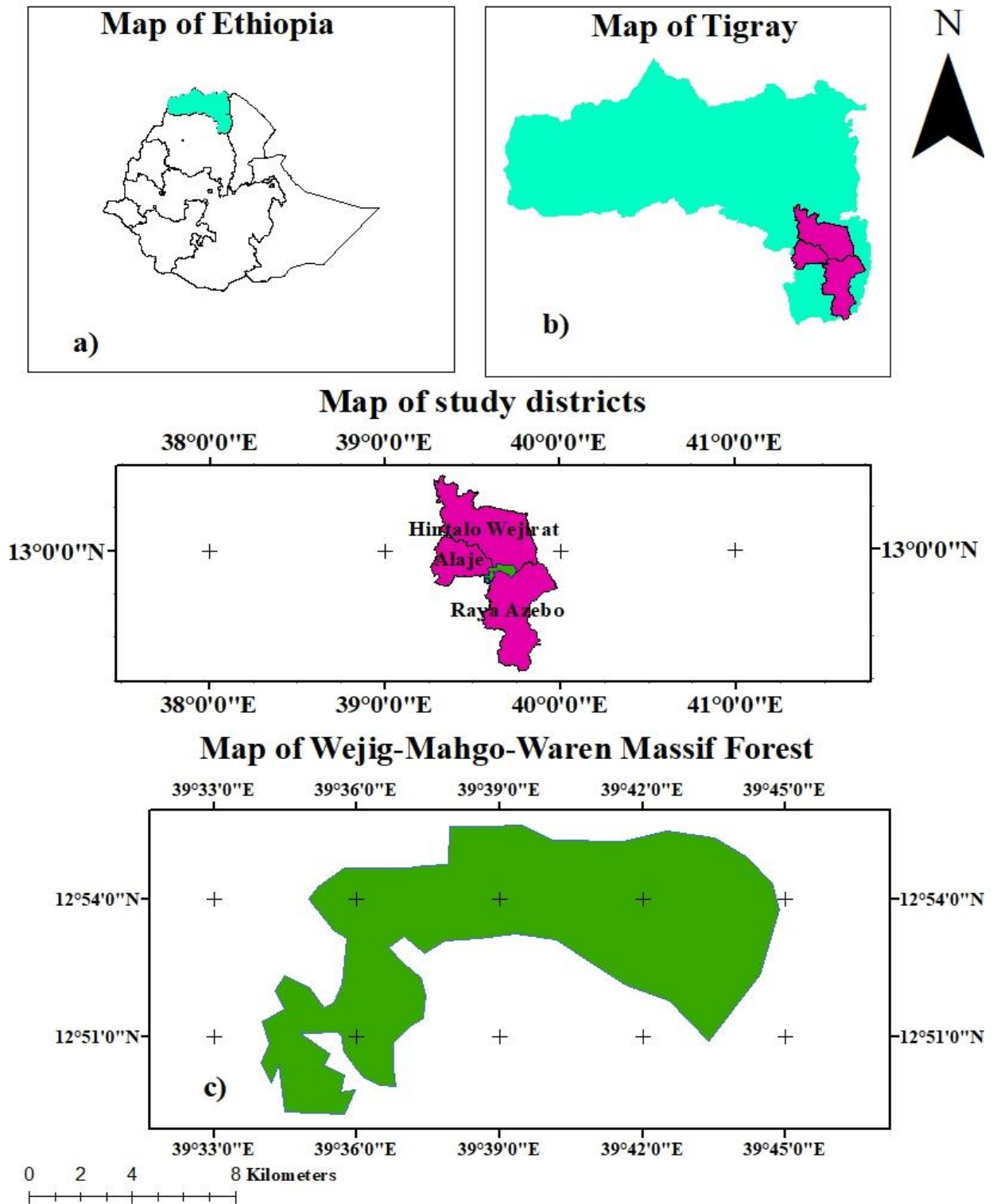
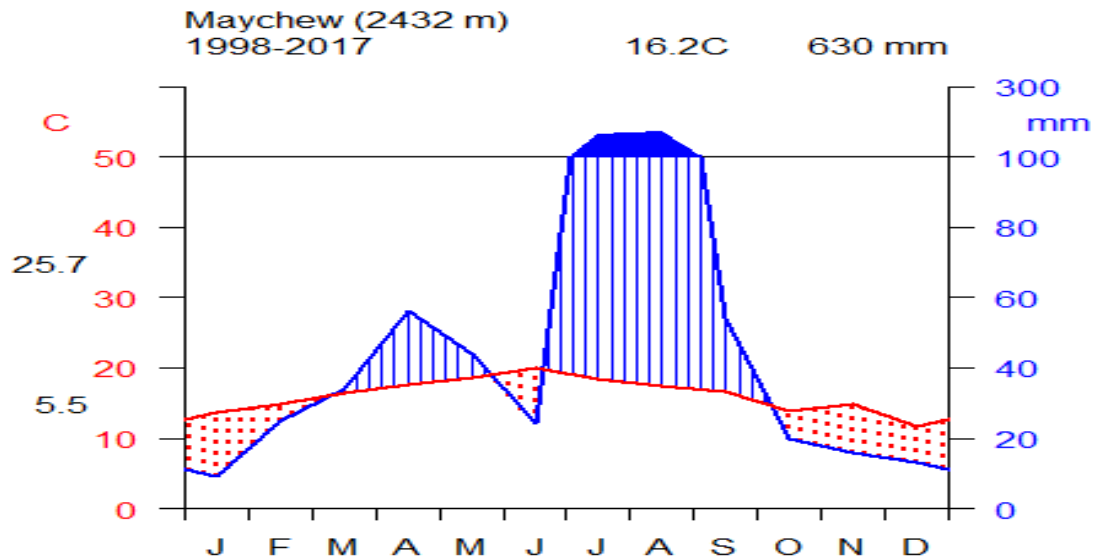


Figure 1. a) Map of Ethiopia, b) Tigray Regional State, c) The study areas (*Hintalo-Wejirat*, *Alaje* and *Raya-Azebo* districts), d) The forest area shaded green (developed using ArcGIS 10.4.1).

3.1.3 Rainfall and temperature (climate)

The rainfall and temperature data for this study were collected from Maychew, the nearest meteorological station. The data were collected from 1998-2017 by the National Meteorological Service Agency (NMSA, 2017). Climate diagram was computed by using R for window version 3.4.1 statistical package (R Development Core Team, 2017). The average maximum temperatures in the study area were recorded in June (25.68 °C) and May (24.90 °C). On the other hand, average minimum temperatures were observed in December (5.47 °C) and January (6.88 °C). According to the twenty years rainfall summarized data, the study area has a high rainfall distribution between July and August and a little bit between March, April and May. The mean annual rainfall of the study area is 630 mm (NMSA, 1998-2017). Generally, the study area has bimodal rainfall pattern, with low rainfall from March to May and the main rainy season from July to September (Figure 2).



Data Source: NMSA (1998-2017) data bank

Figure 2. Climatic diagram of Maychew town

3.1.4 Geology and soil

Wejig-Mahgo-Waren massif forest is formed on tertiary basalt, alkali-alluvial basalt and tuff. A number of transgressions of the sea in the Mesozoic era deposited sandstones (Adigrat Sandstone) and limestones (Antalo Limestone) in the Tigray Upland floristic region (TU) of northern Ethiopia (Friis *et al.*, 2010). They appear in the deep river valleys of the central part of the Western highlands, and are visible in the much eroded TU floristic region, where Enticho Sandstone, Adigrat Sandstone, Angula Shale and Antalo limestone are particularly prominent. Large parts of the undulating terrains in northern Ethiopia are characterized by shallow soils and frequent rock outcrops, while relatively thick soils are found along valley bottoms. Vertisols, Cambisols, Fluvisols, Regosols, and Leptosols are dominant soil types in the study area (Amanuel Zenebe *et al.*, 2015).

3.1.5 Vegetation

Based on the broad classification of the forests of Ethiopia (Friis *et al.*, 2010), *Wejig-Mahgo-Waren* massif forest is categorized as a dry evergreen Afromontane and grassland complex forest. It is characterized by dry climate with an annual mean precipitation less than 1000 mm and with *Juniperus procera* and *Olea europaea* subsp. *cuspidata* as dominant canopy trees. According to Friis (1992), this forest occurs on the highlands in the TU floristic regions at altitudes between (1600-) 2200 and 3200 (-3300) meters with annual rainfall between 500 and 1500 millimeters. Smaller trees and shrubs recorded from the northern forests include *Acokanthera schimperi* (dominant species), *Carissa spinarum*, *Clusia abyssinica*, *Discopodium penninervium*, *Euclea racemosa* subsp. *schimperi*, *Grewia ferruginea*, *Maesa lanceolata*, *Psydrax schimperianum*, *Teclea nobilis* and *Rhus natalensis*. There are comparatively few lianas, epiphytes, shrubs and

forest floor herbs, apart from some species of grasses. It conserves a number of wildlife species, including Leopard, Greater Kudu, Warthog, Ratel, Grey Duiker, Serval Cat, Wild Pig and Spotted Hyena (EWNHS, undated).

3.1.6 Demographics analysis

Based on the 2007 national census conducted by the Central Statistical Agency of Ethiopia (CSA, 2007), *Alaje*, *Hintalo-Wejirat* and *Raya-Azebo* districts have a total population of 107,972, 153,505 and 135,870, respectively. The population growth as compared with the 1994 national census, there is significant increment of population. Based on the 1994 national census conducted by the CSA, *Alaje*, *Hintalo-Wejirat* and *Raya-Azebo* districts had a total population of 83,692, 110,926 and 87,638, respectively. Languages spoken in *Alaje* districts are 98.78% (Tigrigna) and 0.96% (Agaw Kamyir), in *Raya-Azebo* 85.52% of the people speak Tigrigna, 11.04% Amharic, 1.83% Afaan Oromo and 1.53% Afarigna, while in *Hintalo-Wejirat* almost all speak Tigrigna as their first language (99.8%). Concerning education, *Alaje*, *Hintalo-Wejirat* and *Raya-Azebo* districts have literacy rates of 10.46%, 9.12% and 8.44%, respectively.

3.1.7 Economic activities

The community in the study area practice rain-fed subsistence mixed farming. All rural lands are collectively owned by the state. Distribution of land was done based on the fertility status of the soil (fertile, less fertile and moderately fertile), and the functional category of the plot (field plot and backyard plot). Concerning the land holding size in the area, it is estimated that a household owns 0.85 ha on average. Crops are produced in rain fed agriculture mainly for subsistence purpose. The major crops growing in the area are teff, maize, barley, wheat, beans, peas, lentils,

sorghum and pepper. Livestock husbandry is an integral part of the mixed farming system in the study area.

3.2 Methods

3.2.1 Ecological data collection

3.2.1.1 Vegetation sampling

A reconnaissance survey was made from 25 September to 10 October 2015 to get a general impression of the physiognomy of the forest and select sampling sites. Field trips were undertaken covering both dry and wet seasons from March 2016 to September 2017.

Vegetation data were collected from sample quadrats placed along transect lines, which were systematically laid at Southeast edge of forest part. Quadrats were laid along transect lines following systematic sampling approach as described by Mueller-Dombois and Ellenberg (1974). For trees and shrubs, 150 quadrats of 20 m x 20 m (400 m²) were laid at every 200 meters drop of elevation following transect lines (10 in total); the distance between two consecutive transects was 700 meters. For herbaceous species, sub quadrats of 1 m x 1 m at four corners and in the center of each quadrat were laid down. Transects were used because they are considerably important in description of vegetative change along an environmental gradient, or in relation to some marked feature of topography.

All plant species encountered in each quadrat were recorded and samples were collected. Vernacular (local) names were recorded when available. The habits of species were identified based on the conventions in the *Flora of Ethiopia* and *Eritrea*. For each quadrat, altitude,

longitude and latitude were measured using GPS and slope measured using clinometer. Grazing scale was classified visually as ‘no or slightly grazed (0)’, ‘intermediately grazed (1)’, ‘intensively grazed (2)’ and ‘over grazed (3)’ as used by Getinet Masresha (2014). State of human interference at each quadrat was estimated following Leul Kidane (2015). A 0-3 subjective scale was taken into consideration to record stumps, logs and signs of fuelwood collection. Therefore, the magnitude of impact was quantified as follows: 0=nil; 1= low; 2= moderate; and 3=heavy.

For population structure analysis, woody species having height greater than 2 m and circumference greater than 6 cm at breast height, i.e., 1.3 m were measured and counted. Circumference was later converted into a diameter. Diameter was calculated from circumference (C) using the formula $C = \pi d$, where d is diameter at breast height. Plant species were selected based on the following two or three criteria: higher frequency, higher relative abundance and higher basal area. For other woody and herbaceous plants, percentage cover-abundance was estimated and later converted into modified 1-9 Braun-Blanquet scale (van der Maarel, 1979; DCR-DNH, 2011). The scales are 1 \leq 0.1%, 2 = 0.1 to 1%, 3 = 1 to 2%, 4 = 2 to 5%, 5 = 5 to 10%, 6 = 10 to 25%, 7 = 25 to 50%, 8 = 50 to 75% and 9 >75%.

Plant specimens were brought to the National Herbarium (ETH) of Addis Ababa University for identification. Specimens were identified by referring to volumes of *Flora of Ethiopia* and *Eritrea* (1-8) and comparing with authenticated specimens at the National Herbarium and finally were deposited there with their labels.

3.2.2 Determination of vegetation regeneration status

For the purpose of the study “seedlings”, “saplings” and “mature trees/shrubs” were defined as plants with heights less than 1 m, 1-3 m and greater than 3 m, respectively. Following Tadesse Woldemariam (2003), five sub quadrats (3 m x 3 m) were laid at four corners and center of main sample quadrats to count sapling and seedling of all tree and shrub/tree species.

Densities of seedlings and saplings were considered as indicators of the regeneration potential of a forest and this was recorded as good, fair, and poor. A species is considered in a good regeneration state if the number of seedlings is greater than the number of saplings, which in turn is greater than that of trees (seedlings > saplings > trees). Fair regeneration is a condition of a species where the number of seedlings is greater than the number of saplings, and the number of saplings is also less than the number of trees (seedlings > saplings < tree). Poor regeneration is a status of regeneration where density of saplings is greater than that of seedlings (saplings may be <, > or = tree). A species is considered not regenerating if it survives only in adult stage. A species is considered as newly if it is presents only in the seedling stage (Khan *et al.*, 1987; Pokhriyal *et al.*, 2010). To apply regeneration analysis for conservation priority setting, tree and selected shrub species recorded in the study area were grouped into three categories, based on density of seedlings following Simon Shibru and Girma Balcha (2004) and Getinet Masresha (2014). Those species with no or one seedling were grouped under category I, whereas those species with seedlings between 2 and 14 were categorized under category II and species with seedling number greater than or equal to 15 were grouped under category III.

3.2.3 Soil sampling

Soil samples were collected to assess species richness, vertical distribution, composition, density in the soil seed bank and restoration potential of the study area. For soil samples, 20 m × 20 m quadrats (75 in total) were established at every 400 meters drop of altitude following transects; the distance between two consecutive transects was 700 meters. The samples were taken from five points covering 15 cm x 15 cm (one at the center and the other four at the corners). The reduction of quadrats number was because of manageability. A total of 150 soil samples were collected from the 2 separate soil layers, each layer had 5 cm depth (0-5 cm, 5-10 cm) following Price *et al.* (2010), Li *et al.* (2011) and Wang *et al.* (2015) using a digger. The probability of predation, rain-washing seeds from the surface resulted in poorly guided to conclude restoration potential of woody species from litter layer. Consequently, the litter layer was deliberately removed. Soil samplings were completed within one month (January 2017) to avoid differences in temporal bias in seed availability and composition following the method used by Toledo and Ramos (2011). The samples from each soil layer were used to determine variations of seed distribution at each depth of the soil layers (Mulugeta Lemenih and Demel Teketay, 2006). Soil samples from each layer were packed in to plastic bags and then it was transported to greenhouse of Addis Ababa University, Ethiopia.

Each soil sample was first passed through a 2 mm sieve to remove plant materials and gravels buried in the soil. Seeds with diameter > 2 mm were retrieved and then re-deposited in the final sieved soil samples (Dainou *et al.*, 2011). Several methods have been used to estimate the density and composition of soil seed banks and there has been much debate on the utility of the different approaches (Brown 1992; Bernhardt *et al.*, 2008). The ‘seedling emergence method’,

which involves spreading a thin layer of soil over a sterilized medium in a greenhouse and identifying and counting emerging seedlings, is commonly used (Ter Heerd *et al.*, 1996; Ter Heerd *et al.*, 1999). This method gives a good indication of the readily germinable seed bank, but does not provide a complete assessment of the seed bank flora unless the soil sample is maintained for extended periods at simulated habitat conditions (Thompson and Grime, 1979). Consequently, it was kept for about eleven months (February 2017 – December 2017). Indeed, the seedling emergence method can underestimate the density of the seed bank due to errors associated with seed dormancy and specific environmental requirements for germination (Brown, 1992; Bernhardt *et al.*, 2008; Wright and Clarke, 2009).

Soil samples remained after removal of debris and these were spread immediately in 20×20×6 cm rectangular plastic trays in Addis Ababa University greenhouse for germination of the seeds (Appendix 21). Each plastic tray was perforated at the bottom and plugged by cotton to facilitate proper drainage of water without losing soil. The seedling trays were kept continuously moist by daily watering following the method used by Toledo and Ramos (2011). The emerging seedlings were identified, counted and recorded. Unidentified seedlings were transplanted into additional germination trays for further growth until the species could be identified. After the newly germinated seedlings were identified and removed, the soil in each tray was thoroughly stirred to stimulate germination of remaining viable seeds from the deeper in the soil to come to the surface (Smith *et al.*, 2002). Ten seed trays filled with sterilized sand were kept under the same conditions as a control for seed contamination; no seedlings were found in these control trays. Each of the plant specimens were pressed, deposited in the National Herbarium (ETH) of Addis Ababa University. Photographs of seedlings were taken after they produced reproductive organs and became viable for identification.

3.2.4 Ethnobotanical data collection

3.2.4.1 Site selection

Selection of districts was carried out by considering sub-districts and villages that were found in the forest and nearby villages. The following were sub-districts found in and around the forest:

Ayba, Tsehafti, Ebo and Tsigea.

3.2.4.2 Sample size determination

The sample size for collecting quantitative data for this research to ensure the required representative sample size of households from the five sub-districts was determined using Cochran's (1977) formula as indicated by Bartlett et al. (2001) as follow:

$$n = \frac{N}{1 + N(e)^2}$$

n = sample size the research uses; N= total number of households in all five sub-districts

e= maximum variability or margin of error 5% (.05); 1= the probability of the event occurring.

According to sub-districts administrations, the total number of households in five villages of *Arera*, *Ago*, *Mahgo*, *Waren* and *Wejig* were 2236. Applying the above formula, the total sample size for general and key informants was determined to be 339 households. Accordingly, the number of households to be interviewed in each village was calculated using the proportion of the number of households in each village to the total number of households of the two sub-districts. Therefore, the sample size of *Arera* village (*Alaje* district) with a total household of 450

was 68 ($450 \times 339/2236 = 68$). Same calculations were employed for the other study sub-districts (Table 1).

Table 1. Number of households included in the study sub-districts and villages that were found around the forest

| District | Sub-district | Village | Total No. households | Key informants | General informants | Total informant |
|------------------------|--------------|----------|----------------------|----------------|--------------------|-----------------|
| <i>Alaje</i> | Ayba | Arera | 450 | 5 | 63 | 68 |
| | | Subtotal | 450 | 5 | 63 | 68 |
| <i>Hintalo-Wejirat</i> | Tsehafti | Ago | 463 | 5 | 66 | 71 |
| | | Waren | 386 | 4 | 54 | 58 |
| | | Subtotal | 849 | 9 | 120 | 129 |
| <i>Raya-Azebo</i> | Ebo | Mahgo | 418 | 8 | 56 | 64 |
| | | Wejig | 519 | 8 | 70 | 78 |
| | | Subtotal | 937 | 16 | 126 | 142 |
| Grand total | | | 2236 | 30 | 309 | 339 |

3.2.3.3 Selection of informants

The total number of informants involved in the ethnobotanical survey from the study sites were 339 (200 male and 139 female). Informants' ages ranged from 22-86 years (254 were < 50, and 85 were > 50 years old). Three hundred nine informants were selected based on gender stratified random sampling for collecting data about cognitive domain, use, management and threats of the forest during random visits made to houses in the study sub-districts. A total of 30 key informants were purposively selected with the help of local administrators, elders and other community members for ranking exercise and collection of specific ailments known by key informants. The key informants include elders, local experts in some domains and plant use

categories and knowledgeable men and women. Oral consent was obtained from each informant after informing the purpose of the study.

3.2.3.4 Semi-structured interview, guided field walks and focus group discussion

Ethnobotanical data were collected through semi-structured interview (Appendix 17), guided field walks and group discussion with informants on forest use, cognitive domain, management and level of threats following Martin (1995) and Cotton (1996). Guided field walks give the chance to observe and discuss signs of harvesting or patterns of plant distribution. It also gives local people the opportunity to observe and get to know the researchers who are new to the study area (Cunningham, 1996).

Five focus group discussions were conducted, one group from each village with purposely selected informants, and each group containing four to five informants. The groups contained village administrator, development agents for natural resource (DA), male households, female households and guards of the forest. This method was used to gain an insight in to the forest management systems and their suggestions for appropriate solutions to the current problems facing the forest. Interviews and discussions were conducted in the local language of the people in the districts, which is Tigrigna.

3.3 Data analysis

3.3.1 Species accumulation curve

Since number of species is highly dependent on sample size, comparing communities having different sample size is problematic (Peet, 1974; Magurran, 2004). Hence, to overcome this problem, all samples from different communities were standardized to a common sample size of the same number of individuals as recommended by Krebs (1999). Species accumulation curve is a statistical method for estimation of the number of species expected in a random sample of individuals taken from a collection (Krebs, 1999). Species accumulation curves have also been used to estimate the expected number of new species to be detected given a level of additional sampling effort, which can lead to efficient planning and sampling protocols (Krebs, 1999).

Species accumulation curves were plotted for the vascular plants recorded in *Wejig-Mahgo-Waren* massif forest. A graph was plotted for the cumulative number of species recorded as a function of sampling effort, i.e., the number of samples pooled. Species accumulation curve helps to illustrate the rate at which new species were included as the sampling effort proceed.

3.3.2 Plant community analysis

3.3.2.1 Cluster analysis

Classification by means of hierarchical cluster analysis is the most common multivariate technique to analyze community data. Cluster analysis helps to group a set of observations (here quadrats or vegetation samples) together based on their attributes or floristic similarities (Kent and Coker, 1992; McCune and Grace, 2002). Clustering is a multidimensional analysis that consists in partitioning the collection of sampling quadrats (Legendre and Legendre, 1998).

Accordingly, hierarchical cluster analysis was performed using R for window version 3.4.1 statistical package (R Development Core Team, 2017) to classify the vegetation into plant community types based on cover-abundance data of the species in each quadrat. The Similarity Ratio (SR) was measured using Ward's method. The data matrix contained 150 quadrats and 252 species collected from the sample quadrats. The SR was used because it eliminates the differences in total abundance among sample units and the Ward's method was used, because it minimizes the total within group mean of squares or residual sum of squares and is favorite as it produces nicely compact clusters (van Tongeren, 1995; McCune and Grace, 2002).

The three-column data, abundance or cover, were imported to R Package 3.4.3 (R Core Team, 2018) and matrified to carry out analysis for various parameters.

3.3.2.2 Ordination

To analyze the relationship between the plant communities and environment gradients, ordination method was employed. Leps and Smilauer (2001) proposed a simple rule to decide whether to use unimodal or linear method based on the length of the first Detrended Correspondence Analysis axis. If the length is > 4 , data are heterogeneous and unimodal methods should be used. If the length is < 3 , data are homogeneous and linear methods should be used. According to Zerihun Woldu (2017), the unimodal model is closer to reality of ecological data and is more suitable for heterogeneous data sets (structured by strong or long ecological gradients, with high species turnover and many zeroes in the species matrix).

Community environment relationship was analyzed with ordination program "Canonical Correspondence Analysis (CCA)" using R for window version 3.4.1 statistical package (R Development Core Team, 2017). CCA can be used to analyze unimodal relationships between

species and environmental variables. CCA performs quite well with skewed species distributions, with quantitative noise in species abundance data, with samples taken from unusual sampling designs, with highly intercorrelated environmental variables and with situations where not all of the factors determining species composition are known (Palmer, 1993). According to ter Braak and Šmilauer (1998), CCA is preferred for most ecological data sets (since unimodality is common). Moreover, the ordination diagram generated by CCA visualizes not only a pattern of community variation, but also the main features of the distributions of species along the environmental variables (ter Braak, 1987).

Before the CCA analysis, forward and backward stepwise selection of environmental variables was carried out to identify variables that could significantly explain the variability of species or plant community distributions. This assumes that the large majority of the explanatory variables may add little to the explanation since it is highly likely that many of them are collinear (significantly correlated to each other) (Graham, 2003; Dormann *et al.*, 2012; Zerihun Woldu, 2017). According to Dormann *et al.* (2012), there is some degree of collinearity between predictor variables in all real-world data.

3.3.3 Diversity indices

Shannon-Wiener's diversity index was computed to determine diversity and evenness of plant communities. It is given as:

$$H' = - \sum_{i=1}^s p_i \ln p_i$$

Where, H' = Shannon-Wiener Diversity Index; Σ = Summation symbol; p_i = the proportion of individuals or the abundance of i^{th} species expressed as a proportional of total cover in the sample and \ln = log base (natural logarithms).

Equitability or evenness index was calculated from the ratio of diversity observed to maximum diversity using the equation.

$$E = H' / \ln(S) = H' / H_{\text{max}}$$

Where, E = Evenness; H' = Shannon-Wiener Diversity Index; $H_{\text{max}} = \ln S$; S = total number of species in the study area. The value of evenness index falls between zero and one. The higher the value of evenness index, the more even the species is in their distribution within the given area.

Floristic similarity analysis between *Wejig-Mahgo-Waren* massif forest and eight other montane forests of Ethiopia was computed using;

Sorensen's similarity coefficient (S_s),

$$S_s = \frac{2a}{(2a + b + c)}$$

Where, a = number of species common to both forests compared; b = number of species in one forest; and c = number of species in the other forest (Kent and Coker, 1992).

3.3.4 Population structure

Diameter at breast height, basal area, tree density, height, frequency and important value index were used for description of vegetation structure.

3.3.4.1 Diameter at Breast Height (DBH)

Measurements of diameter were made at about 1.3 m from the ground using a measuring tape for those woody plants having circumferences greater than 6 cm. Plants with multiple stems below 1.3 m height were treated as a single individual and the DBH of all the stems were taken and then the average of the diameter was used for basal area calculation. If a tree was buttressed and abnormal at 1.3 m, the diameter was measured just above the buttress. Later the diameter were calculated from circumference (C) using the formula $C = \pi d$, where d is diameter at breast height. This technique is easy, quick, inexpensive and relatively accurate. There is direct relationship between DBH and basal area (Kent and Coker, 1992).

3.3.4.2 Basal area (BA) and relative dominance

Basal area is the area outline of a plant near ground surface. It is expressed in square m/hectare (Mueller-Dombois and Ellenberg, 1974). Basal area was used to calculate the dominance of species using the following formula:

$BA = \pi(d/2)^2$, where d is diameter at breast height.

$$\text{Relative dominance (RDo)} = \frac{\text{Total basal area of a species}}{\text{Total basal area of all tree species}} \times 100$$

3.3.4.3 Density (D)

Density is a count of the numbers of individuals of a species within a quadrat (Kent and Coker, 1992). It is closely related to abundance, but more useful in estimating the importance of a species. Counting is usually done in quadrats placed several times in vegetation communities

under study. Afterwards, the sum of individuals per species is calculated in terms of species density per convenient area unit, such as hectare (Mueller-Dombois and Ellenberg, 1974).

$$D = \frac{\text{The numebr of above ground stems of a species}}{\text{Sampled area in hectare}} \times 100$$

$$\text{Relative density} = \frac{\text{Number of ground stems of a species}}{\text{Total number of above stems in the sample area}} \times 100$$

3.3.4.4 Frequency (F)

Frequency is defined as the probability or chance of finding a species in a given sample area or quadrat. It is dependent on quadrat size, plant size and patterning in the vegetation (Kent and Coker, 1992).

It is calculated using the following formula:

$$F = \frac{\text{Number of plots in which a species occur}}{\text{Total number of plots}} \times 100$$

The frequencies of the tree species in all quadrats were computed. The higher the frequency, the more important the plant is in the community. Although a high frequency value means that the plant is widely distributed through the study area, the same is not necessarily true for a high abundance value. This abundance is not always an indicator of the importance of a plant in a community. Better ideas of the importance of a species with the frequency were obtained by comparing the frequency of occurrences of all of the tree species present. The result is called the relative frequency and is given by the formula:

$$\text{Relative frequency(RF)} = \frac{\text{Frequency of species}}{\text{Total frequency of all species}} \times 100$$

3.3.4.5 Importance Value Index (IVI)

Important value index (IVI) is useful to compare the ecological significance of species. It combines data from three parameters (relative frequency, relative density and relative dominance). It often reflects the extent of the dominance, occurrence and density of a given species in relation to other associated species in an area (Kent and Coker, 1992).

$$\text{IVI} = \text{RD} + \text{RF} + \text{RDO}$$

Where, RD is Relative Density, RF is Relative Frequency and RDO is Relative Dominance.

3.3.5 Analysis of soil seed bank

The species richness of soil seed bank in each soil profile were analyzed. Sorensen coefficient of similarity (Ss, Sorensen, 1948) was used to analyze the similarity between soil seed bank compositions with the standing vegetation and it is given as:

$$Ss = \frac{2a}{(2a + b + c)}$$

Where, a= species common to soil seed bank and standing vegetation; b = total species recorded in standing vegetation; c = total species recorded in soil seed bank.

The composition, frequency and density of seeds in the soil was determined from the germination data recorded in the green house.

3.3.6 Ethnobotanical data analysis

The data were summarized using Microsoft Office Excel 2010 computer programme and SPSS 20. Descriptive and Inferential (T-test) statistical methods were employed to analyze and summarize and test significance level of the ethnobotanical data. The relevant analytical tools of ethnobotany were used to determine the ranks, importance as level of benefits accumulated from various ethnocategories of plants.

3.3.6.1 Preference Ranking (PR)

Preference ranking is a system of ranking of lists or group of plants or other resources in order of first choice (Martin, 1995). This tool was used to determine the level of people's preference based on ranking most threatened plant species; most preferred medicinal plants used against febrile illness, most preferred wild edible plants and honeybee forage species. The selection of ailment for PR of medicinal plants was carried based on the highest number of medicinal plants recorded for healing. Therefore, informants were asked to list, score and rank plants based on their threats and uses stated above. A total of eight to fifteen informants were purposively selected for the collection of PR data.

3.3.6.2 Informant Consensus Factor (ICF) and Fidelity Level (FL)

The calculation of informant consensus factor (ICF) is used for testing homogeneity on the informant's knowledge in choosing certain medicinal plants against a given ailment group. The factor provides a range of 0 to 1, where a high value acts as a good indicator for a high rate of informant consensus. ICF was obtained by “number of use citation (reports) in each category

(n_{ur}) minus the number of species used (n_t), divided by the numbers of use citations (reports) in each category minus one” (Heinrich *et al.*, 1998).

$$ICF = \frac{n_{ur} - n_t}{n_{ur} - 1}$$

High ICF values are obtained when only one or a few plant species are reported to be used by a high proportion of informants to treat a particular ailment category, whereas low ICF values indicate that informants disagree over which plant to use. The main use of ICF is to select ailment categories where there is consensus on the use of plants among the informants and to identify species with particular importance in a culture. To use ICF, it is necessary to classify human illnesses into broad ailment categories (several ailments based on the organ systems in one category). The original categories used by Heinrich *et al.* (1998) and Ermias Leulekal (2014) include conditions common to eight ailment categories in the study area: (1) gastrointestinal, (2) dermatological, (3) external injuries, bleeding and snakebites, (4), evil eye, (5) febrile illness, (6) oral, dental and pharyngeal, (7) sensory and nervous system and (8) others.

The relative healing potential of each reported medicinal plant used against human ailments was evaluated using an index of fidelity level (FL) (Alexiades, 1996), given as:

$$FL = \frac{I_p}{I_u} \times 100$$

Where, I_p is the number of informants who independently cited the importance of a species for treating a particular ailment and I_u is the total number of informants who reported the plant for any given ailment. Independent T- test was employed to the test significance among different social groups on average number of medicinal plants.

3.3.6.3 Direct Matrix Ranking (DMR)

Direct matrix ranking is a system of ranking items in which instead of arranging a series of objects on one characteristic, informants were requested to order them by considering several attributes one at a time, i.e., it draws explicitly upon multiple dimensions (Martin, 1995). As it is described by Martin (1995), direct matrix ranking method was employed to score the use diversity and selected informants were asked to assign use values (5 = best, 4 = very good, 3 = good, 2 = less used, 1 = least used and 0 = not used) to each species. However, it was modified for efficient data recording as high, medium and low (no use) which corresponds to 1.0, 0.5 and 0.0, respectively and for convenience an average of the scores given by the informants were taken. Plants species selected for DMR were *Olea europaea* subsp. *cuspidata*, *Rhus glutinosa*, *Opuntia ficus-indica*, *Podocarpus falcatus*, *Becium grandiflorum*, *Carissa spinarum*, *Teclea nobilis*, *Pittosporum viridiflorum*, *Dodonaea angustifolia*, *Juniperus procera*. Most cited plant species were selected for DMR exercise. Eight informants were purposively selected for the collection of DMR data.

3.3.6.4 Cultural Significance Index (CSI)

The Cultural Significance Index (CSI), an anthropological approach presented by Turner (1988) and modified by Stoffle *et al.* (1990) and Silva *et al.* (2006), calculates importance through researcher- determined weighted ranking of multiple factors. Turner (1988) assigned scores on a five-point scale to the variables of quality and intensity of use and assigned a score of two, one, or 0.5 for the exclusivity or preference of use. To reduce the subjectivity of this approach, Silva *et al.* (2006) revised the CSI with a two-point scale for the variables. They also incorporated a consensus method called a correction factor to reduce the sensitivity of this method to sampling

intensity. The ethnographic, qualitative approach of the CSI method requires considerable experience and rapport with a cultural group for meaningful results.

Cultural Significance Index (Silva *et al.*, 2006) computed as follows:

$$CSI = \sum_{i=1}^n (i * e * c) * CF$$

Where, i = species management [non-managed (1) or managed (2)] e = Use Preference [not preferred (1) or preferred (2)] c = Use Frequency [rarely used (1) or used frequently (2)] CF = Correction factor [the number of informant citations for a given species divided by the number of informant citations for the most cited species].

CHAPTER FOUR

4. RESULTS

4.1 Vegetation study

4.1.1 Floristic diversity

A total of 264 species that belong to 162 genera and 82 families were identified (Appendix 1). Of these, 252 species were recorded in sampling quadrats and 12 species outside sampling quadrats. Angiosperms were represented by 254 species, gymnosperms by two species and Pteridophytes by eight species. The highest number of species were recorded for the families Asteraceae (27 species, 32.93%), Poaceae (24 species, 29.27%) and Fabaceae (23 species, 20.05%) followed by Lamiaceae (14 species, 17.07%), Solanaceae (12 species, 14.63%), Asclepiadaceae (9 species, 10.98%), Euphorbiaceae (7 species, 8.53%) and Amaranthaceae, Acanthaceae, Malvaceae and Scrophulariaceae (5 species, 6.09% each). Thirteen (about 4.92%) of species were endemic to Ethiopia (Appendix 2). Herbs were found to occur more abundantly (111 species, 42.04%) than shrubs (63 species, 23.86%), trees (57 species, 21.59%), lianas or woody climbers (19 species, 7.19%) and herbaceous climbers (14 species, 5.30%).

4.1.2 Plant community types

Five plant community types were identified from the hierarchical cluster analysis (Figure 3). Species with the highest synoptic values in the group were used to name the corresponding plant communities (Table 2).

Hierarchical clustering using SR with colored leaves and labels

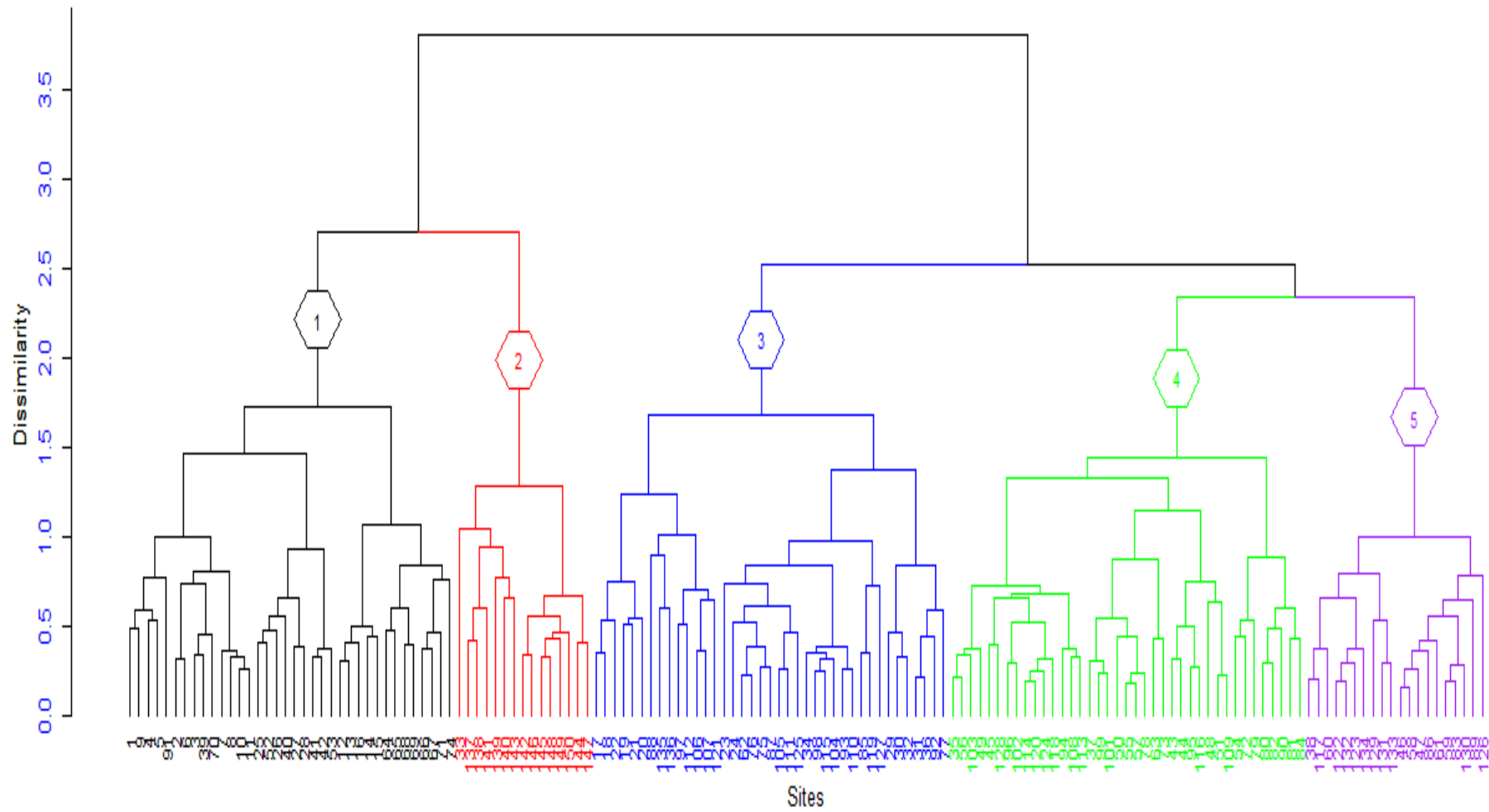


Figure 3. Dendrogram of the vegetation data from Agglomerative hierarchical cluster analysis using similarity ratio

Table 2. Synoptic cover abundance values of species in each community type. Values in **bold** refer to species used to name community types.

| Plant Community | 1 | 2 | 3 | 4 | 5 |
|--|-------------|-------------|-------------|------|------|
| Total quadrats number | 36 | 17 | 39 | 39 | 19 |
| <i>Cadia purpurea</i> | 4.61 | 1.11 | 2.32 | 0.21 | 2.80 |
| <i>Carissa spinarum</i> | 2.76 | 0.29 | 1.05 | 0.26 | 1.20 |
| <i>Calpurnia aurea</i> | 2.21 | 0.34 | 0.30 | 0.16 | 0 |
| <i>Celtis africana</i> | 1.84 | 0.32 | 0.14 | 0.21 | 0.20 |
| <i>Ficus sur</i> | 2.21 | 0.26 | 0.14 | 0.16 | 0 |
| <i>Podocarpus falcatus</i> | 2.71 | 0.61 | 0.07 | 0.32 | 0 |
| <i>Psydrax schimperiana</i> | 1.45 | 0.24 | 0.43 | 0.05 | 0 |
| <i>Pterolobium stellatum</i> | 2.26 | 0.08 | 0.20 | 0 | 0 |
| <i>Rhus natalensis</i> | 1.34 | 0.32 | 0.8 | 0.74 | 1.1 |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 2.87 | 4.84 | 3.14 | 2.58 | 1.9 |
| <i>Juniperus procera</i> | 1.71 | 4.32 | 1.93 | 4.21 | 0 |
| <i>Maytenus undata</i> | 0.05 | 1.13 | 0.11 | 0.53 | 0 |
| <i>Pittosporum viridiflorum</i> | 0.08 | 1.08 | 0.73 | 0.68 | 0 |
| <i>Dodonaea angustifolia</i> | 1.11 | 1.63 | 4.98 | 4.32 | 0 |
| <i>Acacia abyssinica</i> | 0.89 | 0.74 | 3.75 | 2.37 | 0 |
| <i>Becium grandiflorum</i> | 0.26 | 0.53 | 1.09 | 0.47 | 0 |
| <i>Maytenus arbutifolia</i> | 1.42 | 0.61 | 1.52 | 1.21 | 0 |
| <i>Osyris quadripartita</i> | 0.34 | 0.74 | 1.45 | 1.16 | 0 |

| | | | | | |
|------------------------------|------|------|------|-------------|-------------|
| <i>Rhus glutinosa</i> | 1.76 | 1.71 | 1.86 | 1.37 | 0 |
| <i>Erica arborea</i> | 0 | 0.32 | 0.57 | 6.53 | 0 |
| <i>Myrsine africana</i> | 0.03 | 1.84 | 2.39 | 2.95 | 0 |
| <i>Nuxia congesta</i> | 0.34 | 0.55 | 0.3 | 1.11 | 0.20 |
| <i>Acacia tortilis</i> | 0.29 | 0 | 0 | 0 | 5.40 |
| <i>Acacia etbaica</i> | 0.05 | 0 | 0 | 0 | 5.20 |
| <i>Acacia senegal</i> | 0.21 | 0 | 0 | 0 | 2.80 |
| <i>Dichrostachys cinerea</i> | 0.05 | 0 | 0 | 0 | 2.60 |
| <i>Acacia sieberiana</i> | 0 | 0 | 0 | 0 | 1.50 |
| <i>Balanites aegyptiaca</i> | 0 | 0 | 0 | 0 | 1.40 |
| <i>Grewia ferruginea</i> | 0.03 | 0 | 0 | 0 | 1.10 |
| <i>Grewia bicolor</i> | 0.16 | 0.11 | 0 | 0.11 | 1.10 |

1. *Cadia purpurea* – *Carissa spinarum* community

The *Cadia purpurea* – *Carissa spinarum* community is found on the edge of the forest. Severe livestock grazing and human impacts were observed. The altitudinal range of this community was from 2045 to 2481 m.a.s.l. The upper canopy was occupied by *Celtis africana*, *Podocarpus falcatus*, *Psydrax schimperiana*, *Bersama abyssinica* and *Ekebergia capensis*. The understory was dominated by *Cadia purpurea*, *Calpurnia aurea*, *Carissa spinarum*, *Acokanthera schimperi* and *Justicia schimperiana*. The shrubs-herbs layer includes *Achyranthes aspera*, *Bidens pilosa*, *Cynodon dactylon*, *Hypoestes forskoolii*, *Leucas abyssinica*, *Rumex nervosus*, *Solanum schimperianum*, *Tagetes minuta* and *Withania somnifera*. *Senecio hadiensis* is a climber.

The data obtained for plant species that produce non-timber forest products were from the ethnobotanical data analysis. This community also contained 45 species that produced non-timber forest products. The following are among the common species: *Achyranthes aspera*, *Clematis simensis*, *Acokanthera schimperi*, *Abutilon longicuspe*, *Aloe camperi*, *Bidens pilosa*, *Cadia purpurea*, *Calpurnia aurea*, *Carissa spinarum*, *Celtis africana*, *Ficus sur*, *Justicia schimperiana*, *Dovyalis verrucosa*, *Ekebergia capensis*, *Phytolacca dodecandra* and *Opuntia ficus-indica*.

2. *Olea europaea* subsp. *cuspidata* – *Juniperus procera* community

The altitudinal range of this community was from 2453 to 2719 m.a.s.l. The upper canopy was dominated by *Olea europaea* subsp. *cuspidata*, *Juniperus procera*, *Allophylus abyssinicus* and *Cassipourea malosana*. The understory layer was occupied by *Clerodendrum myricoides*, *Dovyalis abyssinica*, *Pavetta abyssinica* and *Pittosporum viridiflorum*. The shrub-herb layer was occupied by *Asparagus racemosus*, *Berberis holstii*, *Clutea abyssinica*, *Colutea abyssinica*, *Gerbera piloselloides*, *Smilax aspera*, *Periploca linearifolia* and *Helichrysum quartinianum*.

This community type was also rich in having 63 species that produced non-timber forest products. Some of the abundant species are the following: *Olea europaea* subsp. *cuspidata*, *Asparagus racemosus*, *Cassipourea malosana*, *Dombeya torrida*, *Dovyalis abyssinica*, *Eucalyptus globulus*, *Heliotropium strigosum*, *Juniperus procera*, *Kalanchoe petitiiana* and *Maytenus undata*.

3. *Dodonaea angustifolia* – *Acacia abyssinica* community

The altitudinal range of this community was from 2224 to 2640 m.a.s.l. The upper canopy was dominated by *Dodonaea angustifolia*, *Acacia abyssinica*, *Maytenus arbutifolia* and *Rhus*

glutinosa. The shrub-herb layer was occupied by *Becium grandiflorum* and *Gomphocarpus fruticosus*.

A total of 47 species that produced non-timber forest products were distributed in this community type. *Becium grandiflorum*, *Clerodendrum myricoides*, *Cyphostemma adenocaulis*, *Dodonaea angustifolia*, *Euphorbia petitiana*, *Myrica salicifolia*, *Rhus glutinosa* and *Rhus natalensis* were some of the most cited plant species.

4. *Erica arborea* – *Myrsine africana* community

The altitudinal range of this community was from 2497 to 2970 m.a.s.l. The upper canopy was dominated mainly by *Erica arborea*. The shrub-herb layer was occupied by *Myrsine africana*, *Anthospermum herbaceum*, *Epilobium hirsuta*, *Ferula communis*, *Hypericum revolutum* and *Smilax anceps*.

This community type also contained 36 species that produced non-timber forest products amongst the major species are the following: *Asparagus racemosus*, *Clusia lanceolata*, *Clerodendrum myricoides*, *Erica arborea*, *Laggera tomentosa*, *Maytenus arbutifolia* and *Nuxia congesta*.

5. *Acacia etbaica* – *Acacia tortilis* community

The altitudinal range of this community was from 1627 to 1830 m.a.s.l. The upper canopy was occupied by *Acacia etbaica*, *Acacia senegal*, *Acacia tortilis* and *Ziziphus mucronata* (Appendix 18). The shrub-herb layer was dominated by *Ziziphus spina-christi*, *Dichrostachys cinerea*, *Grewia bicolor*, *Chloris gayana*, *Cissus quadrangularis*, *Cynanchum gerrardii*, *Sporobolus pyramidalis* and *Verbascum sinaiticum*.

This community type also comprised 17 species that produced non-timber forest products amongst the major species are the following: *Verbascum sinaiticum*, *Acacia etbaica*, *Acacia senegal*, *Ziziphus mucronata*, *Euclea racemosa* subsp. *schimperi*, *Datura stramonium*, *Grewia mollis* and *Acokanthera schimperi*.

4.1.3 Species diversity

The Shannon-Wiener diversity index and evenness values of plants in the study area were 3.87 and 0.89, respectively. Among the communities, community two (*Olea europaea* subsp. *cuspidata* – *Juniperus procera*) had the highest diversity value (3.87), while evenness was high in community three (*Dodonaea angustifolia* – *Acacia abyssinica*) (0.89) (Table 3).

Table 3. Overall species richness, diversity and evenness values of plant communities

| Community types | Altitudinal | | Shannon-Wiener | |
|-----------------|-------------|----------------------|----------------------|------------------------|
| | range | Species richness (S) | Diversity Index (H') | Shannon's-Evenness (J) |
| 1 | 2045-2481 | 68 | 3.64 | 0.86 |
| 2 | 2453-2719 | 121 | 3.87 | 0.81 |
| 3 | 2224-2640 | 56 | 3.59 | 0.89 |
| 4 | 2497-2970 | 141 | 3.58 | 0.72 |
| 5 | 1627-1830 | 52 | 3.21 | 0.81 |

4.1.4 Ordination

Ordination with CCA of *Wejig-Mahgo-Waren* massif forest revealed the following relationship between community types and environmental factors. CCA1 reflects gradients of altitude and slope. Along CCA1 *Myrsine africana* - *Erica arborea* community type four found at relatively higher altitude or associated by altitude. CCA2 of the ordination diagram reflects livestock grazing and human impacts. Along CCA2, the differentiation is based on livestock grazing and human impact that was commonly found in *Acacia etbaica* – *Acacia tortilis* community type five (Figure 4 & 5).

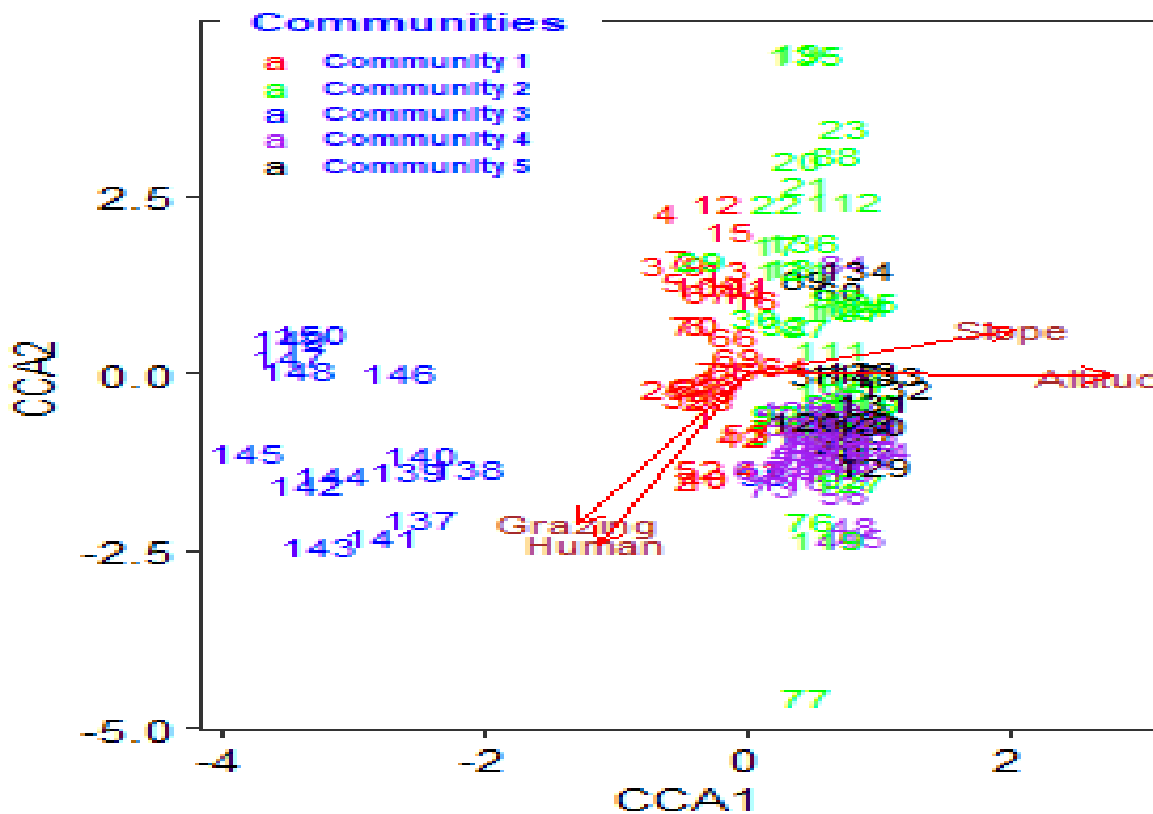


Figure 4. CCA displaying sites constrained by some selected environmental variables and community types identified by different colors

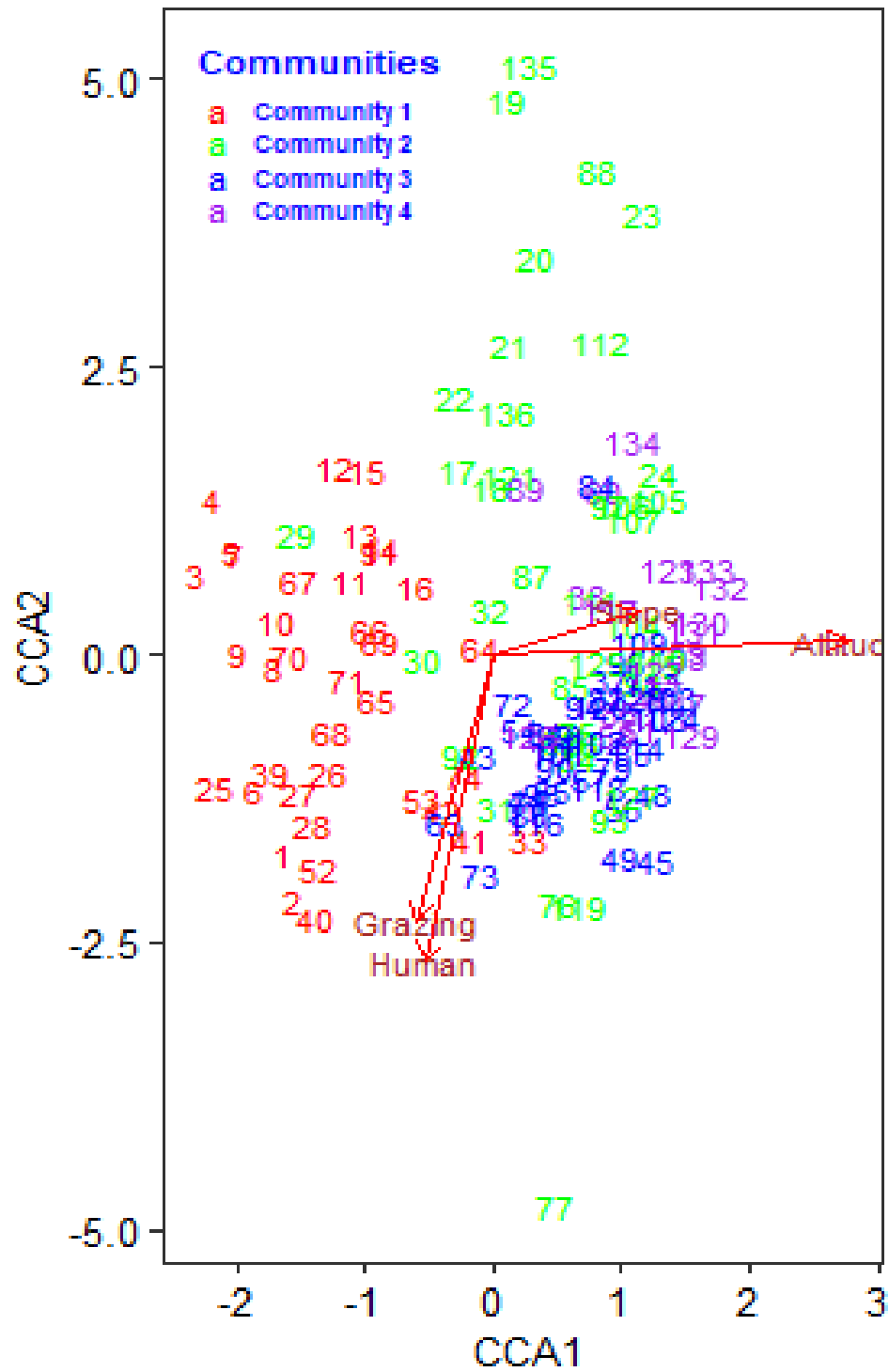


Figure 5. CCA displaying sites constrained by some selected environmental variables and community types identified by different colors

4.1.5 Structure of species population

The population structure of selected tree species followed four general diameter class distribution patterns (Figure 6). These are: A) **inverted-J-shaped**, which shows that the species frequency was highest in the lower diameter classes and decreased gradually towards the higher classes e.g. *Maytenus arbutifolia* B) **bell-shaped**, which shows a type of frequency distribution in which the number of individuals in the middle diameter classes was high and lower in the lower and higher diameter classes, e.g. *Cadia purpurea* C) **broken J-shaped**, which shows that there few individuals in the lower diameter class absent in the intermediate diameter class and in relatively higher number in the highest diameter class, e.g. *Bersama abyssinica* and D) **upward 'F'**, which shows that individuals concentrate in the lower (first) and intermediate (third and fourth) diameter classes. The rest of the classes had almost equally few numbers of individuals, e.g. *Myrsine africana*.

4.1.5.1 Density of some woody species

The higher selected woody plant species density per hectare (1351.50 ha⁻¹) was recorded. Densities of selected tree species per hectare are shown in (Table 4).

Table 4. Density of woody plants in the study area

| Scientific name | Density (ha⁻¹) |
|--|----------------------------------|
| <i>Podocarpus falcatus</i> | 25.50 |
| <i>Acacia abyssinica</i> | 121.67 |
| <i>Cadia purpurea</i> | 213.00 |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 202.00 |
| <i>Juniperus procera</i> | 143.67 |
| <i>Dodonaea angustifolia</i> | 243.33 |
| <i>Myrsine africana</i> | 141.00 |
| <i>Maytenus arbutifolia</i> | 104.33 |
| <i>Rhus glutinosa</i> | 117.67 |
| <i>Bersama abyssinica</i> | 39.33 |

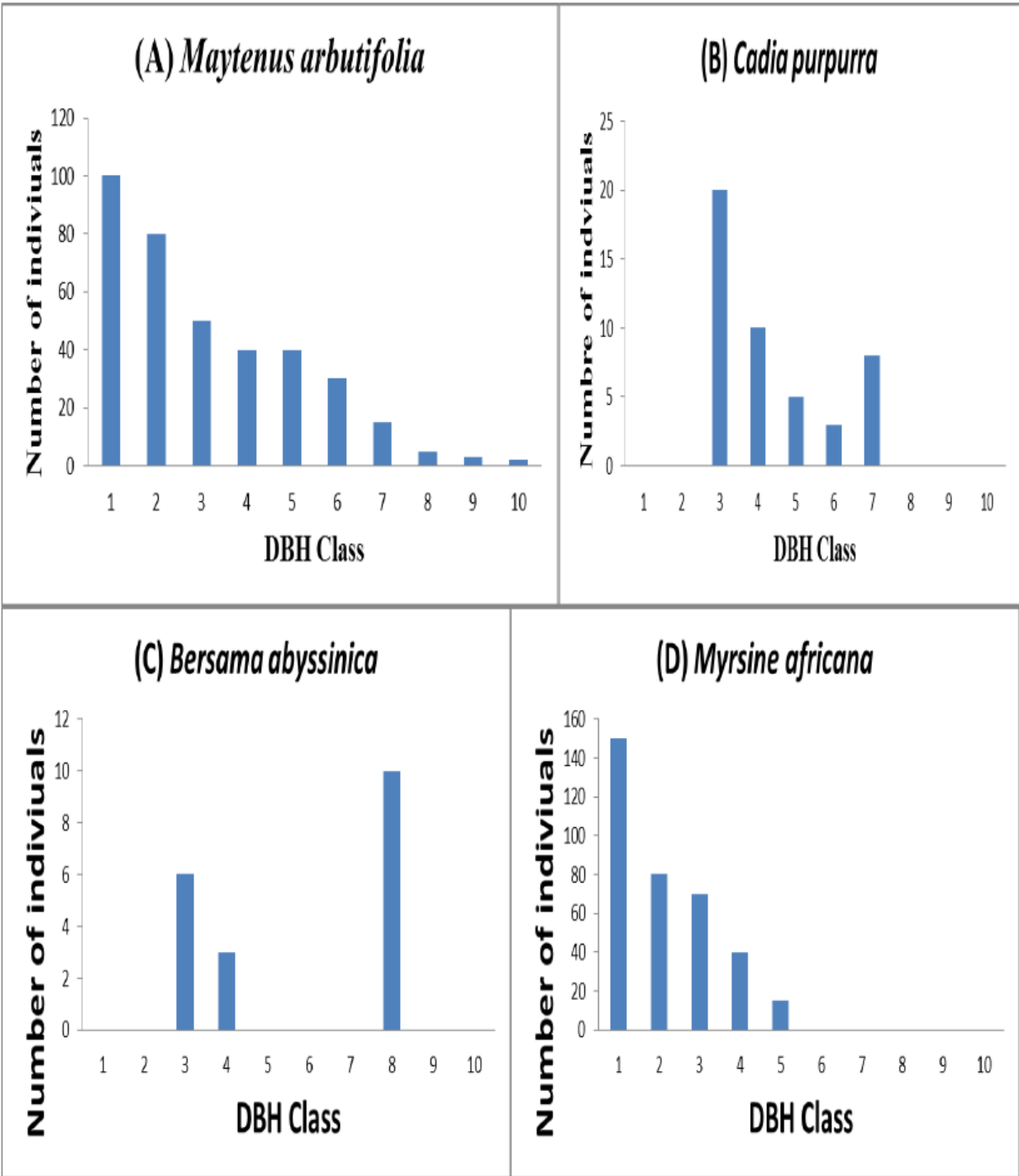


Figure 6. Four representative patterns of population structure based on DBH. DBH class: 1 = ≤ 2 cm, 2 = 2.01 – 7 cm, 3= 7.01 – 12 cm, 4 = 12.01 – 17 cm, 5 = 17.01 – 22 cm, 6 = 22.01 – 27 cm, 7 = 27.01 – 32 cm, 8 = 32.01 – 37 cm, 9 = 37.1 – 42 cm and 10 >42.01 cm.

4.1.5.2 Basal area

The total basal area was dominated by five woody species (57.45%), i.e., *Podocarpus falcatus*, *Dodonaea angustifolia*, *Olea europaea* subsp. *cuspidata*, *Juniperus procera* and *Myrsine africana* (Table 5).

Table 5. Basal area (BA) ($\text{m}^2 \text{ha}^{-1}$) of most dominant woody species

| Scientific name | BA ($\text{m}^2 \text{ha}^{-1}$) | Percent Contribution |
|--|------------------------------------|----------------------|
| <i>Podocarpus falcatus</i> | 0.95 | 13.41 |
| <i>Acacia abyssinica</i> | 0.30 | 4.23 |
| <i>Cadia purpurea</i> | 0.50 | 7.06 |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 0.82 | 11.58 |
| <i>Juniperus procera</i> | 0.73 | 10.31 |
| <i>Dodonaea angustifolia</i> | 0.92 | 12.99 |
| <i>Myrsine africana</i> | 0.72 | 10.16 |
| <i>Maytenus arbutifolia</i> | 0.30 | 4.23 |
| <i>Rhus glutinosa</i> | 0.21 | 2.97 |
| <i>Bersama abyssinica</i> | 0.21 | 2.97 |

4.1.5.3 Frequency of species

Olea europaea subsp. *cuspidata* was found to be the most frequent species occurring in 82% of all quadrats sampled. Species, including *Rhus glutinosa* (73%), *Maytenus arbutifolia* (67%) and *Juniperus procera* (65%) were also common. A list of the most frequent trees in the quadrats sampled is given in Table 6.

Table 6. Most-frequent trees and shrubs in the study area

| Scientific name | Frequency (%) |
|--|---------------|
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 82 |
| <i>Rhus glutinosa</i> | 73 |
| <i>Maytenus arbutifolia</i> | 67 |
| <i>Juniperus procera</i> | 65 |
| <i>Dodonaea angustifolia</i> | 63 |
| <i>Cadia purpurea</i> | 58 |
| <i>Acacia abyssinica</i> | 54 |
| <i>Myrsine africana</i> | 45 |
| <i>Bersama abyssinica</i> | 32 |
| <i>Podocarpus falcatus</i> | 23 |

4.1.5.4 Importance Value Indices

Importance Value Indices (IVI) were computed for trees and shrubs in the forest. The highest IVI values were obtained for *Dodonaea angustifolia* (36.75) and *Olea europaea* subsp. *cuspidata* (35.70) (Table 7).

Table 7. Importance Value Indices (IVI) trees and shrubs species (RD = Relative density, RF = Relative frequency, RDo = Relative dominance)

| Scientific name | RD | RF | RDo | IVI |
|--|------|-------|-------|-------|
| <i>Podocarpus falcatus</i> | 1.04 | 4.09 | 16.00 | 21.13 |
| <i>Acacia abyssinica</i> | 4.09 | 9.61 | 5.21 | 18.91 |
| <i>Cadia purpurea</i> | 8.65 | 10.32 | 8.69 | 27.66 |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 7.20 | 14.59 | 13.91 | 35.70 |
| <i>Juniperus procera</i> | 5.83 | 11.57 | 12.38 | 29.78 |
| <i>Dodonaea angustifolia</i> | 9.89 | 11.21 | 15.65 | 36.75 |
| <i>Myrsine africana</i> | 5.73 | 8.01 | 12.17 | 25.91 |
| <i>Maytenus arbutifolia</i> | 4.24 | 11.92 | 5.21 | 21.37 |
| <i>Rhus glutinosa</i> | 4.78 | 12.99 | 3.48 | 21.25 |
| <i>Bersama abyssinica</i> | 1.6 | 5.69 | 3.48 | 10.77 |

4.2 Height distribution pattern of woody species

The cumulative height class distribution pattern was an inverted J-shape, showing a decline in density of woody plants with increasing height classes (Figure 7). The abundance of individuals (78.13%) were found in the first, second and third height class (≤ 6 m). The rest of the height classes (i.e., class 4 to class 11) contributed only 22.87% of the total individuals (Figure 7).

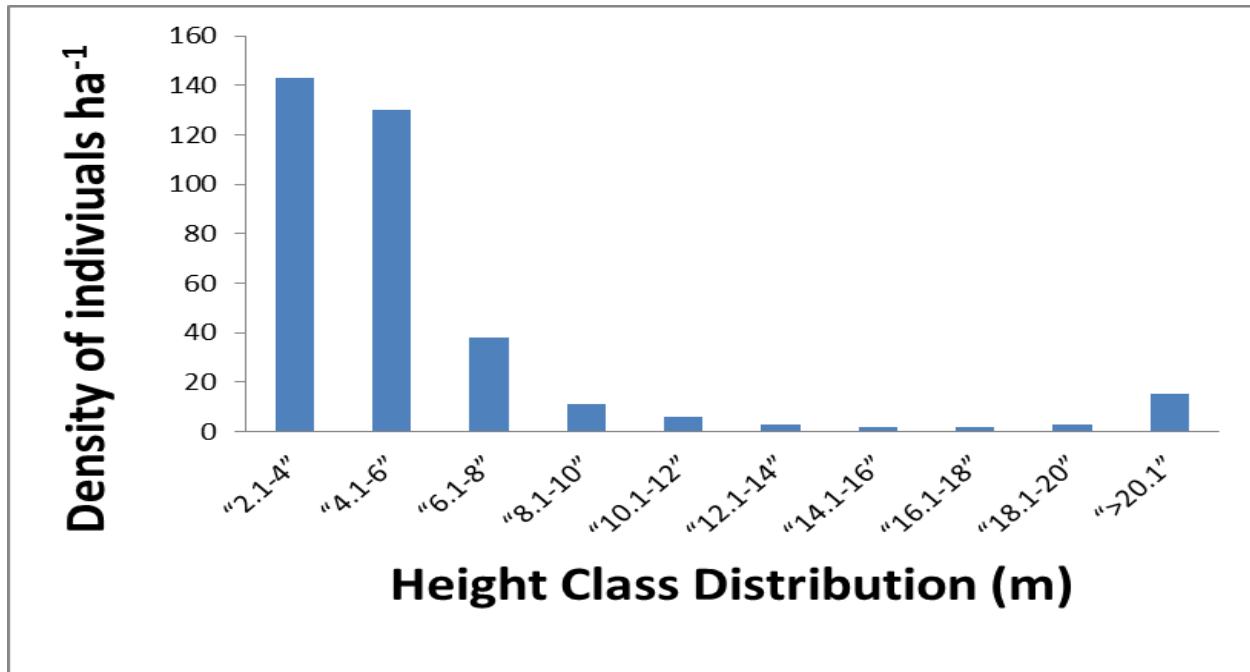


Figure 7. Height class distribution of woody species

4.3 Diameter at Breast Height distribution pattern of woody species

The density distribution of woody species by DBH class shows an inverted J – shaped pattern (Figure 8). For the whole data set, 30.92% of the individuals were found in class one (2.01 - 7 m) while class two (7.01 – 12 m) accounted for 25.77% (Figure 8). The greater percentage i.e., 38.50% were in classes 3 to 5. Individuals from the upper diameter class (greater than 27 m) contributed less than 9.73%. *Podocarpus falcatus*, *Olea europaea* subsp. *cuspidata*, *Juniperus procera*, *Ficus sur*, *Ekebergia capensis* and *Acacia abyssinica* were found to be the dominant large-sized trees in the forest, with DBH > 47.01 m.

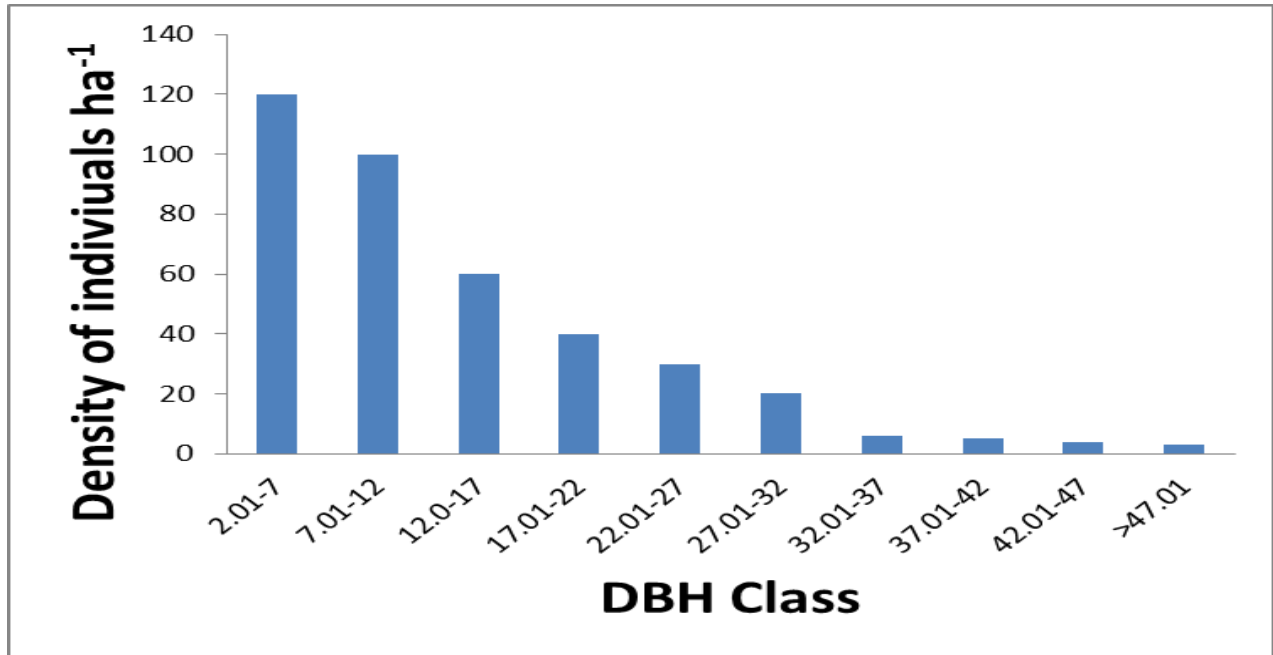


Figure 8. Cumulative frequency distribution by diameter class of woody species.

4.3 Regeneration status

A total of 301.00 (502 ha⁻¹) seedlings, 689.00 (1013.23 ha⁻¹) saplings and 571.00 (841.67 ha⁻¹) mature plants were recorded for all woody species (Figure 9).

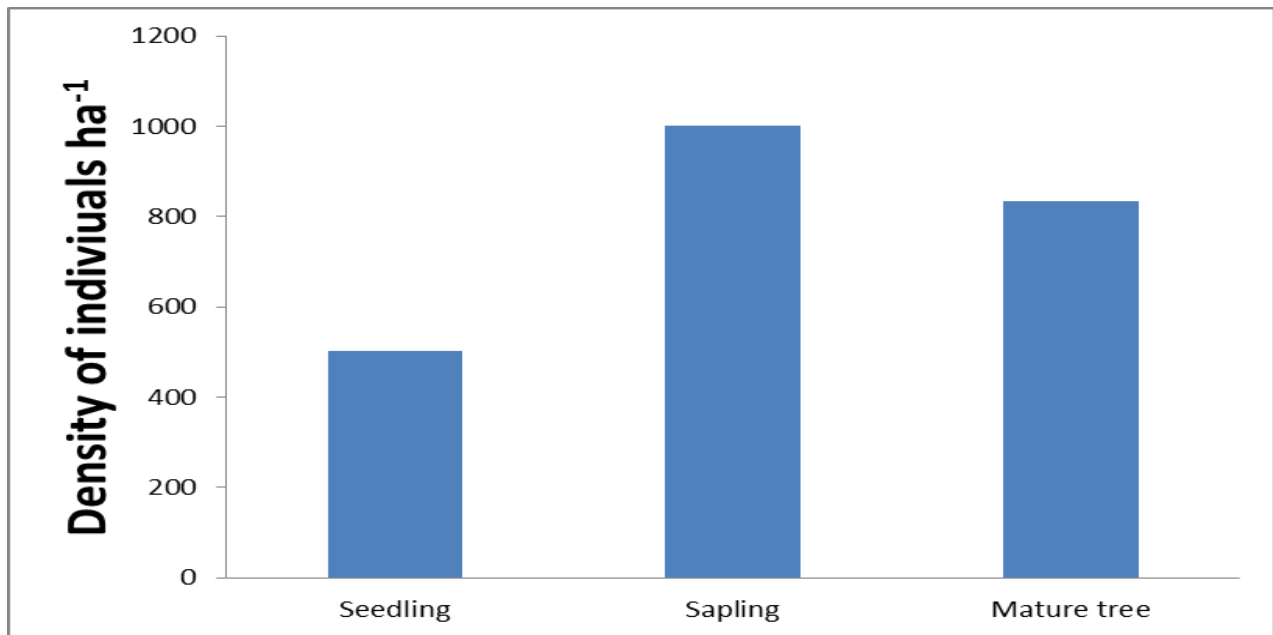


Figure 9. Diameter class distribution of woody species

To apply regeneration analysis for conservation priority setting, tree and selected shrub species recorded in the study area were grouped into three categories, based on the density of seedlings (Table 8). Therefore, those species with no or one seedling were grouped under category I, whereas those species with seedlings between 2 and 14 were categorized under category II and lastly species with seedling number greater than or equal to 15 were grouped under category III.

Table 8. Tree and shrub species categories for conservation priorities

| Regeneration status | | |
|-------------------------------|---------------------------------|--|
| Category I | Category II | Category III |
| Seedling density (0-1) | Seedling density (2-14) | Seedling density (≥ 15) |
| <i>Ekebergia capensis</i> | <i>Pittosporum viridiflorum</i> | <i>Olea europaea</i> subsp. <i>cuspidata</i> |
| <i>Ficus sur</i> | <i>Maytenus undata</i> | <i>Maytenus arbutifolia</i> |
| <i>Allophylus abyssinicus</i> | <i>Acokanthera schimperi</i> | <i>Cadia purpurea</i> |
| <i>Hagenia abyssinica</i> | <i>Bersama abyssinica</i> | <i>Calpurnia aurea</i> |
| <i>Dombeya torrida</i> | <i>Teclea nobilis</i> | <i>Erica arborea</i> |
| <i>Rubus steudneri</i> | <i>Osyris quadripartita</i> | <i>Myrsine africana</i> |
| <i>Balanites aegyptica</i> | <i>Rosa abyssinica</i> | <i>Acacia abyssinica</i> |
| <i>Cassipourea malosana</i> | <i>Celtis africana</i> | <i>Dodonaea angustifolia</i> |
| <i>Grewia bicolor</i> | <i>Nuxia congesta</i> | <i>Rhus glutinosa</i> |
| <i>Rhus retinorrhea</i> | <i>Podocarpus falcatus</i> | <i>Pterolobium stellatum</i> |
| | <i>Rhus natalensis</i> | |
| | <i>Dovyalis verrucosa</i> | |
| | <i>Psydrax schimperiana</i> | |

4.4 Floristic similarity among the study forest and some dry afro-montane forests

Wejig-Mahgo-Waren massif forest shared considerable number of species with *Dense*, *Hugumburda-Gratkahsu*, *Dess'a* and *Alemsaga* in decreasing order. Lower number of species was shared between the study area and *Tara Gedam-Abebaye*, *Dodola*, *Chencha* and *Peninsula-Zegie*. Output of a floristic similarity analysis between *Wejig-Mahgo-Waren* massif forest and some other Ethiopian montane forests, based on Sorensen's coefficient of similarity analysis was summarized in Table 9.

Table 9. Composition of floristic similarities among *Wejig-Mahgo-Waren* massif forest and some dry afro-montane forests in Ethiopia (N= Number of species included in comparison; a= species composition to both forests compared; b= species available only in the study area; c= species available only in the other forest compared; SS = Sorensen's coefficient of similarity).

| Forests | Elevations range (m.a.s.l) | N | a | b | c | SS |
|---------------------------------------|-------------------------------|-----|----|-----|-----|------|
| Alemsaga ^a | 1970-4135 | 124 | 57 | 207 | 67 | 0.30 |
| Chencha ^b | 1180-2250 | 174 | 45 | 219 | 129 | 0.21 |
| Dense ^c | 2125-2921 | 158 | 78 | 186 | 80 | 0.37 |
| Dess'a ^d | 1000-2760 | 130 | 65 | 199 | 65 | 0.33 |
| Dodola ^e | 2500-3500 | 113 | 46 | 218 | 67 | 0.24 |
| Hugumburda- Gratkahsu ^f | 1560-2688 | 102 | 72 | 192 | 30 | 0.39 |
| Peninsula-Zegie ^g | 1770-1975 | 113 | 34 | 230 | 79 | 0.18 |
| Tara Gedam- | 2142-2484 | 143 | 47 | 217 | 96 | 0.23 |

Abebaye^h

Wejig-Mahgo- 1627-2970 264 264 0 0 1

Waren

Data source used for comparison: ^aGetinet Masresha *et al.* (2015), ^bDesalegn Wana and Zerihun Woldu (2005), ^cErmias Leulekal (2014), ^dErmias Aynekulu (2011), ^eKitessa Hundera *et al.* (2007), ^fLeul Kidane *et al.* (2010), ^gAlemnew Alelign *et al.* (2007), ^hHaileab Zegeye *et al.* (2011).

4.5 Soil seed bank study

4.5.1 Species composition

The total number of species recorded was 54, representing 42 genera and 23 families from soil samples collected (Appendix 3). Of the identified species recorded from the forest, 85.16% were herbs and 14.84% were woody plants. Five families, namely Poaceae (380 seedlings), Asteraceae (361 seedlings), Chenopodiaceae (68 seedlings), Brassicaceae (53 seedlings) and Amaranthaceae (46 seedlings) accounted for 81.43 percent of these seedlings. The dominant species in the seed bank were herbs and the proportion of woody species was very low.

4.5.2 Densities of seeds

The number of viable seeds in the soil samples, from germination corresponded to a seed bank density down to 10 cm in the soil was 1115 m⁻². The six species with the highest soil seed densities in descending order include *Tagetes minuta*, *Digitaria velutina*, *Galinsoga parviflora*, *Eragrostis cilianensis*, *Erucastrum abyssinicum* and *Chenopodium murale*. The most frequent

species in the soil seed banks were *Chenopodium murale*, *Tagetes minuta*, *Galinsoga parviflora*, *Digitaria velutina* and *Eragrostis cilianensis* (Table 10).

Table 10. Density and frequency of species from soil samples collected from the study area

| Scientific name | Density (%) | Frequency (%) |
|--------------------------------|--------------------|----------------------|
| <i>Amaranthus hybridus</i> | 1.70 | 9.00 |
| <i>Chenopodium murale</i> | 4.80 | 43.00 |
| <i>Chenopodium opulifolium</i> | 1.10 | 18.00 |
| <i>Digitaria velutina</i> | 16.14 | 39.00 |
| <i>Eragrostis cilianensis</i> | 7.35 | 27.00 |
| <i>Erucastrum abyssinicum</i> | 4.75 | 26.00 |
| <i>Galinsoga parviflora</i> | 11.12 | 41.00 |
| <i>Oxalis procumbens</i> | 2.20 | 20.00 |
| <i>Solanum nigrum</i> | 00.99 | 14.00 |
| <i>Tagetes minuta</i> | 22.87 | 41.00 |

4.5.3 Vertical distribution of seeds

The depth distribution of the seed bank was more or less consistent with the highest densities in the upper five centimeters of soil and decreasing densities in the lower centimeters. The soil seed bank density exhibited a declining trend with increasing soil depth accounting for 668 seeds m⁻² (0-5 cm) and 447 seeds m⁻² (5-10 cm) (Figure 10).

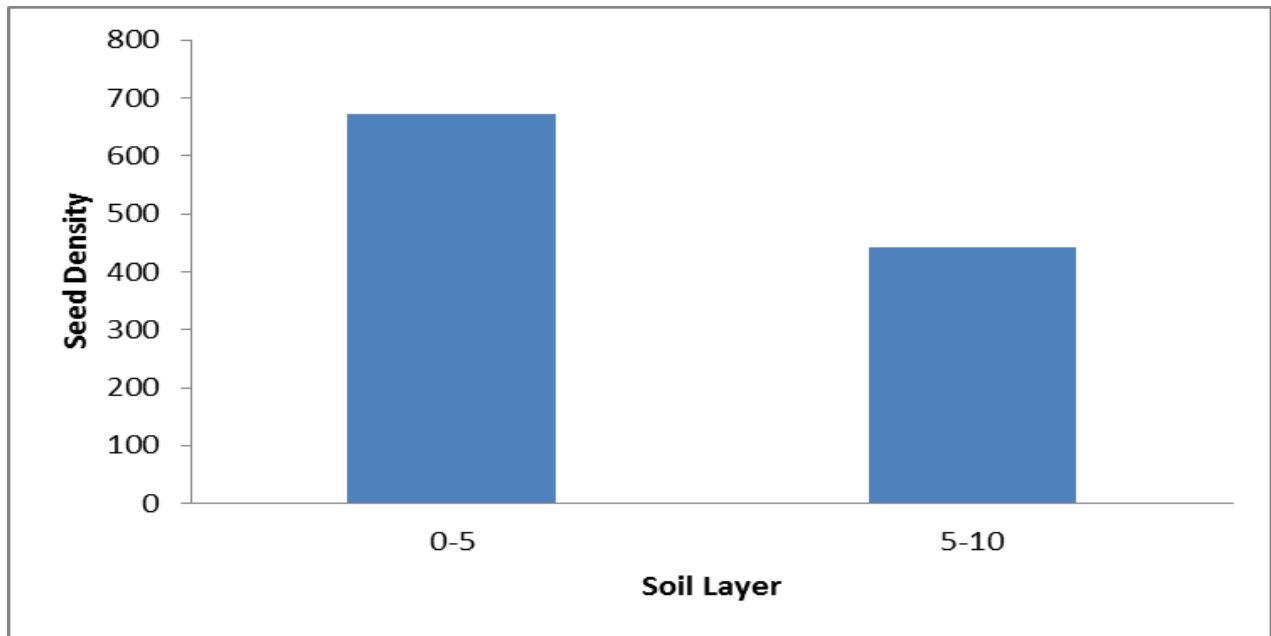


Figure 10. Depth distribution of species density recorded from soil samples

There was variation in relation to depth distribution that decreased to the lower centimeters for the sampled species (Table 11 and Table 12).

Table 11. Ten selected species with their depth distribution of seeds

| Layer | Density of seeds m ⁻² | | | | | | | | |
|---------|----------------------------------|----|-----|----|----|-----|----|----|-----|
| | AH | CM | DV | EC | EA | GP | OP | SN | TM |
| 0-5 cm | 18 | 34 | 106 | 67 | 42 | 64 | 19 | 7 | 136 |
| 5-10 cm | 1 | 24 | 76 | 15 | 10 | 60 | 8 | 5 | 123 |
| Total | 19 | 58 | 182 | 82 | 52 | 124 | 27 | 12 | 259 |

(Abbreviations: AM = *Amaranthus hybridus*; CM = *Chenopodium murale*; DV = *Digitaria velutina*; EC = *Eragrostis cilianensis*; EA = *Erucastrum abyssinicum*; GP = *Galinsoga parviflora*; OP = *Oxalis procumbens*; SN = *Solanum nigrum*; TM = *Tagetes minuta*)

Table 12. Densities of seeds of identified woody species

| Layer | Density of seeds m ⁻² | | | | | | | |
|-------|----------------------------------|----|----|----|----|----|----|----|
| | BG | CP | DA | FP | OL | RN | SA | VS |
| 0-5 | 2 | 1 | 2 | 9 | 4 | 1 | 2 | 2 |
| 5-10 | 0 | 0 | 0 | 1 | 2 | 1 | 0 | 0 |
| Total | 2 | 1 | 2 | 10 | 6 | 2 | 2 | 2 |

(Abbreviations: BG = *Becium grandiflorum*; CP = *Cadia purpurea*; DA = *Dodonaea angustifolia*; FP = *Ficus palmata*; OL = *Ocimum lamifolium*; RN = *Rumex nervosus*; SA = *Solanum adoense*; VS = *Vernonia* sp.)

4.5.4 Similarity between soil seed bank and standing vegetation

Species recorded only in the standing vegetation were 228, Species recorded only in soil seed bank were 18 and species recorded both in soil seed bank and standing vegetation were 36. The Sorensen Similarity coefficient was 23%. Eight woody species (14.84%) were encountered in the soil seed bank. 6.67% soil seed bank woody species were also found in the standing woody species. The similarity of woody species composition found in the seed bank and standing vegetation was low. Woody species found in soil seed bank were 100% represented in the standing vegetation.

4.6 Plants and People interaction

4.6.1 Concept about the forest and forest resources

Informants reported that forest is important to attract rain and to serve as home to wildlife. They also reported its importance for beekeeping and foraging and as source of straw for thatching and wood used as fuelwood. It is possible to use resources properly from the forest with the permission of the district administration at any time as long as it did not deplete later.

Emic categorizations of plants in the study area were practiced. Plants were described in physiognomic and generic terms. Based on growth form, individual plants were locally described as “*O'm*” (tree), “*Kotkoat*” (bush), “*Hareg*” (liana), “*Sa'eri*” (grass) and “*Tsahyai*” (weed).

4.6.2 Management of forest plant resources

The local communities have had indigenous knowledge of managing forest resources by their own rules. They nominated “*kessasi*” (accuser) and “*wardia*” (guards) and collected money used for buying ox or goat; slaughtered the ox or goat. Guidelines prepared to the guards how they would protect the forest. Punishment conducted when human enhanced dieback and illegal cutting of plant species have manifested. Moreover, there was a taboo related to *Olea europaea* subsp. *cuspidata* that illegal cutting encountered, it was considered as person was killed. The forest was used to oxen grazing from October to December. Currently, the regional government hired guards having monthly salary.

4.6.3 Major plant use categories

The local people around the study area had accumulated traditional knowledge on the use categorization of forest plant resources into wild edible plants, medicinal plants, farm implements, building materials, fuel wood, livestock fodder/ forage and honeybee forage. The study recorded 79 plant species belonging to 70 genera and 47 families. The families best represented were Fabaceae (7 species), Asteraceae (5 species), Lamiaceae, Solanaceae (4 species each) and Malvaceae (3 species). Thirteen families were represented by two species each and 30 families were represented by one species each.

Traditionally, local people in the study area had their own way of categorizing important plant species in the forest based on the value they later provided. Accordingly, from the 79 plant species, 52 (66%) were used as traditional medicine (human and livestock), 28 (35%) as livestock fodder/ forage and 27 (34%) as honeybee forage (Table 13).

Table 13. The seven major use categories of plants in the study area identified by local communities.

| Use category | Plant category | | | % of total species | Rank |
|--------------------------|----------------|--------|---------|--------------------|-----------------|
| | Family | Genera | Species | | |
| Traditional medicine | 35 | 49 | 52 | 66 | 1 st |
| Livestock fodder/ forage | 20 | 24 | 28 | 35 | 2 nd |
| Honeybee forage | 20 | 25 | 27 | 34 | 3 rd |
| Fuel wood | 19 | 20 | 23 | 29 | 4 th |
| Farm implements | 19 | 20 | 22 | 28 | 5 th |
| Building materials | 16 | 17 | 19 | 24 | 6 th |
| Wild edible plants | 12 | 14 | 17 | 22 | 7 th |
| Total | 47 | 70 | 79 | 100 | |

Table 13 shows that the forest was primarily used as source of traditional medicine for both livestock and human and forage for the livestock and honeybees. Species cited for each category are listed in Appendices 13, 14, 15, 16, 17 and 18.

4.6.3.1 Medicinal plants collected from the study area

Fifty-two plant species were reported to be used against human and/or livestock ailments in the study area, of which eight were used against both human and livestock ailments. Growth habits of the medicinal plants included shrubs (23 species, 44.23%), trees (12 species, 25.00%), herbs (11 species, 21.15%), woody climbers (2 species, 3.84%) and herbaceous climbers (3 species, 5.77%).

4.6.3.1.1 Ailments treated

The reported medicinal plants were used to treat 50 human and 5 livestock ailments. Of the total number of medicinal plants, 45 (68.18%) were used to treat human ailments (Appendix 4), 13 (19.69%) were employed to treat ailments of livestock (Appendix 5) and 8 (12.12%) were used against ailments of both human and livestock ailments (Appendix 6). With regard to human ailments, febrile illness was the one, against which, a high number of medicinal plants (6 species) were prescribed, followed by evil eye (5 species), jaundice (4 species) and wound (4 species). Cancer, cholera, malaria, devil and snakebite were treated by 3 species each. Both eye infection and dislocation were the prevailing livestock ailments in the study area against each three species were used, followed by those (2 species) used against infestation by leeches. Based on the reports of the interview, a single plant could be used for treatment of different ailments as stated in Table 14.

Table 14. Plants with five or more medicinal uses as identified by the local people

| Scientific name | Ailments treated |
|--|---|
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | Febrile illness, Cholera, Typhus, Shanbu*, Fnfan*, Eye Infection, Stomachache |
| <i>Balanites aegyptica</i> | Cholera, Gulhay*, Hangnail, Dandruff, Anthrax, Tonsillitis, |
| <i>Clematis simensis</i> | Chibiti*, Cancer, Kidney Ailment, Dislocation, Paralysis, |
| <i>Datura stramonium</i> | Hemorrhoid, Tuberculosis, Werchi*, Wound, Toothache |
| <i>Withania somnifera</i> | Malaria, Satan, Wound, Febrile illness, Dislocation |

*It refers to local name of an ailment that did not come across common name.

4.6.3.1.2 Plant parts used and mode of preparation

Interview results given in Table 15 shows that the leaf is the most commonly used plant part accounting for 59.04% of the total reported medicinal plants, followed by roots (22.86%) and leaf/root (8.57%).

Table 15. Plant parts used for traditional medicine preparations

| Plant part | Total response | Percentage |
|--------------|----------------|------------|
| Leaf | 62 | 59.04 |
| Root | 24 | 22.86 |
| Leaf or Root | 9 | 8.57 |
| Bark | 4 | 3.81 |

| | | |
|--------------|-----|-------|
| Young shoot | 2 | 1.90 |
| Latex | 1 | 0.95 |
| Fruit | 1 | 0.95 |
| Whole plant | 1 | 0.95 |
| Leaf or bark | 1 | 0.95 |
| Total | 105 | 99.98 |

It was found (Table 16) out that most remedies were processed by crushing (83.33%), chewing (9.26%) and boiling (4.63%) and the majority (68%) of the remedies were prepared from fresh materials only. Some (24%) remedies were prepared from either dried or fresh materials while few (8%) were prepared from dried materials only. Considerable preparations were made from mixture of different plant species with the use of water and different additives, such as honey, butter, water and cheese. The additives were believed to reduce poisonings, improve flavor and mitigate other side effects.

Table 16. Preparation methods of traditional medicine

| Form of preparation | Total response | Percentage |
|----------------------------|-----------------------|-------------------|
| Crushing | 90 | 83.33 |
| Chewing | 10 | 9.26 |
| Boiling | 5 | 4.63 |
| Others | 3 | 2.78 |
| Total | 108 | 100 |

4.6.3.1.3 Modes of administration, dosage and use of antidotes

Informants indicated that drinking was the most employed mode of administration with 41 (34.75%) reports, followed by tying 28 (23.73%), smoking 15(12.71) and washing 11 (9.32%). Others were administered by rubbing on the affected/infected part and through swallowing (11.01%) (Table 17). Results show that there was no agreement among informants in materials used for measurement or units used. Most informants used cup, spoon, drops and fingers to determine doses and different doses were reported even for the same or similar ailments. Most of the remedies were reported to have no adverse effects except for some species, such as *Phytolacca dodecandra*, *Justicia schimperiana*, *Clematis simensis* and *Solanum marginatum* that were indicated to be poisonous both to human and livestock. Milk, honey, yoghurt and butter were the commonly reported antidotes in case of adverse side effects.

Table 17. Method of administration of traditional medicine

| Administration | Total response | Percentage |
|-----------------------|-----------------------|-------------------|
| Drinking | 41 | 34.75 |
| Tying | 28 | 23.73 |
| Smoking | 15 | 12.71 |
| Washing | 11 | 9.32 |
| Rubbing | 9 | 7.63 |
| Swallowing | 4 | 3.38 |
| Smearing | 5 | 4.24 |
| Others | 5 | 4.24 |
| Total | 118 | 100 |

4.6.3.1.4 The most-preferred plants for treating febrile illness

Preference ranking exercise on six medicinal plants that were reported to be used against febrile illness revealed *Eucalyptus globulus* as the best-preferred species. *Withania somnifera* and *Olea europaea* subsp. *cuspidata* were also among the most-preferred species ranked 2nd and 3rd, respectively and others followed (Table 18).

Table 18. Preference ranking to medicinal plants used to treating febrile illness

| Medicinal plants | Informants labeled A to O | | | | | | | | | | | | | | | Total score | Rank |
|---|---------------------------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|-------------|-----------------|
| | A | B | C | D | E | F | G | H | I | J | K | L | M | N | O | | |
| <i>Eucalyptus globulus</i> | 6 | 6 | 6 | 6 | 5 | 6 | 5 | 6 | 6 | 6 | 6 | 5 | 6 | 6 | 6 | 87 | 1 st |
| <i>Withania somnifera</i> | 5 | 5 | 5 | 5 | 6 | 4 | 6 | 5 | 5 | 4 | 5 | 6 | 5 | 5 | 5 | 76 | 2 nd |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 2 | 4 | 4 | 4 | 4 | 2 | 3 | 4 | 2 | 5 | 4 | 3 | 3 | 4 | 4 | 52 | 3 rd |
| <i>Zehneria scabra</i> | 3 | 3 | 3 | 1 | | 5 | 4 | 3 | 4 | 3 | 3 | 4 | 4 | 3 | 3 | 46 | 4 th |
| <i>Becium grandiflorum</i> | 4 | 2 | 2 | 2 | 1 | 3 | 1 | 1 | 3 | 1 | 1 | 2 | 2 | 2 | 2 | 29 | 5 th |
| <i>Carissa spinarum</i> | 1 | 1 | 1 | 3 | 2 | 1 | 2 | 2 | 1 | 2 | 2 | 1 | 1 | 1 | 1 | 22 | 6 th |

NB: Scores given in the table indicate ranks of medicinal plants based on their efficacy. The highest number (6) given to the medicinal plant which informants thought most effective and the lowest number (1) for the least effective plant.

4.6.3.1.5 Efficacy of medicinal plants

About eight major ailment categories were identified from the 50 human ailments reported in the study area. The category with the highest Informat Consensus Factor (ICF) value was that of dermatological ailments (0.98), followed by external injuries, bleeding and snakebites (0.92) and gastrointestinal complaints (0.84) (Table 19). The highest plant use citation (17.72% each) was found for external injuries, bleeding and snakebites and sensory and nervous system, followed by dermatological ailments (15.19% each).

Table 19. ICF values of traditional medicinal plants used for treating human ailments

| No. | Ailment category | No. of Species | % all species | Use citations | ICF |
|-----|---|-------------------|---------------|---------------|------|
| 1 | Dermatological ailments | 12 | 15.19 | 52 | 0.98 |
| 2 | External injuries, bleeding and snakebites | 14 | 17.72 | 43 | 0.92 |
| 3 | Gastrointestinal complaints | 7 | 8.86 | 39 | 0.84 |
| 4 | Evil eye | 5 | 6.33 | 24 | 0.82 |
| 5 | Febrile illness | 6 | 7.59 | 27 | 0.80 |
| 6 | Sensory and nervous system | 14 | 17.72 | 60 | 0.77 |
| 7 | Oral, dental and pharyngeal ailment | 10 | 12.66 | 18 | 0.47 |
| 8 | Others | 7 | 8.86 | 12 | 0.45 |

4.6.3.1.6 Healing potential of medicinal plants used for treating human ailments

The highest fidelity level (96.15%) was obtained for *Verbascum sinaiticum*, followed by *Withania somnifera* (89.47%), *Eucalyptus globulus* (88.24%) and *Datura stramonium* (75.86%) (Table 20). *Verbascum sinaiticum* and *Withania somnifera* were reported to be used against dermatological ailments and sensory and nervous systems, respectively.

Table 20. Fidelity level values of medicinal plants commonly reported against a given human ailment category

| No. | Medicinal plant | Therapeutic category | Ip* | Iu* | FL (%)* |
|-----|--|--|-----|-----|---------|
| 1 | <i>Verbascum sinaiticum</i> | Dermatological ailments | 25 | 26 | 96.15 |
| 2 | <i>Withania somnifera</i> | Sensory and nervous systems | 17 | 19 | 89.47 |
| 3 | <i>Eucalyptus globulus</i> | Febrile illness | 15 | 17 | 88.24 |
| 4 | <i>Datura stramonium</i> | External injuries, bleeding and snakebites | 22 | 29 | 75.86 |
| 5 | <i>Clematis simensis</i> | Ailments of sensory and nervous systems | 18 | 24 | 75.00 |
| 6 | <i>Olea europaea</i> subsp. <i>cuspidata</i> | Gastrointestinal complaints | 20 | 27 | 74.07 |
| 7 | <i>Rumex nepalensis</i> | Evil eye | 10 | 19 | 52.63 |
| 8 | <i>Balanites aegyptica</i> | Oral, dental and pharyngeal ailments | 6 | 12 | 50.00 |

4.6.3.1.7 Comparison between different social groups

Significant differences ($p < 0.05$) were observed between the average numbers of medicinal plants claimed by men and women informants, by informants belonging to age groups below 50 and above 50 years, illiterate and literate informants and by key and general informant categories (Appendices 7,8,9 and 10). The higher averages were calculated for men than women, for older people than younger ones, for illiterate people than literate ones and for key informants than general informants (Table 21).

Table 21. Statistical test of significance on average number of medicinal plants among different informant groups.

| Parameters | Informant groups | N | Average \pm SD | t -value** | p -value |
|--------------------|---------------------|-----|-------------------|------------|----------|
| Gender | Male | 200 | 4.21 \pm 9.20 | 5.88 | 0.000* |
| | Female | 139 | 0.32 \pm 1.39 | | |
| Age | Young members (<50) | 254 | 1.61 \pm 5.99 | -1.38 | 0.030* |
| | Old members (>50) | 85 | 2.66 \pm 6.22 | | |
| Literacy level | Literate | 140 | 1.63 \pm 4.27 | -2.79 | 0.000* |
| | Illiterate | 199 | 3.55 \pm 8.29 | | |
| Informant Category | Key informant | 30 | 14.57 \pm 13.74 | 17.86 | 0.000* |
| | General informant | 309 | 0.34 \pm 1.12 | | |

*Significant difference ($p < 0.05$); ** t (0.05) (two tailed), df = 337, N= number of informants

4.6.3.1.8 Source and transfer of indigenous knowledge on medicinal plants

In the study area, it was widely observed that parents (50.66%) and grandparents (30.22%) served as the main source of traditional medical knowledge (Table 22). They commonly transferred their knowledge orally to their eldest son and other trusted family members secretly.

Table 22. Sources of knowledge on the practice of traditional medicine

| Source of knowledge for traditional healer | Percentage |
|---|-------------------|
| Parents | 50.00 |
| Grandparents | 35.22 |
| Wife/Husband | 8.55 |
| Neighbors | 3.00 |
| Friends | 1.22 |
| Sister/Brother | 0 |
| Uncle/Aunt | 0 |
| Other | 2.00 |
| Total | 100 |

4.6.3.2 Wild edible plants

Seventeen plant species were identified as edible. Of the recorded edible plants, shrubs constituted 15 species (88.23%) and trees two species (11.76%). In terms of plant parts used, fruits included 17 species (94.12%) and only one species (5.88%) was reported to be consumed for its young shoots (Appendix 11).

The preference ranking exercise on wild edible plants ranked: *Opuntia ficus-indica*, *Ficus sur* and *Carissa spinarum*: 1st, 2nd and 3rd, respectively (Table 23).

Table 23. Results of preference ranking of eight wild edible plants

| Wild edible plants | Informants labeled A to K | | | | | | | | | | | Total score | Rank |
|-----------------------------|---------------------------|---|---|---|---|---|---|---|---|---|---|-------------|-----------------|
| | A | B | C | D | E | F | G | H | I | J | K | | |
| <i>Opuntia ficus-indica</i> | 8 | 8 | 6 | 4 | 8 | 8 | 7 | 7 | 8 | 8 | 8 | 80 | 1 st |
| <i>Ficus sur</i> | 6 | 6 | 3 | 8 | 7 | 3 | 8 | 8 | 7 | 7 | 3 | 66 | 2 nd |
| <i>Carissa spinarum</i> | 7 | 5 | 8 | 6 | 5 | 7 | 5 | 6 | 5 | 4 | 5 | 63 | 3 rd |
| <i>Dovyalis verrucosa</i> | 3 | 3 | 2 | 7 | 6 | 6 | 2 | 4 | 6 | 5 | 4 | 48 | 4 th |
| <i>Rosa abyssinica</i> | 5 | 7 | 4 | 3 | 2 | 2 | 4 | 5 | 3 | 6 | 6 | 47 | 5 th |
| <i>Dovyalis abyssinica</i> | 4 | 4 | 5 | 2 | 3 | 4 | 3 | 3 | 2 | 2 | 7 | 39 | 6 th |
| <i>Sageretia thea</i> | 2 | 2 | 7 | 5 | 4 | 1 | 1 | 2 | 4 | 2 | 2 | 32 | 7 th |
| <i>Rubus steudneri</i> | 1 | 1 | 1 | 1 | 1 | 5 | 6 | 1 | 1 | 1 | 1 | 20 | 8 th |

NB: Scores in the table indicate ranks given to wild edible plants based on their edibility. Highest number (8) given for the edible plant which informants thought most preferred and the lowest number (1) for least preferred.

4.6.3.3 Fuelwood

Twenty-three plant species were preferred sources of fuelwood. The different plants harvested by the local community as a source of fuelwood included shrub (16 species, 69.57%) and tree (7 species, 30.43%) species. Fifteen species (65.22%) were used for their main trunk (stems) and eight species (34.78%) for their branches and were mostly harvested from the forest (Appendix 12).

4.6.3.4 Farm implements and building materials

Twenty-two plants were reported to be used for making traditional farm implements. Shrubs (15 species, 68.18%) and trees (7 species, 31.81%) were used. Concerning the plant part used, stem was commonly used (Appendix 13). Plough and its parts, yoke, handlers of (spade, pickaxe, scythe, harrow and hoe) were few among the specific uses of the plants species recorded.

Nineteen plant species were used for traditional construction of houses. Of these, shrubs (13 species, 68.42%) and trees (6 species, 31.58%) were frequently used. Stems were the commonly used plants parts (Appendix 14).

4.6.3.5 Livestock fodder/ forage

Twenty-six plant species were used as livestock fodder/ forage. Shrubs (19 species, 73.07%) were the most represented plant habits, followed by trees (4 species, 15.38%) and herb (3 species, 11.53%). With reference to the plant parts used only leaves were used as a source of fodder/ forage in the study area (Appendix 15).

4.6.3.6 Honeybee forage

Twenty-seven plant species were identified as sources of honeybee forage. The study shows that Fabaceae with four species followed by Lamiaceae, Acanthaceae, Malvaceae and Anacardiaceae with three species each were the major sources of nectar/pollen. Based on growth habit, shrubs contributed the highest number (22 species, 81.48%) followed by that of trees (4 species, 14.81%) and herb (1 species, 3.70%) (Appendix 16). Nectar/pollen was used the sole sources of honeybee forage. Preference ranking indicated *Erica arborea*, *Hypoestes forskaolii*, *Leucas abyssinica* and *Becium grandiflorum* as 1st, 2nd, 3rd and 4th, respectively (Table 24).

Table 24. Results of preference ranking of eight honeybee plants (**NB**: Scores in the table indicate ranks given to honeybee plants based on pollinators visiting. Highest number (8) given for the honeybee plant which informants thought most visited and the lowest number (1) for least visited)

| Honeybee plants | Informants labeled A to J | | | | | | | | | | Total score | Rank |
|--|---------------------------|---|---|---|---|---|---|---|---|---|-------------|-----------------|
| | A | B | C | D | E | F | G | H | I | J | | |
| <i>Erica arborea</i> | 8 | 8 | 1 | 7 | 4 | 6 | 7 | 6 | 7 | 8 | 62 | 1 st |
| <i>Hypoestes forskaolii</i> | 5 | 7 | 2 | 6 | 8 | 8 | 8 | 2 | 6 | 7 | 59 | 2 nd |
| <i>Becium grandiflorum</i> | 7 | 5 | 4 | 5 | 2 | 4 | 5 | 7 | 8 | 6 | 53 | 3 rd |
| <i>Leucas abyssinica</i> | 6 | 4 | 3 | 4 | 7 | 7 | 4 | 8 | 5 | 5 | 53 | 3 rd |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 2 | 6 | 8 | 8 | 6 | 5 | 6 | 4 | 4 | 4 | 53 | 3 rd |
| <i>Opuntia ficus-indica</i> | 4 | 3 | 6 | 3 | 1 | 2 | 2 | 3 | 3 | 3 | 30 | 6 th |
| <i>Aloe camperi</i> | 3 | 2 | 5 | 2 | 5 | 3 | 3 | 1 | 1 | 1 | 26 | 7 th |
| <i>Justicia schimperiana</i> | 1 | 1 | 7 | 1 | 3 | 1 | 1 | 5 | 2 | 2 | 24 | 8 th |

4.6.4 Multipurpose plants

Informants in all study sites reported that many forest plants have multiple uses. For example, *Olea europaea* subsp. *cuspidata* and *Rhus glutinosa* were reported to have about six uses each followed by *Teclea nobilis* with five uses. The rest of the plants had three to four uses each (Table 25).

Table 25. Direct matrix ranking of top seven multipurpose plants. Note: **Fo** = food; **Me** = medicine; **Fi** = farm implement; **Bm** = building materials; **Fw** = fuelwood; **Fd** = fodder; **Hb** = honeybee forage; **R**= rank)

| Species | Use category | | | | | | | | R |
|--|--------------|-----|-----|-----|-----|-----|-----|-------|-----------------|
| | Fo | Me | Fi | Bm | Fw | Fd | Hb | Total | |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 0.0 | 0.5 | 1.0 | 1.0 | 1.0 | 0.5 | 0.5 | 4.5 | 1 st |
| <i>Rhus glutinosa</i> | 0.5 | 0.0 | 1.0 | 1.0 | 0.5 | 0.5 | 0.5 | 4 | 2 nd |
| <i>Podocarpus falcatus</i> | 0.0 | 1.0 | 1.0 | 0.5 | 1.0 | 0.0 | 0.0 | 3.5 | 3 rd |
| <i>Teclea nobilis</i> | 0.0 | 0.0 | 0.5 | 0.5 | 1.0 | 1.0 | 1.0 | 3.5 | 3 rd |
| <i>Opuntia ficus-indica</i> | 1.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.0 | 1.0 | 3 | 5 th |
| <i>Becium grandiflorum</i> | 0.0 | 0.5 | 0.0 | 0.0 | 1.0 | 0.5 | 1.0 | 3 | 6 th |
| <i>Juniperus procera</i> | 0.0 | 0.5 | 1.0 | 1.0 | 1.0 | 0.0 | 0.0 | 3.0 | 7 th |
| <i>Carissa spinarum</i> | 1.0 | 1.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.5 | 2.5 | 8 th |
| <i>Pittosporum viridiflorum</i> | 0.0 | 0.0 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 2.5 | 8 th |
| <i>Dodonaea angustifolia</i> | 0.5 | 0.0 | 0.0 | 0.5 | 1.0 | 0.5 | 0.0 | 2.5 | 8 th |

4.6.5 Major threats to forest resources

Information showed that the forest had been subjected to frequent tree cutting by members of the local community, because of population growth and poverty. According to the informants, the major factors reported for forest destruction were deforestation (38%), agricultural expansion (25%), fuelwood collection (18%), livestock grazing (9%), drought (5%) and settlement (3%) in decreasing order.

There were a number of plant species that were commonly over exploited, because of human influence. Priority ranking was employed to identify those locally threatened plant species as perceived by the local people. The results indicated that *Olea europaea* subsp. *cuspidata*, *Erica arborea* and *Podocarpus falcatus* were the 1st, 2nd and 3rd most threatened plant species in the forest, respectively (Table 26).

Table 26. Priority ranking of locally threatened tree/shrub species as identified by the local people

| Species | Informants and scores | | | | | | | | | | | | | | | Total score | Rank |
|--|-----------------------|---|---|---|---|---|---|---|---|----|----|----|----|----|----|-------------|-----------------|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | | |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 7 | 8 | 8 | 8 | 7 | 118 | 1 st |
| <i>Erica arborea</i> | 5 | 6 | 7 | 7 | 7 | 4 | 6 | 5 | 7 | 6 | 8 | 7 | 8 | 7 | 8 | 98 | 2 nd |
| <i>Podocarpus falcatus</i> | 4 | 5 | 5 | 5 | 6 | 6 | 7 | 7 | 5 | 7 | 4 | 5 | 7 | 6 | 6 | 85 | 3 rd |
| <i>Juniperus procera</i> | 7 | 4 | 6 | 6 | 4 | 7 | 6 | 6 | 6 | 5 | 5 | 4 | 5 | 5 | 4 | 80 | 4 th |
| <i>Ficus sur</i> | 2 | 3 | 1 | 3 | 5 | 2 | 5 | 4 | 4 | 4 | 6 | 2 | 8 | 3 | 1 | 53 | 5 th |
| <i>Teclea nobilis</i> | 6 | 7 | 2 | 4 | 2 | 3 | 1 | 1 | 3 | 3 | 1 | 6 | 2 | 1 | 3 | 45 | 6 th |
| <i>Psydrax schimperiana</i> | 3 | 1 | 3 | 1 | 1 | 5 | 3 | 3 | 2 | 1 | 3 | 1 | 1 | 4 | 5 | 37 | 7 th |
| <i>Dovyalis abyssinica</i> | 1 | 2 | 4 | 2 | 3 | 1 | 2 | 2 | 1 | 2 | 2 | 3 | 5 | 2 | 2 | 34 | 8 th |

NB: Scores in the table indicate ranks given to locally threatened plants. Highest number (8) given for the most threatened plant which informants thought and the lowest number (1) for least threatened.

4.6.6 Cultural Significance Index

Table 27 presents a cultural significance index of eight most cited plant species. The calculation revealed *Olea europaea* subsp. *cuspidata*, *Podocarpus falcatus*, *Acacia etbaica*, *Juniperus procera* and *Balanites aegyptiaca* as the first five most culturally significant plant species.

Table 27. Cultural Significance Index (CSI) of some most cited plants: **NIC** = Number of informant citations; **WV** = Weighted Variables (**i** = species mgmt [non-managed (1) or managed (2)] **e** = Use Preference [not preferred (1) or preferred (2)] **c** = Use Frequency [rarely used (1) or used frequently (2)], **Fo** = food; **Me** = medicine, **Fi** = farm implement, **Bm** = building materials; **Fw** = fuelwood, **Fd** = fodder; **Hb** = honeybee forage, [**CF** = Correction factor the number of informant citations for a given species divided by the number of informant citations for the most cited species].

| No. | Plant species | NIC | WV | Specific uses (SU) | | | | | | | Sum | | |
|-----|-----------------------------|-----|---------|--------------------|----|----|----|----|----|----|---------|------|-------|
| | | | | Fo | Me | Fi | Bm | Fw | Fd | Hb | (i*e*c) | CF | CSI |
| 1 | <i>Acacia etbaica</i> | 46 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 2 | 1 | 2 | 1 | 2 | 1 | 1 | | | |
| | | | (c) | 2 | 2 | 2 | 2 | 2 | 2 | 2 | | | |
| | | | (i*e*c) | 4 | 2 | 4 | 2 | 4 | 2 | 2 | 20 | 0.76 | 15.20 |
| 2 | <i>Balanites aegyptiaca</i> | 55 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (c) | 2 | 2 | 2 | 2 | 1 | 1 | 1 | | | |
| | | | (i*e*c) | 2 | 2 | 2 | 2 | 1 | 1 | 1 | 11 | .91 | 10.01 |

| No. | Plant species | NIC | WV | Specific uses (SU) | | | | | | | Sum | | |
|-----|--|-----|---------|--------------------|----|----|----|----|----|----|---------|------|-------|
| | | | | Fo | Me | Fi | Bm | Fw | Fd | Hb | (i*e*c) | CF | CSI |
| 3 | <i>Becium grandiflorum</i> | 30 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 2 | 1 | 1 | 2 | 2 | 2 | | | |
| | | | (c) | 1 | 2 | 1 | 1 | 2 | 2 | 2 | | | |
| | | | (i*e*c) | 1 | 4 | 1 | 1 | 4 | 4 | 4 | 19 | 0.50 | 9.5 |
| 4 | <i>Cadia purpurea</i> | 25 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 1 | 2 | 2 | 2 | 1 | 1 | | | |
| | | | (c) | 1 | 1 | 2 | 2 | 2 | 2 | 2 | | | |
| | | | (i*e*c) | 1 | 1 | 4 | 4 | 4 | 4 | 2 | 20 | 0.41 | 8.2 |
| 5 | <i>Erica arborea</i> | 19 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 1 | 1 | 1 | 1 | 1 | 2 | | | |
| | | | (c) | 2 | 1 | 1 | 2 | 2 | 2 | 2 | | | |
| | | | (i*e*c) | 2 | 1 | 1 | 2 | 2 | 2 | 4 | 14 | 0.32 | 4.48 |
| 6 | <i>Juniperus procera</i> | 57 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 2 | 1 | 1 | 2 | 1 | 1 | | | |
| | | | (c) | 1 | 2 | 1 | 1 | 2 | 1 | 1 | | | |
| | | | (i*e*c) | 1 | 4 | 1 | 1 | 4 | 1 | 1 | 13 | 0.95 | 12.35 |
| 7 | <i>Olea europaea</i> subsp. <i>cuspidata</i> | 60 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 2 | 2 | 2 | 2 | 2 | 2 | | | |
| | | | (c) | 1 | 2 | 2 | 2 | 2 | 2 | 2 | | | |
| | | | (i*e*c) | 1 | 4 | 4 | 4 | 4 | 4 | 4 | 25 | 1 | 25.00 |
| 8 | <i>Podocarpus falcatus</i> | 50 | (i) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | | |
| | | | (e) | 1 | 2 | 2 | 2 | 2 | 1 | 1 | | | |
| | | | (c) | 1 | 2 | 2 | 2 | 2 | 1 | 1 | | | |
| | | | (i*e*c) | 1 | 4 | 4 | 4 | 4 | 1 | 1 | 19 | 0.83 | 15.77 |

CHAPTER FIVE

5. DISCUSSION, CONCLUSION AND RECOMMENDATIONS

5.1 Discussion

5.1.1 Vegetation study

5.1.1.1 Floristic composition and diversity

According to this study, there are 264 species in the area with the possibility of discovering few species more as seen from species accumulation curve, which seems to indicate the possibility for discovering few additional species with increasing sampling effort. The richness of the study site agrees with the fact that the pattern of species richness in East African montane forests (Conservation International, 2007). Variations of species richness occurred among the forests could be attributed to the area sampled (Hill and Curan, 2003) and large range of altitude (Tamrat Bekele, 1993), climate variability (Schmitt *et al.*, 2013) and forest disturbance (Økland, 1990). The Ethiopian vegetation was highly influenced by climate, which was associated with elevation (Dugdale, 1964). Pollen studies in Kenya (Lamb *et al.*, 2003) and northern Ethiopia (Darbyshire *et al.*, 2003) indicated that the flora of East Africa has experienced several changes due to climate change and anthropogenic disturbances.

Consequently, species richness in the study area was greater than in *Alemsaga* with 124 species (Getinet Masresha *et al.* 2015), *Chencha* with 174 species (Desalegn Wana and Zerihun Woldu, 2005), *Dense* with 158 species (Ermias Lulekal, 2014)), *Dess'a* with 130 species (Ermias Aynekulu, 2011), *Dodola* with 113 species (Kitessa Hundera *et al.*, 2007), *Hugumburda-Gratkahsu* with 102 species (Leul Kidane *et al.* 2010), *Peninsula-Zegie* with 113 species

(Alemnew Alelign *et al.*, 2007) and *Taragedam Abebaye* with 143 species (Haileab Zegeye *et al.*, 2011). However, the study area less species-rich than *Borena Sayint* with 397 species (Hussien Adal, 2014) and *Hugumburda-Gratkahsu* with 326 species (Leul Kidane, 2015).

The family with high species representation was Asteraceae (27 species, 32.93%). Mesfin Tadesse (2004) stated that Asteraceae was among the richest family in the flora area (Flora of Ethiopia and Eritrea (FEE)). According to Hedberg (1964), wind dispersal is one of the main mechanisms affecting long-distance dispersal for Afromontane landscapes. Moreover, many species of Asteraceae have umbrella shape capitula that are adapted for wind dispersal (Hedberg, 1964) and increase their opportunity for the successful establishment of their populations. The study was in accordance with results of other studies, for instance, *Boditi* (39 species, 17%; Haile Yineger *et al.*, 2008), *Komto* (17 species, 9.44%; Fekadu Gurmessa *et al.*, 2013), *Dense* (20 species, 27%; Ermias Lulekal, 2014), *Hugumburda-Gratkahsu* (38 species, 11%; Leul Kidane, 2015). However, in terms of species richness found in the FEE, Fabaceae (678 species), Poaceae (609 species), Asteraceae (472 species) and Euphorbiaceae (472 species) were the richest families (Ensermu Kelbessa and Sebsebe Demissew, 2014). Compared to other life forms or habits, high richness of herbaceous species was recorded. This may be attributed to disturbance events that created gaps in canopy cover increasing open spaces in favor of pioneer herbaceous and shrub species. As disturbance increase, pioneer shrubs and herbs may encroach into tree-dominated landscapes (Ermias Aynekulu, 2011), which may be the case in the present study area. Similar studies have reported related results from other dry Afromontane forest whereby herbs are found to be dominant life form or habits (Haile Yineger *et al.*, 2008; Fekadu Gurmessa *et al.*, 2013; Ermias Leulekal, 2014; Leul Kidane, 2015).

The number of endemic species harbored in the forest was 13 species. Of this, six species were included in the IUCN Red List Categories of Ethiopia and Eritrea (Vivero *et al.*, 2005). Asteraceae and Lamiaceae were the largest families having three endemic species each in the study area. Studies conducted elsewhere in the flora area reported Asteraceae as the largest family (Abiyou Tilahun *et al.*, 2011; Ensermu Kelbessa and Sebsebe Demissew, 2014; Abiyot Berhanu, 2017). Nearly half of the world's vascular plant species are endemic to 25 hotspots (Myers *et al.*, 2000), of which 17 are in tropical forests each having at least 1500 endemic plant species (Brooks *et al.*, 2002). The hotspot concept is based on species richness, endemism and potential threat (with 70% of its natural habitats degraded; Myers *et al.*, 2000). However, many plant species have a too small population size to be viable and this may increase the rate of extinction in the long term (Whitemore, 1997). It has been known that the Afromontane forests contained higher endemism (White, 1978) than the other regions in Africa. Consequently, over 3,000 endemic plant species estimated to be found in this region (White, 1978). However, there has been a threat to the endemic plants found in the study area and thus need immediate action to protect them.

The Shannon - Wiener diversity index was high ($H' = 3.87$) in the study area. Kent and Coker (1992) explained that the Shannon – Wiener diversity index normally varies between 1.5 and 3.5, rarely exceeds 4.5. This value is found to be higher than that of other montane forests, such as *Chilimo* ($H' = 2.72$; Tadesse Woldemariam *et al.*, 2000), *Tara Gedam* ($H' = 2.98$; Haileab Zegeye *et al.*, 2011). Similarly, the evenness ($J = 0.89$) was higher than that of *Chilimo* ($J = 0.68$; Tadesse Woldemariam *et al.*, 2000), *Tara Gedam* ($J = 0.65$; Haileab Zegeye *et al.*, 2011). Altitude and slope were important environmental variables to influence species richness and dispersal activities (Tamrat Bekele, 1994; Ermias Aynekulu, 2011). Generally, habitat diversity

is a widely accepted determinant of species diversity (Mutke and Barthlott, 2005). The theory of spatial heterogeneity (Pianka, 1966) stated that more heterogeneous and complex physical environments support more plants that are diverse.

5.1.1.2 Plant communities

The vegetation types in the study site can be broadly classified as dry single-dominant Afromontane forest of the Ethiopian highlands, transition between single dominant Afromontane and east African evergreen and semi-evergreen bushland (Friis, 1992), and modified to dry evergreen Afromontane forest and grassland complex (DAF) (Undifferentiated Afromontane forest, Afromontane woodland, wooded grassland and grassland and dry single-dominant Afromontane of the Ethiopian highlands) (Friis *et al.*, 2010).

The plant communities identified from Agglomerative hierarchical cluster analysis are described below:

Cadia purpurea – *Carissa spinarum* community belongs to the undifferentiated Afromontane forests. *Cadia purpurea* occurred in association with *Calpurnia aurea*, *Acokanthera schimperi* and *Carissa spinarum*. They were found on the edge of the forest that was exposed to frequent livestock grazing and human impacts. It can be classified as degraded shrub vegetation. All the indicator species were shrub species. Thus, the expansion of *Cadia purpurea* in the community can be considered as an indicator of disturbance. *Podocarpus falcatus*, *Allophylus abyssinicus*, *Celtis africana* and *Bersama abyssinica* were common in the undifferentiated Afromontane forests (Friis, 1992), though small number, because of the presence of human-induced disturbances.

Olea europaea subsp. *cuspidata* – *Juniperus procera* community is a dry single dominant Afromontane forest with *Juniperus procera* and *Olea europaea* subsp. *cuspidata* as dominant species (Friis, 1992). It was found on the higher altitude and sloppy area for having little disturbance in the community. The forest occurred NW and SE Ethiopian highlands (especially the Tigrean and the Harar plateau), at altitudes from 2200- 3200 at annual average temperature 12 – 18 °C and annual rainfall between 500 and 1500 mm (Friis, 1992). A sample of dry forest from central Tigray has been described by Wilson (1977) and perhaps be taken as an example of the northern plateau forests.

Dodonaea angustifolia – *Acacia abyssinica* community formed like a belt on the massif that preferred non-calcareous soil and at an altitude 2224 to 2640 m.a.s.l. It can be classified into dry single-dominant Afromontane forest and East African evergreen and semi-evergreen bushland. This category includes a range of physiognomic types, from typical forest to evergreen scrub with dispersed trees, but floristically the whole range is connected. The forest-like types exist in parts of Eritrea, on the eastern escarpment of the NW Highlands (part of Tigray) and on the northern escarpment of the SE highlands (Friis, 1992).

Erica arborea – *Myrsine africana* community was dominated by *Erica arborea* in the upper canopy and the lower story was dominated by *Myrsine africana*. It was found on the primary high and medium altitude montane evergreen bushland (Friis, 1992). This was in agreement with the general pattern presence of *Erica arborea* dominated communities at higher ranges of Afromontane forests (Demel Teketay and Tamrat Bekele, 1995a). *Erica arborea* was found as low as 2100 m.a.s.l, but they were few in number.

Acacia etbaica – *Acacia tortilis* community was found at the lower altitudes, having shallow soils and that received a lower amount of annual rainfall with higher temperatures. This community was highly influenced by people collecting firewood, charcoal making and livestock grazing. This was due to the place where it was found on the junction of the Afar and Tigray regions (*Raya-Azebo* and *Hintalo-Wejirat* districts), where the tragedy of the commons affected the community. However, there were attempts to protect from severe livestock grazing and overexploitation by the *Hintalo-Wejirat* district for they believed that the forest had a paramount role in bringing rainfall to the rest of their area (Agricultural and pastureland).

5.1.1.3 Ordination

Based on the CCA examination of the relationship between environmental gradients revealed that a strong relationship between altitude and slope, which means that steep slope was more common at higher altitudes (Tamrat Bekele, 1993; Leul Kidane, 2010; Ermias Aynekulu, 2011). Altitude and slopes have mainly influenced *Olea europaea* subsp. *cuspidata* – *Juniperus procera* and *Erica arborea* - *Myrsine africana* communities. Hedberg (1964) stated that altitude affects atmospheric pressure, moisture and temperature in an area whereby the latter directly influence the growth and development of plants and the corresponding patterns of vegetation distribution. Altitude and slope were important environmental gradients portioning forest communities in the Afromontane forests of Ethiopia (Zerihun Woldu and Backeus, 1991; Tamrat Bekele, 1994). *Cordia purpurea* – *Carissa spinarum* community experienced disturbances. It was found that shrubs were very common in this community. Canopy gaps may favor the spread of shrub and herb species, like *Cordia purpurea* (Ermias Aynekulu, 2011). The disturbances were human impacts and livestock grazing that have also had a connection in the pattern. If there was

livestock grazing, there were illegal cutting. *Acacia etbaica* – *Acacia tortilis* community type was highly influenced by livestock grazing and human impacts and habitat fragmentation from the continuous mountainous forest. CCA2 was positively correlated to livestock grazing and human impacts, while negatively correlated to slope and altitude. Anthropogenic disturbance affects species diversity through habitat loss and habitat fragmentation (Brooks *et al.*, 2002). Habitat fragmentation affects biodiversity by reducing the area of a habitat, which reduces the persistence of a species and through the negative edge effect that increases mortality while decreasing reproduction (Farhig, 2003). The vegetation on the Afromontane belt (900-3200 m) of Ethiopia has been under tremendous pressure from human activities and overgrazing, which has led to the replacement of the evergreen forests by grasslands (Tewolde Berhan Gebre Egziabher, 1988).

5.1.1.4 Vegetation structure

Woody species density in the study area showed relatively high value (1351.50 ha⁻¹; Table 4) as compared to other afromontane forests, such as *Dense* (1138 stems ha⁻¹; Ermias Leulekal, 2014) and *Hugumburda* (1235.9 stems ha⁻¹; Ermias Aynekulu, 2011). Variations in density distributions can be attributed to variations in habitat preferences of species and topographic gradients forming the forest and the degree of anthropogenic influences (Whittaker *et al.*, 2003).

The stand basal area at the study forest (7.08 m² ha⁻¹) is far less than that of other tropical and subtropical dry forests (17-40 m² ha⁻¹; Murphy and Lugo, 1986), which is evidence for the over-exploitation of the forest, especially of tree species, such as *Olea europaea* subsp. *cuspidata*, *Podocarpus falcatus* and *Juniperus procera*. The plant species with the largest basal area in the

forest could be the most important in terms of economic, cultural, ecological and social significance.

The most frequent species were *Olea europaea* subsp. *cuspidata*, *Juniperus procera*, *Rhus glutinosa* and *Maytenus arbutifolia*. Canopy trees, such as *Olea europaea* subsp. *cuspidat* and *Juniperus procera* are also dominant species in the dry Afromontane forests between (1600-) 2200 and 3200 (-3300) meters with annual rainfall between 500 and 1500 millimeters (Friis *et al.*, 2010). High frequency of a species always depends on factors, which relate to habitat preferences, adaptation, degree of exploitation and availability of suitable conditions for regeneration (Rey *et al.*, 2000).

5.1.1.5 Vertical structure of the forest

The International Union of Forest Research Organizations (IUFRO) classification scheme classifies vertical structure of forests into three layers/stories based on mean heights of canopy tree species (Lampercht, 1989). This classification is entirely related to woody species (trees and shrubs). The upper story species were *Podocarpus falcatus*, *Celtis africana*, *Olea europaea* subsp. *cuspidata*, *Juniperus procera*, *Allophylus abyssinicus*, *Ekebergia capensis*, *Acacia abyssinica*, and *Pittosporum viridiflorum*. The most common middle story species were *Psydrax schimperiana*, *Bersama abyssinica*, *Teclea nobilis*, *Cadia purpurea*, *Calpurnia aurea*, *Carissa spinarum*, *Acokanthera schimperi* and *Justicia schimperiana*, *Cassipourea malosana*, *Dovyalis abyssinica*, *Pavetta abyssinica*, *Erica arborea*, *Dodonaea angustifolia*, *Maytenus arbutifolia* and *Rhus glutinosa*, *Acacia etbaica*, *Acacia senegal*, *Ziziphus spina-christi* and *Dichrostachys cinerea*. Some of the most common lower story species were *Acacia tortilis*, *Myrsine africana*, *Clerodendrum myricoides*, *Asparagus racemosus*, *Berberis holstii*, *Clutea abyssinica*, *Colutea*

abyssinica, *Smilax aspera*, *Becium grandiflorum* and *Gomphocarpus fruticosus*. Most of the shrubs were in this category. According to White (1983), this layer is mixed and the shrub layer is less distinguished from the lower tree layer.

5.1.1.6 Importance value index of woody species

Importance value index (IVI) analysis showed higher values for *Dodonaea angustifolia* and *Olea europaea* subsp. *cuspidata* than for any other species in the study area. Higher basal area for *Dodonaea angustifolia* and higher frequency for *Olea europaea* subsp. *cuspidata* of these species were the contributing factors for having high IVI values. The relative ecological significance and/or dominance of a tree species in a forest ecosystem could be identified from IVI analysis (Mueller-Dombois and Ellenberg, 1974; Curtis and McIntosh, 1951). Hence, in this study, the IVI results also confirmed that these two species (*Dodonaea angustifolia* and *Olea europaea* subsp. *cuspidata*) are the most important/dominant species in the study area.

5.1.1.7 Species population structure and regeneration status

Measurements of DBH for some woody plant species showed a different species population structure. The first population structure represented by *Maytenus arbutifolia* (Figure 6), revealed the presence of the highest density in the lower DBH classes with a gradual decrease in the density towards the higher classes. It represents an inverted J – shaped distribution except for a slight decrease in the first DBH class. Such population was an indication of stable population structure and good regeneration status (Fekadu Gurmessa *et al.*, 2012; Ermias Leulekal, 2014; Leul Kidane, 2015).

The second structure was represented by *Cadia purpurea* (Figure 6), it represented by Gauss-type distribution structure or bell-shaped distribution formed by species where the frequency distribution of individuals in the lower and higher DBH classes was lower than the middle classes. It was found on the edge and disturbed area that experienced selective cutting of big individuals and grazed the seedling severely.

The third structure was represented by *Bersama abyssinica* (Figure 6) revealed that few individuals in the lower diameter class and individuals were absent in the intermediate diameter class and relatively higher number of individuals in the highest diameter class. It represented Broken J-shaped. This structure was caused by the selective removal of a mature individual for construction purpose (Feyera Senbeta, 2006; Alemnew Alelign *et al.*, 2007). The fourth structure was *Myrsine africana* (Figure 6), showed that individuals concentrate in the first (lower) and intermediate (third and fourth) diameter classes. The rest of the classes had almost equally few numbers of individuals that were few.

According to Popma *et al.* (1988), plant population structures help to understand population dynamics and regeneration status of species in the forest. Overall, population structure of tree species depending on size-class distribution yielded an inverted J-shaped curve in the forest (Figure 8). This agrees with a general of normal population structure where the number of individual's decreases as the height and DBH class increases, showing the forest was in a stage of a sign of regeneration. This type of population structure was an indicator of a healthy forest. Nevertheless, this does not meant each species found in the study area followed the same population structure. There were species that do not have seedlings so that it was the sum total of the whole forest indicated promising result for regeneration. Climatic factors and biotic

interference influence the regeneration of different species in the vegetation (Khan *et al.*, 1987). According to Alemayehu Wassie (2007), some species livestock-induced disturbance might be among the major factors constraining regeneration and recruitment of woody species and contributing, ultimately, to the decline of woody species population in the forests.

5.1.1.8 Floristic similarities with other dry Afromontane forests

The observed high Sorensen similarity values obtained from the floristic comparison of *Wejig-Mahgo-Waren* and *Hugumburda-Gratkahsu* (0.39; Leul Kidane, 2015) forests and *Wejig-Mahgo-Waren* and *Dense* (0.37; Ermias Leulekal, 2014) forests indicated that *Wejig-Mahgo-Waren* massif forest was floristically more close to *Hugumburda-Gratkahsu* and *Dense* forests than to the other six forests compared. The least similarity was revealed with *Peninsula-Zegie* (0.18; Alemnew Aleign *et al.*, 2007) and *Chencha* (0.26; Desalegn Wana and Zerihun Woldu, 2005). The regional climate has a paramount role in differences for species distribution patterns in tropical montane forests, especially in those with highly diverse topography (Schmitt *et al.*, 2013). The higher similarity observed was both forests found in the dry Afromontane forests that believed they share a close similar climate, altitude interval and geographical proximity with *Hugumburda-Gratkahsu*. The ecological distance between any two areas has also been noted to play an important part for differences in species composition (Mueller-Dombois and Ellenberg, 1974). For example, *Chencha* forest found in the southern nation, nationalities and people region that was very far from the study area, which is at the northern tip of the country. Thus, the dissimilarity observed between the study area and *Chencha* forest was owing to ecological distance.

5.1.2 Soil seed bank study

Knowledge of which species are not represented in the persistent soil seed bank can be just as important as knowing which species are represented. Soil seed banks reflect partly the history of the vegetation and can play an important role in its restoration after disturbances. They have been exploited in two contexts: to manage the composition and structure of existing vegetation and to restore or establish native vegetation (van der Valk and Pederson, 1989). In forest management, natural seed banks play a vital role in restoration after disturbances. The findings of the study area revealed that the prevailing habits in the soil seed bank of the forest were herbs and only a small proportion of seed abundance and species richness of woody species. The predominance of herbaceous may be attributed to small seed size, which makes them more easily incorporated into the soil to form seed bank and also less prone to predation (Thompson, 1987, Guo *et al.*, 1998, Luzuriaga *et al.*, 2005). Previous researches have indicated that seeds of non-woody species can be dormant for a long time under unfavorable environmental conditions, while the tree species with large seed masses easily tend to be rotten or germinate soon after seed fall (Decocq *et al.*, 2004; Daws *et al.*, 2005).

The buildup of seeds in the soil was favored by the dormancy of many of the seeds, which is attributed to either the presence of embryo dormancy or impermeable seed coat or both. Dormancy is selected in most dry Afromontane species, which are characterized by long dry seasons and unreliable rainy periods. In non-dormant seeds, dormancy will be induced if the seeds are dispersed under a dense canopy or buried in the soil (Demel Teketay and Granstrom, 1995b; Demel Teketay and Granström, 1997).

Most of the woody component of the forest lacked reserves of long-lived seeds in the soil. Of the 120 woody species recorded in the standing vegetation of the forest, only eight species (6.67%) were represented in the soil seed banks. Low similarity levels between standing vegetation and the seed banks indicate that one should increase work on forest protection and management because of its potentially low restoration ability (Cui *et al.* 2016). Indeed, the seedling emergence method can dramatically underestimate the density of the seed bank due to errors associated with seed dormancy and specific environmental requirements for germination (Bernhardt *et al.*, 2008; Wright and Clarke, 2009). These findings agreed with those of different authors (Kebrom Tekle and Tesfaye Bekele, 2000; Feyera Senbeta and Demel Teketay, 2001; 2002; De Villiers *et al.*, 2003; Ericksson *et al.*, 2003; Amiaud and Touzard, 2004; Tefera Mengistu *et al.*, 2005; Alemayehu Wassie, 2007) who reported higher and lower proportions of herbaceous and woody species in the soil seed banks, respectively. The low values of similarity indices between the seed bank and the standing vegetation provided more proof of the weak contribution of soil seed banks to mature stands. Similar conclusions were made from other studies (Perera, 2005; Uasuf *et al.*, 2009).

The spatial distribution of seeds of different species in vertical distribution varied greatly. Seed abundance and number of species generally decrease with the soil depth, especially the seeds of woody species accumulated in the upper 0-5 cm soil layers. For example, seeds of *Becium grandiflorum*, *Cadia purpurea* and *Dodonaea angustifolia* characterized relatively by their large sizes, when compared with the herbaceous ones, were found on the upper 0-5 cm soil layers. These variations may reflect differences of species in terms of seed longevity in the soil, mode of seed dispersal, seed predation and probable differences in altitude and local edaphic conditions where seeds land. This result is also in agreement with past similar works (Feyera Senbeta and

Demel Teketay, 2001; 2002; Ericksson *et al.*, 2003; Matus *et al.*, 2005; Tefera Mengistu *et al.*, 2005; Alemayehu Wassie, 2007).

Soil seed bank species have been reported to have the following characters related to their success in establishing after disturbance: production of numerous small seeds means of long-distance dispersal, formation of persistent soil seed banks and the capacity to remain viable in a dormant state for a long period of time (Demel Teketay, 2002; Alemayehu Wassie, 2007).

Dry Afromontane forests can be characterized by possessing large populations of buried seeds of herbs whereas trees mainly have persistent seedling banks and the ability to sprout from damaged roots or shoots. The fact that most of the dominant tree species do not accumulate seeds in the soil suggests that their restoration from seeds would be prevented by removal of mature individuals in the standing vegetation. This, in turn, implies that restoration forests would be difficult and slow to accomplish if they are destroyed (Demel Teketay, 2002). The herbaceous vegetation has a better chance of recovery since it has a diverse soil seed bank with great seed longevity (Alemayehu Wassie, 2007). The results provide further evidences that consolidate the conclusions of previous studies on soil seed banks, i.e., the future existence of the woody flora characteristic of dry Afromontane areas in Ethiopia depends on the conservation and sustainable utilization of the few remnant natural forests (Alemayehu Wassie, 2007), revegetation of indigenous seedlings from nursery and exclosure for natural restoration.

The soil seed banks appeared not to contribute largely to the restoration of woody species. Clearly, the soil seed bank could not provide a good supply of diaspores of woody species. For such purpose, planting in logging gaps with threatened indigenous seedlings coming from the nursery should be more efficient (Doucet *et al.*, 2009).

5.1.3 People and plant interactions

5.1.3.1 Concept about the forest and forest resources

The results indicated that traditional concept about forest resources was tied up with the use of plants. Forest is the most precious gift, nature has provided to us, as it is meeting all kinds of essential requirements of the humans in the form of medicine, food, fodder, fuel. According to Zemedu Asfaw (2006), the people of Ethiopia are knowledgeable about the name and classification of their environment, plants in their surroundings and their value for the local people, which they have gained orally from generation to generation. Forest for the majority of the rural people meant everything, no-forest-no-life (Melese Damite, 2001). Indigenous knowledge about plants has been a matter of survival to the people who have generated these systems (Grenier, 1998) and indigenous or local people throughout the world have their own distinct culture, linguistic, geographical separation, values and beliefs (Furze *et al.*, 1997). The concept of local people on plants is developed over time involvement, reflection and discussion that affect their beliefs and attitudes towards forest management practices (Cotton, 1996).

5.1.3.2 Management of forest plant resources

There was a belief related to *Olea europaea* subsp. *cuspidata* in the community when a single tree of the plant was cut, the community considered it as if a person was killed. In seeking to understand how indigenous peoples might live in harmony or balance with the natural world, the idea of taboo as a conservation strategy was popularized. Cultural taboos are informal activities that determine human behavior (Colding and Folke, 2001). They involve restrictions by certain sectors of society on the use of particular resources and habitats, sometimes only at particular

times or in particular places (Colding and Folke, 2001). Cultural taboos are an important aspect of plant selection by societies throughout the world. It is a taboo to use certain plant species to protect them from overexploitation and extinction.

This study has brought into picture the traditional management system of the forest held by the local people that has been handed over down to the present. The forest massif was found on the common borders of the three districts and that resulted in competition for resources. The tragedy of the commons has been observed in the study area. People fought for grazing, fuelwood and agricultural lands. As a result, there were places on the border areas that were highly affected. The forest was used for intermittent oxen grazing, though it was illegal.

Currently, the Tigray Bureau of Agriculture (Natural Resource Division) had managed to take the responsibility of protecting the forest. Guards were hired for a round clock protection of the forest. There were two types of local forest management types in the area, namely communal and protected “Hizaeti” where the ownership right was given to both the government and the community. The communal forest was protected through community consensus and grazing was allowed seasonally. While government protected forest was not allowed for grazing anytime, but there were times (September to November) that the community was allowed to let oxen in the forest graze without the permission of the district administration. This agrees with the report of the Kerreyu people of Ethiopia, where land is seasonally grazed (Kebu Balemie *et al.*, 2004). The local communities in Mukogodo forest of Kenya similarly use the forest for dry grazing (Komwihangilo and Mwilawa, 2006). All these studies showed similar plant use knowledge among different traditional people of different geographic areas regarding grazing area management. When there happened to be a special program, which could be a wedding or

funeral, collection of dried trees from the forest was possible with permission from sub-district natural resource agent.

One very fascinating phenomenon was beehive keeping inside the forest on the branches of *Podocarpus falcatus* and *Ficus sur*, owned by some so-called early settlers. Both the plants and traditional knowledge were received from their ancestors through legacy. There were no conflict of interests on the trees used for beehive inside the forest, though the forest was believed to be a property of the government and the community. However, there were orders from the government to remove beehive from the forest for fear of forest fire. Similar results have been reported from elsewhere in Ethiopia: Dess'a forest (Abrha Tesfay, 2008) and Borena Sayint National Park (Hussien Adal, 2014).

5.1.3.3 Major plant use categories

5.1.3.3.1 Medicinal plants

The local communities have accumulated substantial knowledge on natural resource found in their environment, including plants. Consequently, 79 plants were recorded to be used as medicinal plants, wild edible plants, fuelwood, farm implements, building materials, livestock fodder/ forage and honeybee forage. Among these resources, none has received more recent scientific and popular attention than tropical medicinal plants (Voeks and Leony, 2004). Of these resources, 52 were medicinal plants and the number was found to be higher compared to those reported elsewhere in Ethiopia: *Zay* people (33 species; Mirutse Giday, 2001), *Tanqua-Abergele* and *Kolla-Tembien* (29 species; Gebremedhin Gebrezgiher *et al.*, 2013), in and Around *Alamata* (16 species; Gidey Yirga, 2010a), *Mekelle* town (25 species; Gidey Yirga, 2010b) and

less as compared to those reported from studies conducted in *Kilte Awlalelo* (114 species; Abraha Teklay *et al.*, 2013), *Ofla* (113 species; Nurya Abdurhman, 2010), *Asgede Tsimbila* (68 species; Girmay Zenebe *et al.*, 2012), *Ofla* and *Raya-Azebo* (83 species; Mirutse Giday and Gobena Ameni, 2003), *Atsbi* and *Adi Keyih* (58 species; Moravec *et al.*, 2014) and *Ankober* (151 species; Ermias Leulekal, 2014). Results showed the role played by traditional medicinal plants in meeting the primary health care of people. Ease of accessibility, economic, culture and efficacy related aspects might have played key roles for the people to rely on traditional medicine (Ermias Leulekal, 2014). The informants agreed that more medicinal plants were used in the past than today. Moreover, there was deforestation and drought that resulted in the loss of medicinal plants in the study area.

The fact that shrubs were dominant in the study area may indicate the dry Afromontane forest has been converted into shrubland, because of massive deforestation. Other studies conducted elsewhere in Ethiopia also indicated the dominance of shrubs (Ermias Leulekal *et al.*, 2008; Abrha Tesfay, 2008; Anteneh Belayneh, 2006). However, studies conducted elsewhere in Ethiopia noted that herbs were dominating (Nurya Abdurhman, 2010; Abraha Teklay *et al.*, 2013; Moravec *et al.*, 2014; Gebremedhin Gebrezgabiher *et al.*, 2013). Herbs are more easily accessible from the nearby areas than trees and shrubs that are often harvested from patches of forests distantly located from resident areas (Ermias Leulekal, 2014).

High numbers of medicinal plants were used in the treatment of febrile illnesses (6 species) and this may suggest the high prevalence of the ailment in the study area. The fact that *Eucalyptus globulus* was the most frequently used plant to treat febrile illness could indicate better efficacy of the plant.

Leaves were the most used plant part in the preparation of remedies in the district as compared to other parts. Many studies conducted elsewhere in Ethiopia also showed the dominance of leaves in the preparation of remedies (Abraha Teklay *et al.*, 2013; Nurya Abdurhman, 2010; Girmay Zenebe *et al.*, 2012; Mirutse Giday and Gobena Ameni, 2003). This practice helps to reduce the rate of threat on plant species or helps for sustainable harvesting of plants as compared with utilization of roots. However, studies conducted elsewhere in the country showed the most common use of roots in the preparation of remedies (Moravec *et al.*, 2014; Ermias Leulekal, 2014). The popularity of roots might be due to an easy way of their use, as they were mainly chewed or put on fire and inhaled. Roots can be also dried and powdered or homogenized with water. The extensive root excavation can be devastating and can lead to a threat of the species survival or extinction (Moravec *et al.*, 2014; Ermias Leulekal, 2014).

The result showed that most remedies were prepared from fresh materials only. Some remedies were prepared from either dried or fresh materials while few were prepared from dried materials only. The frequent use of freshly collected medicinal plant could be attributed to the widespread traditional belief of attaining high efficacy from fresh remedies due to the higher presence of active ingredients in the cases of fresh plant parts, which community members thought, could be lost on drying. Considerable preparations were made from a mixture of different plant species with water and different additive substances, namely honey, butter, water and yogurt. A study conducted elsewhere in Ethiopia (Abraha Teklay *et al.*, 2013; Nurya Abdurhman, 2010; Teshale Sori *et al.*, 2004) also reported similar results.

Most informants reported the use of measuring units, such as cup, spoon, drops and fingers, but still, there were differences in doses. The variation in quantity, unit of measurement and duration

of treatment of prescribed plant preparations was also noted in a study conducted elsewhere in Ethiopia (Abraha Teklay *et al.*, 2013).

Overdose of remedies was reported to bring adverse effects, like vomiting, diarrhea, burning sensations and sometimes fainting of the patient. Problem of precision and standardization taken as one drawback for the recognition of the traditional health care system (Dawit Abebe, 2001; Ermias Leulekal, 2014; Abraha Teklay *et al.*, 2013). However, there were antidotes to the side effects caused doses problems, like honey, butter, water and yogurt. This finding is consistent with reports elsewhere in the country (Mirutse Giday *et al.*, 2010; Ermias Leulekal, 2014).

The knowledge of some young people may be related to their lack of interest in plant-based medicines, mainly because of their lack of credibility; some believe that modern medicine is more efficient than traditional medicine. Similar findings were reported in studies conducted on other cultural groups in Ethiopia (Mirutse Giday *et al.*, 2009; Ermias Leulekal, 2014). Furthermore, most of the knowledgeable people keep the knowledge secret and within their family members and this may put the continuity of medicinal plants and associated indigenous knowledge in question might be taken as a guarantee for social respect, effectiveness of healing and means of income. Other researchers reported similar findings elsewhere in the country (Mirutse Giday *et al.*, 2009; Tilahun Teklehaymanot, 2009; Ermias Leulekal, 2014).

5.1.3.3.2 Efficacy, healing potential and ranking of medicinal plants

The calculated ICF results indicated the highest agreements among informants on the use of human medicinal plant species for treating dermatological ailments and external injuries, bleeding and snakebites. The observed highest informant agreement coupled with high plant use citations for these ailment categories could also indicate the relatively high incidence of the ailments in the area. High ICF values were important in identifying plants of particular interest in the search for bioactive compounds (Heinrich *et al.*, 1998; Ermias Leulekal, 2014).

The reported highest fidelity level values were for *Verbascum sinaiticum* and *Withania somnifera* against dermatological ailments; and sensory and nervous system ailments, respectively. Fidelity level values could be used as a clue to identify medicinal plants of high healing potential and those needed for further phytochemical and bioactivity studies (Trotter and Logan, 1986; Heinrich *et al.*, 1998).

Informants ranked the plants and identified the most-preferred medicinal plant species to treat febrile illness. Accordingly, *Eucalyptus globulus*, *Withania somnifera* and *Olea europaea* subsp. *cuspidata* scored highest values and were indicated as the most-preferred ones to treat febrile illness.

5.1.3.3.3 Comparison between different gender and social groups

The results revealed that men had better medicinal plant knowledge than women and this could probably be due to the reason that boys were usually selected in the study area for the proper transfer of the knowledge. In addition, this may also be attributed to the common use of

medicinal plants in the area, which were less accessible to women. Other studies conducted elsewhere demonstrated similar findings (Begossi *et al.*, 2002; Collins *et al.*, 2006; Tilahun Teklehaymanot, 2009). Teferi Gedif and Hahn (2002) stated that parents in Ethiopia prefer to pass their traditional medical knowledge more to sons than to daughters. However, a study conducted elsewhere in Ethiopia demonstrated that there is no significant difference in medicinal plant knowledge between men and women (Hareya Fassil, 2003; Ermias Leulekal, 2014). By contrast, Voeks and Leony (2004) stated that on women are considerably more knowledgeable about the local healing flora than men. Consequently, elderly female achieve considerable community prestige, because of their healing abilities. Generally, gender-based differences in medicinal plant knowledge can be derived from experience and degree of cultural contact with curative plants (Voeks, 2007).

The average number of medicinal plants reported by different age groups in this investigation yielded very highly significant results. As the age of the informant increased, so did the person's knowledge about medicinal plants. Clearly, older people have had more time to assimilate knowledge about medicinal plants and their healing properties than younger people (Silva *et al.*, 2011; Voeks and Leony, 2004). Moreover, the treatment of health problems often falls more on the elder members of a community, who have more time at their disposal and have during their lives been exposed to many more health crises than their younger counterparts. The impact of modernization (including urbanization and the advent of formal education) and the very poor system of sharing indigenous knowledge (through word of mouth, with maximum secrecy and only along family lines) on medicinal plants with the younger generation were the sources of difference in medicinal plants. The scenario is the same for other cultural groups in Ethiopia (Mirutse Giday *et al.*, 2009; Tilahun Teklehaymanot, 2009; Ermias Leulekal 2014) and

elsewhere in the world (Begossi *et al.*, 2002; Uniyal *et al.*, 2006; Voeks and Leony (2004); Silva *et al.*, 2011). The absence of continuous cultural interaction with plants was also reported as one factor for loss of traditional knowledge through generations (Reyes-García *et al.*, 2007; Winter and McClatchey, 2008). The much-used symbol that the death of each elder is like burning a book on traditional medicine (Voeks and Leony, 2004).

The other significant difference between key & general and literate & illiterate informants could relate to the impact of age-old experience and the maximum degree of secrecy in using medicinal plants on the former and modernization on the latter. Similar results were reported by Teferi Gedif and Hahn (2002), Mirutse Giday *et al.* (2009), Tilahun Teklehaymanot (2009) and Ermias Leulekal (2014). Furthermore, community members who have greater contact with medicinal plants were more knowledgeable about therapeutic uses of these plants than those with only intermittent contact (Voeks, 2007).

5.1.3.3.4 Source and transfer of indigenous knowledge

It was widely manifested in the study area that the sources of indigenous medicinal knowledge were mainly parents and grandparents. Such knowledge has frequently been transferred to the eldest son and trusted family members. It was transferred orally from generation to generation and kept secret as it was shared only within the family circle. Other studies conducted elsewhere in Ethiopia also indicated similar results (Nurya Abdurhman, 2010; Ermias Leulekal, 2014). However, currently, there were problems in interest to learn to the indigenous knowledge carefully and use properly owing to the expansion of modern education and healthcare centers.

5.1.3.4 Multipurpose use of forest resources

The forest and forest trees were the main source of seasonal food for the local communities, particularly children and men looking after cattle and collecting fuelwood. Plants that produced edible berries, such as *Opuntia ficus-indica*, *Rhus glutinosa*, *Carissa spinarum* and *Myrsine africana* were consumed as seasonal food. Fruits were the primary preference for local communities as food, because they can simply be consumed uncooked. Similar results were reported elsewhere in the country (Abrha Tesfay, 2008; Ermias Leulekal, 2014; Getachew Addis *et al.*, 2005).

Zemedede Asfaw and Mesfin Tadesse (2001) stated that eight percent of the higher plants of Ethiopia were edible, where 203 wild edible plants with fruits constituted the highest edible parts. However, species richness in the study area was low because of deforestation and lack of interest to continually add wild edible plants to their culture. Tengnas (1994) indicated that food from trees makes supplemental, seasonal and emergency contribution to household food supplies, particularly during drought, famine and war.

According to Getachew Addis *et al.* (2005), the purpose of wild edible plants used by local people was to add dietary variety because of their good taste, as snacks during hungry periods of the day, seasonally during periods of scarcity or extreme famine, to moisten the mouth in the absence of drinking water. The most preferred plant species in the study area were *Opuntia ficus-indica*, *Ficus sur* and *Carissa spinarum* because of their high quantity and quality of fruit part, good taste, availability and easy to consume. However, currently, *Opuntia ficus-indica* has been infested by the cochineal insect that greatly affects its productivity. The dependence on wild edible plants in the study area has been decreasing from, because of modernization (with the young generation not wanting to use them as food), the less availability of the plants, fewer

occurrences of famine and war and deprivation go into the forest. The importance of wild plants as food is believed to have gradually diminished with the advent of cultivation and modernization (Zemedu Asfaw and Mesfin Tadesse, 2001).

According to the informants, the local communities have relied on plants as sources of energy. Stems and branches of woody plants were mainly used as source of fuelwood. Abbiw (1990) reported that 90% of the harvested wood is used as fuel. It is generally known that the vast majority of rural households in Ethiopia depend on wood or farm residues as fuel for cooking and heating (Abrha Tesfay, 2008). The study noted that men, women and children (shepherds) performed the collection of fuelwood. Similar findings were reported elsewhere in the world (Maundu *et al.*, 2001; Mushongi, 2001). However, a study reported in Dess'a forest reported that only men had performed the collection of fuelwood (Abrha Tesfay, 2008). The most preferred plant species in the study area were *Olea europaea* subsp. *cuspidata* and *Acacia etbaica* because of easy burning, energy content, power to produce charcoal and availability. Although the majority of wild woody plant species can be used as a source of fuel by indigenous people, many species are recognized for particular burning qualities (Cotton, 1996). These species (*Olea europaea* subsp. *cuspidata* and *Acacia etbaica*) were also preferred elsewhere in Africa (Maundu *et al.*, 2001). However, a study conducted in and around the semi-arid Awash National Park reported that *Acacia tortilis* and *Acacia nilotica* are the most preferred wood plants, and those used for household consumptions and income generation (Tinsae Bahru *et al.*, 2012). An inefficient and wasteful method of traditional open fire cooking accounts mainly for the consumption of a relatively higher proportion of fuel wood. If open fires were replaced by fuel saving stoves, fuel wood consumption could be reduced by one third by the more efficient use of

fuel wood and in part also by a change in heating and cooking habits, fuel wood consumption in the area could be effectively cut down (Cotton, 1996).

Agricultural practices required certain traditional techniques, including the use of tools and implements due to steep and hilly terrain comprising shallow and stony soils. The high quality of timber wood is always in great demand for construction, making of agricultural implements and as handles of harvesting tools. The main occupation of people residing in the study area is traditional agriculture, which acts as a major source of income. From time immemorial, local plants have been used in making agricultural implements; handles of harvesting tools and their parts. Forest ecosystems have close linkages with traditional farming systems. The local farmers used traditional farm implements, such as plow and yoke for crop production. Forest was the backbone of the livelihood of the farming community, because it provided materials for farm implements. Traditional' agriculture driven rural communities in the plains or more 'traditional' upland societies practicing traditional multi-species complex agro ecosystems are linked with the forest resources on which they depend for a variety of their needs (Ramakrishnan, 1999).

As traditional constructions of houses are common in the countryside in Ethiopia, local communities residing in the study area have used forest resources for construction of traditional house, such as “Hidmo” and “Sekella.” The roof of the traditional stone house “Hidmo” and traditional huts “Sekella” were built with poles and sticks and plastered all round with a mixture of cow dung, straw and soil. These traditional houses were wasteful as they consumed a large amount of wood (Abrha Tesfay, 2008). Plant parts, especially stems and sheets of bark or split wood, were used in roof construction in traditional dwellings (Abbiw, 1990). The roofing plant materials can be chosen according to functional properties, like durability and for being water-

proof (Abbiw, 1990). Forest resources have a paramount role in building a shelter for the local community. Similar findings in Ethiopia reported elsewhere (TFAP, 1996; Maundu *et al.*, 2001; Getachew Desalegn *et al.*, 2003).

Fodder for domestic animals is of vital importance in a livestock keeping society. The traditional livestock feeding system made use of local resources, like tree leaves and pods from the forest. In a place where there was a scarcity of rainfall and soil moisture, trees and shrubs have utmost role due to better tolerance of unfavorable conditions. Tree fodder is particularly important as animal feed in the late dry season (April – June). Similar results were reported in studies conducted in Tigray and Afar regions (Gebremedhin Hadera, 2000; Abraha Tesfay, 2008; Emiru Berhane *et al.* 2014).

Forest plants were an important source of nectar and pollen for honeybees and as beehives. The forest was used as a source of honey for some local people as they kept their beehives inside the forest. Honey in the study area was harvested almost two times a year, namely in October and June during good climatic conditions. *Podocarpus falcatus* and *Ficus sur* were the most preferred plants to use as beehive inside the forest (Appendix 20). There were common occurrences that the so-called former settlers had much beehive trees transferred through generation without conflict of interests. The fact that they kept the beehive in the forest was to provide all the necessary amenities for the honeybees in order to produce much quantity and quality honey from the diversified plants and get water found in the forest. This result agrees with researches conducted elsewhere in Ethiopia (Gebremedhin Hadera, 2000; Abraha Tesfay, 2008; Hussien Adal, 2014). The most preferred plant species in the study area were *Erica arborea*, *Hypoestes forskalii*, *Leucas abyssinica* and *Becium grandiflorum* because of the

copious nectar and pollen production as well as their pleasant flowers they produce that attract honeybees.

The widely used multipurpose plants in the forest were *Olea europaea* subsp. *cuspidata*, *Rhus glutinosa*, *Podocarpus falcatus*, *Teclea nobilis* and *Juniperus procera*. However, study conducted in Dess'a forest (Abrha Tesfay, 2008) reported *Olea europaea* subsp. *cuspidata* followed by *Acacia etbaica*, *Boscia salicifolia*, *Erica arborea*, *Cordia africana*, *Tarchonanthus camphoratus* and *Teclea nobilis* as multipurpose plants. As it is described in FAO (2011), forests by their nature are multipurpose resources.

The cultural significance index of species is used to record the role of plants in a culture (Silvia *et al.*, 2006). Hoffman and Gallaher (2007) noted that many relative cultural importance indices pool in the unique uses of plant species mentioned by informants into major use categories. Plants frequently cited for uses that differ only slightly, thereby receiving exaggerated outlier of relative cultural importance values (Albuquerque *et al.*, 2014). Standardized categorization facilitates compilation, comparison, as well as the efficient presentation of the whole data sets identifying that plant species is relatively the most important and which species is relatively the most preferred. In the study area, *Olea europaea* subsp. *cuspidata*, *Podocarpus falcatus*, *Acacia etbaica*, *Juniperus procera* and *Balanites aegyptiaca* were the most important plants.

5.1.3.5 Major threats and diminishing forest resources

Despite the attempts made by the local community and the government (regional and federal) to conserve the forest, the forest area was severely depleted. The major forest destruction activities were deforestation, agricultural expansion, fuelwood collection, livestock grazing, drought and settlement in decreasing order. Fast population growth in the study area and the demand for extra

land for agriculture, grazing and settlement were the major threats for forest depletion. Moreover, demand for construction material, fuelwood and farm implement has also increased. An increasing demand for arable land and forest products due to the rapidly growing population and poverty of the rural people were the major threats to the survival of many of the Ethiopian plant species (Ensermu Kelbessa *et al.*, 1992). If present trends in population growth continue, deterioration of natural resources will be more rapid in the future and will result in a great loss of biodiversity (Ensermu Kelbessa *et al.*, 1992; Zerihun Woldu, 1999). Alternative resource management systems that involve local people—the key generators, custodians and promoters of local biodiversity becomes useful (Rastogi *et al.*, 1998) and this helps in the effective joint-management of resources between government and local people.

Over-exploitation of plants for various reasons in the study area led to the depletion of some species. Currently, many households have moved and settled at the edge and even inside the forest in order to be closer to the resources they need for their livelihood. Traditional people all over the world, through their indigenous knowledge know which plant species are threatened and need priority conservation (Given, 1994; Aumeeruddy-Thomas and Pei, 2003). Accordingly, *Olea europaea* subsp. *cuspidata*, *Erica arborea*, *Podocarpus falcatus*, *Juniperus procera* and *Ficus sur* were the most locally threatened plants in the study area. The 1997 IUCN Red List of Threatened Plants (Walter and Gillett, 1998) included 163 (2.5%) species from Ethiopia and Eritrea, with 48 woody species. Due to heavy pressure on *Juniperus procera*, the Food and Agriculture Organization (FAO) has listed it as a threatened species that requires *in-situ* conservation priority (FAO, 1975). In addition, it is listed under the Red List of the IUCN threatened species that requires *in-situ* conservation priority (World Conservation Monitoring Centre, 1998). Dry evergreen Afromontane forest and grassland complex (DAF) is the second

richest vegetation type after *Acacia - Commiphora* woodland and bushland (ACB), which characterized by *Olea europaea* subsp. *cuspidata* and *Juniperus procera* (Friis *et al.*, 2010). DAF has been intensely utilized by man for a long while, with the result that the forests have diminished in area, and are replaced by bushland in most areas (Zerihun Woldu, 1999). Hence, the *Olea europaea* subsp. *cuspidata* – *Juniperus procera* community type contains high species richness, contains many rare species and harbors considerable number of non-timber forest products that needs conservation priority.

According to Plotkin (1995), indigenous people living in forests know best on the use and protection of biodiversity. However, the survival and existence of indigenous people and their long-term accumulated knowledge faces challenges, because of modernization, genetic erosion on plant and animal resources, low recognition of their knowledge and varied culture, loss of biodiversity (Martin, 1995; Balick and Cox, 1996; Thrupp, 1997; Almaz Negash, 2001; Bussmann, 2006). Indigenous people are also suffering from the displacement and restriction of using their local ecosystems and resources, because of the establishment of strictly protected areas (Furze *et al.*, 1997) which are believed to be one factor for the encroachment and destruction of most of the protected areas.

5.2 Conclusion

This study found that the forest has a considerable number of plant species of high diversity with 264 recorded species and five community types namely: *Cadia purpurea* – *Carissa spinarum*, *Olea europaea subsp. cuspidata* – *Juniperus procera*, *Dodonaea angustifolia* – *Acacia abyssinica*, *Erica arborea* – *Myrsine africana* and *Acacia etbaica* – *Acacia tortilis*. Among the five community types, three communities (*Olea europaea subsp. cuspidata* – *Juniperus procera*, *Dodonaea angustifolia* – *Acacia abyssinica*, *Erica arborea* – *Myrsine africana*) have overlapping altitudinal range to a certain extent while the remaining two community types (*Acacia etbaica* – *Acacia tortilis* and *Cadia purpurea* – *Carissa spinarum*) were found at low elevations isolated from the others and with relatively less similarity among them. It was found that the distribution of plant communities was influenced by altitude, slope and disturbance (human and grazing by livestock).

Studies of population structure showed a high proportion of small-sized individuals in the lower diameter classes indicating high regeneration potential of the forest. However, there were large variations among different tree species. For instance, some tree species have skewed population structure with no or few individuals at a lower size class, indicating poor regeneration or selective grazing or cutting. Overall, the population structure of the species investigated fall into four population density structure, namely Inverted-J shaped, bell-shaped, Broken J shape and upward 'F'. Assessment of regeneration status based on age classes indicated that a significant proportion of woody species were represented by few or no seedlings, indicating that the species were under threat.

Seed bank studies are known tools for assessing restorative potential of an ecosystem and will be essential for developing appropriate forest restoration strategies. The study on soil seed bank through germination experiment showed that the soil seed bank was mostly dominated by the herbaceous flora and in terms of seed bank distribution, the upper 5 cm soil has high number of species when compared with the lower 5 cm soil. The study showed high variability of soil seed bank density and emerging vegetation among soil samples collected from the sampled quadrates. Overall, the diversity of woody species was very low where only eight woody species germinated from an experiment that run for almost a year. From this result, it was found that the soil seed bank might not be relied upon for restoration of the forest. This means passive restoration approach, such as enclosure may not be sufficient for restoration of the forest. Thus, any restoration attempt needs to take into consideration active restoration approaches, such as growing of seedlings from seeds and planting them in the degraded area of the forest. In addition, there may be a potential of seed rain from standing vegetation or seed dispersal by birds or other vectors from the surrounding.

It is an established fact that fringe communities on the edge of the forest use forests near them for various purposes. Even though the study forest is a protected forest by guards employed by the regional government, there were illegal grazing and cutting. From survey conducted on the local community, it was found that the forest provided important ecosystem services for the community through provision of medicinal plants, wild edible plants, fuelwood, livestock fodder/forage, honeybee forage, farm implements and building material. However, there were threats posed by the local community on the forest due to poverty and lack of awareness, agricultural expansion, deforestation, fuelwood collection and settlement. In addition, it is

concluded that the traditional ecological knowledge of the community is threatened due to lack of documentation. Knowledge has been orally transmitted from generation to generation and this is also shown in high disparity among the present generation. This sort of study that document forest resources and indigenous knowledge's is important for restoring the forest to its original state and raising awareness among the community.

Finally, *Wejig-Mahgo-Waren* massif forest has relatively a considerable plant species richness and diversity in spite of the threats it faces and with the paramount importance of the forest for the fringe communities. However, it requires immediate and active intervention since the restoration potential of the forest is questionable as this study indicates.

5.3 Recommendations

❖ Based on the findings of the research, the following recommendations are forwarded:

1. From the thirteen endemic species found in the forest, six of them: *Becium grandiflorum*, *Leucas abyssinica*, *Lippia adoensis*, *Rhus glutinosa*, *Vernonia leopoldii* and *Vernonia rueppellii*, are registered in the IUCN Red List Categories. Hence, responsible authorities, such as the Ethiopian Biodiversity Institute (EBI) are advised to devise appropriate conservation measures (*in situ* or *ex situ*) to save these valuable plant species.
2. It is important to follow the ecosystem strategy of biodiversity conservation that integrates land, water and living resources. In the study area, to conserve a large number of species and plant communities, it is worth establishing a biodiversity conservation corridor along the landscape of *Wejig-Mahgo-Waren* forest massif. However, with the current socio-economic context in the study area, it may be difficult to implement some of the recommendation stated above. Dry evergreen Afromontane forest and grassland

complex (DAF) is the second richest vegetation type, which is characterized by *Olea europaea* subsp. *cuspidata* and *Juniperus procera* and harbored considerable number of non-timber forest products. Due to heavy pressure on *Juniperus procera*, the Food and Agriculture Organization (FAO) and Red List of the IUCN has listed it as a threatened species. Thus, it is important to prioritize conservation sites. The *Olea europaea* subsp. *cuspidata* – *Juniperus procera* community contained many locally threatened and rare species that requires *in-situ* conservation priority.

3. For plant species with irregular population structure and reduced regeneration status, there is a need to establish nursery sites to prevent local extinction by planting seedlings. The species *Cadia purpurea*, *Bersama abyssinica*, *Allophylus abyssinicus*, *Hagenia abyssinica*, *Ekebergia capensis*, *Ficus sur*, *Dombeya torrida*, *Celtis africana* and *Podocarpus falcatus*.
4. Illegal grazing and cutting mainly at the edges of the forest were found to be the principal drivers for deforestation, and this requires intervention by federal and regional governments in consultation with the community to put in place a long-term plan and strategy to reduce livestock density within and around the forest area to achieve sustainable utilization of forest resources.
5. Since the restoration potential from soil seed bank is limited, to restore the forest in the shortest possible time, restoration strategy that combines active restoration, planting of indigenous seedlings locally threatened plant species, such as *Olea europaea* subsp. *cuspidata*, *Erica arborea*, *Podocarpus falcatus*, *Juniperus procera* and *Ficus sur* and passive restoration techniques, (e.g. exclosures) from the available soil seed bank should be applied. Effort should also be made by the author of this work.

6. The ethnobotanical survey for many species is the preferred strategy for drug discovery. This approach assumes that species contain healing abilities as employed and demonstrated by traditional healers; especially promising medicinal plants should be pursued further. The local community relies on traditional herbal medicine in the area. However, since dosage of use varies from person to person, this practice poses a health problem to the community. Therefore, it is important to take those plants that have the highest FL, ICF and PR values and that require further phytochemical and bioactivity tests. The following plants seem to be the most promising plants obtained from both FL and PR that need extensive investigation: *Verbascum sinaiticum*, *Withania somnifera*, *Eucalyptus globulus*, *Olea europaea* subsp. *cuspidata* and *Datura stramonium*.
7. The fact that forest resources used as fuelwood are the cheapest and accessible energy source in the study area, the wasteful method of traditional open fire cooking are common that consume huge woody species which resulted in deforestation. Hence, to combat the problem of deforestation, there is a need for fuel saving stoves, as well as distribution of electricity in each household by the regional and federal government.
8. Because of poverty, the community relies on the forest resources for most of the activities that caused depletion of forest resources. Community-based conservation of the forest must be done through raising awareness.

References

- Abbiw, D. K. (1990). Useful plants of Ghana: West African uses of wild and cultivated plants. Intermediate technology publications, London and the Royal Botanic Gardens, UK, p. 337.
- Abiyot Berhanu (2017). Vegetation Ecology and Conservation Status of Evergreen Afromontane Forest Patches in Awi Zone of Amhara Region, Northwestern Ethiopia. PhD Dissertation, Addis Ababa University, Ethiopia.
- Abiyot Tilahun, Teshome Soromessa, Ensermu Kelbessa and Abiyot Dibaba (2011). Floristic composition and community analysis of Menagesha Amba Mariam Forest (Egdu Forest) in central plateau of Shewa, Ethiopia. *Ethiopian Journal of Biological Sciences* **10(2)**: 111-136.
- Abraha Teklay (2015). Traditional medicinal plants for ethnoveterinary medicine used in Kilte Awulaelo District, Tigray Region, Northern Ethiopia. *Advancement in Medicinal Plant Research* **3(4)**: 137-150.
- Abraha Teklay, Balcha Abera and Mirutse Giday (2013). An Ethnobotanical Study of Medicinal Plants Used in Kilte Awulaelo District, Tigray Region of Ethiopia. *Journal of Ethnobiology and Ethnomedicine* **9**:65.
- Abraha Tesfay (2008). Ethnobotanical Study of Dess'a Forest, Northeastern Escarpment of Ethiopia, With Emphasis on Use and Management of Forest Resources by the Local People. MSc. Thesis. Addis Ababa University, Ethiopia.
- Albuquerque, U. (2009). Quantitative Ethnobotany or Quantification in Ethnobotany? Available: on-line at: [http:// www. Ethnobotany journal.org/vol5/i1547-3465-07-001](http://www.Ethnobotanyjournal.org/vol5/i1547-3465-07-001).
- Albuquerque, U. P., Cunha, L. V. F. C., Lucena, R. F. P. and Alves, R. R. N. (2014). Methods and Techniques in Ethnobiology and Ethnoecology Springer Science + Business Media New York, USA
- Alemayehu Wassie (2007). Ethiopian Church Forests: opportunities and challenges for restoration. PhD. Thesis, Wageningen University, Wageningen, Netherlands.
- Alemnew Alelign, Demel Teketay, Yonas Yemshaw and Edwards, S. (2007). Diversity and status of regeneration of woody plants on the Peninsula-Zegie, northwestern Ethiopia. *Tropical Ecology* **48(1)**: 37-49.
- Alexiades, M. N. (1996). Selected Guidelines for Ethnobotanical Research: A Field Manual. The New York Botanical Garden. New York, pp. 295.

- Almaz Negash (2001). Diversity and conservation of Enset (*Ensete ventricosum* Welw. Cheesman) and its relation to household food and livelihood security in south- western Ethiopia. PhD Disseration. Wageningen University.
- Amanuel Zenebe; Girmay Gebresamuel and Atkilt Girma (2015). Characterisation of Agricultural Soils in Cascape Intervention Woredas in Southern Tigray, Ethiopia; Mekelle University, Ethiopia.
- Amiaud, B. and Touzard, B. (2004). The relationships between soil seed bank, aboveground vegetation and disturbances in old embanked marshlands of Western France. *Flora* **199**: 25-35.
- Anteneh Belayneh (2006). Floristic Description and Ethnobotanical Study of the Natural Vegetation in the Babile Elephant Sanctuary, Ethiopia. MSc. Thesis in Dryland Biodiversity. Addis Ababa University, Ethiopia.
- Aumeeruddy-Thomas, Y. and Pei, S. (2003). Applied Ethnobotany: case-studies from the Himalayan region. People and Plants working paper 12. WWF, Godalming, UK.
- Ayyad, M.A. (2003). Case studies in the conservation of biodiversity: degradation and threats. *Journal of Arid Environments* **54**: 165-182.
- Balick, M. J. and Cox, P. A. (1996) Plants, People and Culture. The science of ethnobotany. Scientific American Library, New York.
- Bard, K. A., Coltorti, M. and Di-Blasi, M. C. (2000). The environmental history of Tigray (northern Ethiopia) in the middle and late Holocene: A preliminary outline. *African Archaeological Review* **17(2)**: 65-86.
- Bartlett, J. E., Kotrlik, J. W. and Higgins, C., (2001). Organizational research: Determining appropriate sample size for survey research. *Journal of Information Technology, Learning and Performance* **19(1)**: 43-50.
- Baskin, C. C. and Baskin, J. M. (1998). Seeds: Ecology, Biogeography and Evolution of Dormancy and Germination. Academic Press, San Diego, California, USA.
- Begossi, A., Hanazaki, N. and Tamashiro, J. Y. (2002). Medicinal plants in the Atlantic Forest (Brazil): knowledge, use and conservation. *Human Ecology* **30**: 281-299.
- Bennett, B. C. (1992). Plants and people of the Amazonian rainforests. *BioScience* **42**: 599-607.
- Bernhardt, K. G., Koch, M., Kropf, M., Ulbel, M. and Webhofer, J. (2008). Comparison of two methods characterising the seed bank of amphibious plants in submerged sediments. *Aquatic Botany* **88**: 171–177.

- Bobe, R. (2006). The evolution of arid ecosystems in eastern Africa. *Journal of Arid Environments* **66**: 564–584.
- Bossuyt, B. and Hermy, M. (2001). Influence of land use history on seed banks in European temperate forest ecosystems: a review. *Ecography* **24**: 225-238.
- Breitenbach, F.V. (1963). The Indigenous trees of Ethiopia. 2nd revised and enlarged edition, Ethiopian Forestry Association, Addis Ababa.
- Brooks, T. M., Mittermeier, R. A. and Mittermeier, C.G. (2002). Habitat Loss and Extinction in the Hotspots of Biodiversity. *Conservation Biology* **16(4)**: 909-923.
- Brown, D. (1992). Estimating the composition of a forest seed bank: a comparison of the seed extraction and seedling emergence methods. *Canadian Journal of Botany* **70**: 1603–1612.
- Burger, W.C. (1974). Flowering periodicity at four levels in eastern Ethiopia. *Biotropica* **6**: 38-42.
- Bussmann, R. W. (2006). Ethnobotany of the Samburu of Mt. Nyiru, South Turkana, Kenya. Bussmann, licensee BioMed Central Ltd. *Journal of Ethnobiology and Ethnomedicine* **2**: 35.
- Callaway, R. M. (1997). Positive interactions in plant communities and the individualistic continuum concept. *Oecologia* **112**: 143-149.
- Capon, S. J. and Brock, M.A. (2006). Flooding, soil seed bank dynamics and vegetation resilience of a hydrologically variable desert floodplain. *Freshwater Biology* **51**: 206–223.
- Castaneda, H. and Stepp, J. R. (2007). Ethnoecological Importance Value (EIV) Methodology: Assessing the Cultural Importance of Ecosystems as Sources of Useful Plants for the Guaymi People of Costa Rica.
- Cincotta, R. P., Wisniewski, J. and Engelman, R. (2000). Human population in the biodiversity hotspots. *Nature* **404**: 990-992
- Clements, F. E. (1916). Plant succession: An analysis of the development of vegetation Publication No. 242, Carnegie Inst., Washington, DC.
- Cochran, W. G. (1977). Sampling techniques (3rd ed.) New York: John Wiley and Sons.
- Colding, J. and Folke, C. (2001). Social taboos: “Invisible” systems of local resource management and biological conservation. *Ecological Applications* **11**: 584–600.
- Collins, S., Martins, X., Mitchell, A., Awegechew Teshome and Arnason, J. T. (2006). Quantitative ethnobotany of two East Timorese cultures. *Economic Botany* **60**: 347-361.
- Conservation International (2007). Biodiversity hotspots. <http://www.biodiversityhotspots.org/xp/hotspots/caucasus/Pages/default.aspx>. Cited 4 Oct 20015.

- Cotton, C. M. (1996). *Ethnobotany: Principles and Applications*. John Wiley and Sons Ltd. London. England. pp. 402.
- Central Statistical Agency (2007). Tables: Tigray Region, Ethiopia (accessed 30 December 2008)
- Cui, L., Li, W., Zhaoa, X., Zhanga, M., Lei, Y., Zhanga, Y., Gaoa, C., Kanga, X., Suna, B., Zhanga, Y. (2016) The relationship between standing vegetation and the soil seed bank along the shores of Lake Taihu, China. *Ecological Engineering* **96**: 45–54.
- Cunningham, A. B. (1996). People, Park and Plants use recommendations for multiple use zones and development alternatives around Bwindi: Impenetrable National Park, Uganda. In: *People and Plants*, working paper four, Sample, A. (ed.), UNESCO, Paris, pp. 18-23.
- Curtis, J. T. and McIntosh, R.P. (1951). An upland Forest Continuum in the prairie forest border region of Wisconsin. *Ecology* **32**: 476-479.
- Dainou, K., Bauduin, A., Bourland, N., Gillet, J. F., Feteke, F. and Doucet, J. L. (2011). Soil seed bank characteristics in Cameroonian rainforests and implications for post-logging forest recovery. *Ecological Engineering* **37**: 1499-1506.
- Darbyshire, I., Lamb, H. and Umer, M. (2003). Forest clearance and regrowth in northern Ethiopia during the last 3000 years. *The Holocene* **13**: 537-546.
- Dawit Abebe (2001). The role of medicinal plants in healthcare coverage of Ethiopia, the possible benefits of integration. In: Medhin Zewdu, Abebe Demissie (eds). *Proceedings of Workshop on Biodiversity Conservation and Sustainable Use of Medicinal Plants in Ethiopia*. Institute of Biodiversity Conservation and Research, Addis Ababa, Ethiopia, 28 April –May 2001, pp. 6-21.
- Daws, M. I., Garwood, N. C. and Pritchard, H. W. (2005). Traits of recalcitrant seeds in a semi-deciduous tropical forest in Panama: some ecological implications. *Funct. Ecol.* **19**: pp. 874-885.
- DCR-DNH (2011). Outline of procedures for data collection using the standard DCR-DNH plot form. Virginia: Virginia Department of Conservation and Recreation / Division of Natural Heritage. pp. 1-15.
- Decocq, G., Valentin, B., Toussaint, B., Hendoux, F., Saguez, R. and Bardat, J. (2004). Soil seed bank composition and diversity in a managed temperate deciduous forest. *Biodivers. Conserv.* **13**: 2485-2509.
- Demel Teketay (2002). Germination responses of *Discopodium penninervium* to temperature and light. *Flora* **197**: 76–80.
- Demel Teketay and Tamrat Bekele (1995a). Floristic composition of Wof-Washa natural forest, Central Ethiopia: Implications for the conservation of biodiversity. *Feddes Repertorium* **106**: 127–147.

- Demel Teketay and Granstrom, A. (1995b). Soil seed banks in dry afro-montane forests of Ethiopia. *Vegetation Science* **6**: 777-786.
- Demel Teketay and Granström, A. (1997). Germination ecology of forest species from the highlands of Ethiopia. *Journal of tropical ecology* **13(6)**: 805-831.
- Desalegn Wana and Zerihun Woldu (2005). Vegetation of Chenchu Highlands in Southern Ethiopia. *SINET: Ethiop. J. Sci.* **28(2)**: 109–118.
- De Villiers, A. J., Van Rooyen, M. W. and Theron, G. K. (2003). Similarity between the soil seed bank and the standing vegetation in the Strandveld Succulent Karoo, South Africa. *Land Degradat. Dev.* **14**: 527–540.
- Dormann, C. F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G., Marquéz, G., Gruber, B., Lafourcade, B., Leitão, P.J., Münkemüller, T., McClean, C., Osborne, P.E., Reineking, B., Schröder B., Skidmore, A.K., Zurell, D. and Lautenbach, S. (2012). Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. *Ecography* **35**: 001–020.
- Doucet, J. L., Kouadio, Y. L., Monticelli, D. and Lejeune, P. (2009). Enrichment of logging gaps with moabi (*Baillonella toxisperma* Pierre) in a Central African rain forest. *Forest Ecol. Manage.* **258**: 2407–2415.
- Dugdale, J. S. (1964). Ethiopian climates and vegetation: The state of our present knowledge. *Journal of Semitic Studies* **9**: 250-256.
- Edwards, S., Mesfin Tadesse and Hedberg, I. eds. (1995). *Flora of Ethiopia and Eritrea. Canellaceae to Euphorbiaceae (vol. 2:2)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 456.
- Edwards, S., Mesfin Tadesse, Sebsebe Demissew and Hedberg, I. eds. (2000). *Flora of Ethiopia and Eritrea. Magnoliaceae to Flacourtiaceae (vol. 2:1)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 332
- Edwards, S., Sebsebe Demissew and Hedberg, I. (1997). *Flora of Ethiopia and Eritrea Hydrocharitaceae to Arecaceae. (vol. 6)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp.586.
- Ethiopian Forestry Action Program (1994). The Challenge for Development. Volume III.

- Emiru Birhane, Mulubrhan Balehegn, Daniel Kiros and Diress Tsegaye (2014). Distribution, Animal Preference and Nutritive Value of Browse Species in the Rangelands of Afar, Northern Ethiopia. *Ethiop. J. Biol. Sci.* **13(2)**: 135-148.
- Ensermu Kelbessa, Sebsebe Demissew, Zerihun Woldu and Edwards, S. (1992). Some threatened Endemic Plants of Ethiopia. In: Edwards, S. and Zemedu Asfaw (eds). *The status of some plants in parts of tropical Africa*, , NAPRECA, No. 2. Botany 2000, East and Central Africa, pp. 35-55.
- Ensermu Kelbessa and Sebsebe Demissew (2014). Diversity of vascular plant taxa of the Flora of Ethiopia and Eritrea. *Ethiop. J. Biol. Sci.* **13(Supp.)**: 37-45.
- Ericksson, I., Demel Teketay and Granstrom, A. (2003). Response of plant communities to fire in an Acacia woodland and a dry Afromontane forest, southern Ethiopia. *Forest Ecology and Management* **177**: 39-50.
- Ermias Aynekulu (2011). Forest diversity in fragmented landscapes of northern Ethiopia and implications for conservation. PhD Dissertation, University of Bonn, Germany.
- Ermias Lulekal (2014). Plant Diversity and Ethnobotanical Study of Medicinal Plants in Ankober District, North Shewa Zone of Amhara Region, Ethiopia. PhD Dissertation. Addis Ababa University, Ethiopia.
- Ermias Lulekal, Ensermu Kelbessa, Tamrat Bekele and Haile Yineger (2008). An ethnobotanical study of medicinal plants in Mana Angetu District, southeastern Ethiopia. *Journal of Ethnobiology and Ethnomedicine* **4:10**.
- Ethiopian Wildlife and Natural History Society (Undated). A Glimpse at Biodiversity Hotspots of Ethiopia.. Addis Ababa, Ethiopia.
- Food and Agriculture Organization (2011). State of the World's Forests. Rome, Italy
- Farhig, L. (2003). Effects of habitat fragmentation on biodiversity. *Ann. Rev.Ecol.Syst.* **34**: 487-515.
- Fekadu Gurmessa, Teshome Soromessa and Ensermu Kelbessa (2012). Structure and regeneration status of Komto Afromontane moist forest, East Wollega Zone, west Ethiopia. *Journal of Forestry Research* **23**: 205-216.
- Fekadu Gurmessa, Teshome Soromessa and Ensermu Kelbessa (2013). Floristic composition and community analysis of Komto Afromontane moist forest, East Wollega Zone, West Ethiopia. *Science Technology and Arts Research Journal* **2(2)**: 58-69.

- Feoli, E. (1996). Rehabilitation of degraded and degrading areas of Tigray, Northern Ethiopia. *Final project report*, commission of the European Communities (CEC). pp. 411.
- Feoli, E., Vuerich, L.G. and Zerihun Woldu (2002) Evaluation of environmental degradation in northern Ethiopia using GIS to integrate vegetation, geomorphological, erosion and socio-economic factors. *Agriculture, Ecosystems and Environment* **91**: 313-325.
- Ferrier, S., Faith, D. P. and Arponen, A. (2009). Community-level approaches to spatial conservation prioritization. In Moilanen, A. *et al* (eds) Spatial conservation prioritization. Oxford University Press, New York, pp. 95-109.
- Feyera Senbeta (2006). Biodiversity and ecology of Moist Evergreen Afromontane forests with wild *Coffea arabica* L. populations in Ethiopia. PhD Dissertation. Cuvillier Verlag Göttingen. pp. 152.
- Feyera Senbeta and Demel Teketay (2001). Regeneration of indigenous woody species under the canopy of tree plantations in Central Ethiopia. *Tropical Ecology* **42**: 175-185.
- Feyera Senbeta and Demel Teketay (2002). Soil seed banks in plantations and adjacent natural dry Afromontane forests of central and southern Ethiopia. *Tropical Ecology* **43**: 229-242.
- Fitsumbirhan Tewelde, Mebrahtom Mesfin and Semere Tsewene (2017). Ethnobotanical Survey of Traditional Medicinal Practices in LaelayAdi-yabo District, Northern Ethiopia. *International Journal of Ophthalmology and Visual Science* **2(5)**: 80-87
- Friis, I. (1992). Forests and forest trees of northeast tropical Africa: Their natural habitats and distribution patterns in Ethiopia, Djibouti and Somalia. Her Majesty's Stationery Office, London.
- Friis, I. and Sebsebe Demissew (2001). Vegetation maps of Ethiopia and Eritrea. A review of Existing Maps and the need for a new map for the flora of Ethiopia and Eritrea. *Biol. Skr.* **54**: 399-439.
- Friis, I., Sebsebe Demissew and Van Bruegel, P. (2010). Atlas of the Potential Vegetation of Ethiopia. Addis Ababa: Addis Ababa University Press and Shama Books.
- Furze, B., De Lacy, T. and Birckhead, J. (1997). Culture, Conservation and Biodiversity: The social Dimension of linking local level development and conservation through protected areas. John Willey and SONS.
- Gaston, K. J. (2000). Global patterns in biodiversity. *Nature* **405**: 220-227.

- Gebremedhin Gebrezgabiher, Shewit Kalayou and Samson Sahle (2013). An Ethno-Veterinary Survey of Medicinal Plants in Districts of Tigray Region, Northern Ethiopia. *International Journal of Biodiversity and Conservation* **5(2)**: 89-97.
- Gebremedhin Hadera (2000). A study on the Ecology and Management of the Dess'a Natural Forest, in the North-Eastern Escarpment of Ethiopia. M.Sc. Thesis in Dry land biodiversity, Addis Ababa University.
- Genet Atsbeha (2012). Floristic composition of herbaceous flowering plant species with emphasis to ethnobotanical importance of wild grasses in Laelay and Tahtay Michew Districts, Central Zone of Tigray, Ethiopia. MSc. Thesis, Addis Ababa University.
- Gessesse Dessie and Kleman, J. (2007) Pattern and Magnitude of Deforestation in the South Central Rift Valley Region of Ethiopia. *Mountain Research and Development* **27(2)**: 162– 168.
- Getachew Addis, Kelbessa Urga and Dawit Dikasso (2005). Ethnobotanical study of edible wild plants in some selected districts of Ethiopia. *Human Ecology* **33(1)**: 83-118.
- Getachew Desalegn, Wubalem Tadesse, Worku Fekadu, Gemechu Kaba, Demel Teketay and Girma Taye (2003). Effectiveness of protection measures on 32 timber species against subterranean termites and fungi at Zeway research station, central Ethiopia. *Ethiop. J. Biol. Sci.* **2(2)**: 189-216.
- Getachew Tesfaye, Demel Teketay, Masresha Fetena and Erwin, B. (2010). Regeneration of seven indigenous tree species in a dry Afromontane forest, southern Ethiopia. *Flora* **205**: 135-143.
- Getinet Masresha (2014). Diversity, Structure, Regeneration and Status of Vegetation in Simien Mountains National Park, Northern Ethiopia. PhD. Thesis, Addis Ababa University.
- Getinet Masresha, Teshome Soromessa and Ensermu Kelbessa (2015). Status and Species Diversity of Alemsaga Forest, Northwestern Ethiopia. *Advances in Life Science and Technology* **34**.
- Gidey Yirga (2010a). Ethnobotanical Study of Medicinal Plants in and Around Alamata, Southern Tigray, Northern Ethiopia. *Current Research Journal of Biological Sciences* **2(5)**: 338-344.
- Gidey Yirga (2010b). Use of traditional medicinal plants by indigenous people in Mekele town, capital city of Tigray regional state of Ethiopia. *Journal of Medicinal Plants Research* **4(17)**: 1799-1804.

- Girmay Zenebe, Mohammed Zerihun and Zewdie Solomon (2012). An Ethnobotanical Study of Medicinal Plants in Asgede Tsimbila District, Northwestern Tigray, Northern Ethiopia. *Ethnobotany Research and Applications* **10**: 305-320.
- Given, D. R. (1994). Principles and practices of Plant Conservation. Chapman and Hall, London, Timber press, Inc.
- Gleason, H. A. (1926). The individualistic concept of the plant association. *Bull Torrey Bot Club* **53**: 7-26.
- Graham, M. H. (2003). Confronting multicollinearity in Ecological Multiple Regression. *Ecology* **84 (11)**: 2809-2815.
- Green, R.H. (1979). Sampling design and statistical methods for environmental biologists. Wiley-Interscience, New York, Chichester, Brisbane, Toronto.
- Greenway, P. J. (1973). A classification of the vegetation of East Africa. *Kirkia* **9**: 1 -68.
- Grenier, L. (1998). Working with Indigenous Knowledge: A guide for Researchers. Published by the IDRC (International Development Research Centre). Ottawa, Canada. pp. 115.
- Guo, Q. F., Rundel, P. W. and Goodall, D. W. (1998) Horizontal and vertical distribution of desert seed banks: patterns, causes and implications. *J. Arid Environ.* **38**: 465-478.
- Haile Yineger, Ensermu Kelbessa, Tamrat Bekele and Ermias Lulekal (2008). Floristic composition and structure of the dry Afromontane forest at Bale Mountains National Park, Ethiopia. *SINET: Ethiopian Journal of Science* **31(2)**: 103–120.
- Haileab Zegeye, Demel Teketay and Ensermu Kelbessa (2011). Diversity and regeneration status of woody species in Tara Gedam and Ababay forests, northwestern Ethiopia. *Journal of Forestry Research* **22(3)**: 315–328.
- Hareya Fassil (2003). “We do what we know”: Local Health Knowledge and Home-based Medicinal Plant Use in Ethiopia. PhD thesis. Green College, Oxford University.
- Harper, J. L. (1977). Population biology of plants. Academic press, London, pp. 892.
- Hedberg, O. (1964) Features of Afroalpine plant ecology. *Acta Phytogeographica Suecica* **49**: 1-144.
- Hedberg, I. and Edwards, S., eds. (1989): *Flora of Ethiopia and Eritrea. Pittosporaceae to Araliaceae. (vol. 3)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 659.

- Hedberg, I., Edwards, S. and Sileshi Nemomissa (2003). *Flora of Ethiopia and Eritrea Apiaceae to Dipsacaceae. (vol. 4:1)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 352.
- Hedberg, I., Friis I. and Persson, E. (2009a). *Flora of Ethiopia and Eritrea Lycopodiaceae to Pinaceae. (vol. 1)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 305.
- Hedberg, I., Friis I. and Persson, E. (2009b). *Flora of Ethiopia and Eritrea. General Part and Index to Volumes 1-7 (vol. 8)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 331.
- Hedberg, I., Kelbessa, E., Edwards, S., Sebsebe Demissew and Persson, E., eds (2006). *Flora of Ethiopia and Eritrea. Gentianaceae to Cyclocheliceae (vol. 5)*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia.
- Heinrich, M., Ankil, A., Frei, B., Weimann, C. and Sticher, O. (1998). Medicinal plant in Mexico: Healer's Consensus and Cultural importance. *J. Soc.Sci. and Med.* **47**: 1863-1875.
- Hill, J. L. and Curran, P. J. (2003). Area, shape and isolation of tropical forest fragments: effects on tree species diversity and implications for conservation. *Journal of Biogeography* **30**: 1391-1403.
- Hill, S. C. and Morris, D. M. (1992). *The Function of Seed Banks in Northern Forest Ecosystems*. Queen's Printer for Ontario, Ontario.
- Hirsch, B. T., Veronica, R. K., Pereira, E. and Jansen, P. A. (2012). Directed seed dispersal towards areas with low conspecific tree density by a scatter-hoarding rodent. *Ecology Letters* **1**: 1-7.
- Hoffman, B. and Gallaher, T. (2007). Importance indices in Ethnobotany. *Ethnobot. Res. Appl.* **5**: 201-218.
- Höft, M., Barik, S. K. and Lykke, A. M. (1999). Quantitative ethnobotany: Applications of multivariate and statistical analysis in ethnobotany. People and Plants Working Paper 6. UNESCO, Paris. pp. 45.
- Hussien Adal (2014). Plant Diversity and Ethnobotany of Borena Sayint National Park, northern Ethiopia. PhD Dissertation. Addis Ababa University, Ethiopia.

- Janzen, D. H. (1988). Tropical dry forests: the most endangered major tropical ecosystem pp. 130-37, In: Wilson EGO. and Peter F. M. (Eds.). *Biodiversity*, National Academy Press, Washington, pp. 425.
- Johnson, R. G. and Anderson, R. C. (1986). The seed bank of a tallgrass prairie in Illinois. *American Midland Naturalist* **115**: 123–130.
- Kalayu Mesfin, Gebru Tekle and Teklemichael Tesfay (2013). Ethnobotanical Study of Traditional Medicinal Plants Used by Indigenous People of Gemad District, Northern Ethiopia. *Journal of Medicinal Plants Studies* **1(4)**: 32-37.
- Kebrom Tekle and Tesfaye Bekele (2000). The role of soil seed bank in the rehabilitation of degraded hillslopes in Southern Wello, Ethiopia. *Biotropica* **32**: 23–32.
- Kebu Balemie, Ensermu Kelbessa and Zemedede Asfaw (2004). Indigenous medicinal plant utilization, management and threats in Fentalle area, Easter Shewa, Ethiopia. *Ethiop. J. Biol. Sci.* **3(1)**: 37-58.
- Kent, M. and Coker, P. (1992). *Vegetation Description and Analysis. A practical approach*. John Wiley and Sons, New York. pp. 363.
- Khan, M. L, Rai, J. P. N. and Tripathi, R. S. (1987). Population structure of some tree species in disturbed and protected sub-tropical forests of north-east India. *Acta Oecologia: Oecologia Applicata* **8**: 247-255.
- Kindeya Gebrehiwot (1997). Area enclosures as an approach in the management of dryland biodiversity: a case study in Tigray region, Northern Ethiopia. Paper presented at the workshop on Management of dryland biodiversity, 30 July-14 August, 1997, Nairobi, Kenya.
- Kitessa Hundera, Tamrat Bekele and Ensermu Kelbessa (2007). Floristics and Phytogeographics Synopsis of A Dry Afromontane Coniferous forest in The Bale Mountain (Ethiopia): Implication to Biodiversity Conservation. *SINET: Ethiop. J. Sci.* **30(1)**: 1-12.
- Komwihangilo, D. M. and Mwilawa, A. J. (2006). Enhancing utilization of exogenous and indigenous tree and shrubs in livestock production systems of semiarid areas of central Tanzania. In: Nikundiwe, M. A. and Kabigumila, D.L.J eds. *Dryland ecosystem: challenges and opportunities for sustainable Natural Resources Management*, pp. 127-131. Proceedings of the Regional Workshop held at ŠArusha, Tanzania, 7-9 June 2006, RPSUD, Dare Salaam, Tanzania.
- Krebs, C.J. (1999). *Ecological Methodology*. Addison Wiseley Longman, New York. pp. 694.

- Kreft, H. and Jetz, W. (2007). Global patterns and determinants of vascular plant diversity. *PNAS* **104(14)**: 5925-5930.
- Kumelachew Yeshitla and Tamrat Bekele (2002). Plant community analysis and ecology of afro-montane and transitional rainforest vegetation of Southwestern Ethiopia. *SINET: Ethiopian Journal of Science* **25(2)**: 155-175.
- Lamb, H., Darbyshire, I. and Verschuren, D. (2003). Vegetation response to rainfall variation and human impact in central Kenya during the past 1100 years. *The Holocene* **13(2)**: 285-292.
- Lamprecht, H. (1989). *Silviculture in the Tropics: Tropical forest ecosystems and their tree species-possibilities and methods for their long-term utilization*. Federal Republic of Germany, Eschborn.
- Legendre, P. and Legendre, L. (1998). *Numerical Ecology* (2nd English edition). Elsevier Science B.V., Netherlands. pp. 870.
- Lepš, J. and Šmilauer, P. (2001). *Multivariate Analysis of Ecological Data using CANOCO*. Cambridge Press.
- Leul Kidane (2015). *Vegetation ecology and plant diversity of Hugumburda-Gratkhassu National Forest Priority Area, Northeastern Ethiopia: habitat fragmentation in time and space*. PhD Dissertation. Addis Ababa University, Ethiopia.
- Leul Kidane, Tamrat Bekele and Sileshi Nemomissa (2010) *Vegetation Composition in Hugumburda-Gratkhassu National Forest Priority Area, South Tigray*. *MEJS* **2(2)**: 27-48.
- Li, Q., Fang, H. and Cai, Q. (2011). Persistent soil seed banks along altitudinal gradients in the Qilian Mountains in China and their significance for conservation management *African Journal of Agricultural Research* **6(10)**: 2329-2340.
- Lomolino, M.V. (2001). Elevation gradients of species-density: historical and prospective views. *Global Ecology and Biogeography* **10**: 3-13.
- Luzuriaga, A. L., Escudero, A., Olano, J. M. and Loidi, J. (2005). Regenerative role of seed banks following an intense soil disturbance. *Acta Oecol.* **27**: 57-66.
- Magurran, A. E. (2004). *Measuring Ecological Diversity*. Blackwell Science Ltd., Malden
- Mark, V. (2001). Do commonly used indices β -diversity measure species turnover? *Journal of Vegetation Science* **12**: 545-552.
- Martin, G. J. (1995). *Ethnobotany: A Methods Manual*. Chapman and Hall, New York.

- Matus, G., Papp, M. and Tothmeresz, B. (2005). Impact of management on vegetation dynamics and seed bank formation of inland dune grassland in Hungary. *Flora* **200**: 296-306.
- Maundu, P., Berger, D. J., Saitabau, C., Nasieku, J., Kipelian, M., Mathenge, S. G., Morimoto, Y. and Höft, R. (2001). Ethnobotany of the Loita Maasai: Towards Community Management of the Forest of the Lost Child - Experiences from the Loita Ethnobotany Project. People and Plants working paper 8. UNESCO, Paris.
- McCune, B. and Grace, J. B. (2002). Analysis of Ecological Communities. MjM Software Design, USA.
- McKee, J. K., Sciulli, P. W. and Foose, C. D. (2004). Forecasting global biodiversity threats associated with human population growth. *Biological Conservation* **15**: 161-164.
- Meaza Gidey, Tadesse Beyene, Signorini, M. A., Bruschi, P. and Gidey Yirga (2015). Traditional medicinal plants used by Kunama ethnic group in Northern Ethiopia. *Journal of Medicinal Plants Research* **9(15)**: 494-509.
- Ministry of Environment, Forestry and Climate Change (2016). Ethiopia's Forest Reference Level Submission to the UNFCCC. Addis Ababa, Federal Democratic Republic of Ethiopia.
- Melaku Bekele (2003). Forest property rights, the role of the State and institutional exigency: The Ethiopian experience. *Currents* **31/32**: 14-17.
- Melese Damite (2001). Land use and forest legislation for conservation, development and utilization of forests. **In: Imperative Problems Associated with Forestry in Ethiopia** pp. 3144 (Biological society of Ethiopia eds). Proceedings of a workshop. February 1, 2001, Addis Ababa University.
- Mesfin Tadesse (2004). *Asteraceae (Compositae)* In: Hedberg I, Friis I, Edwards S, eds. *Flora of Ethiopia and Eritrea*. (vol. 4:2).. Department of Systematic Botany, Uppsala, Sweden & The National Herbarium, Addis Ababa, Ethiopia. pp. 408.
- Mesfin Woldearegay (2017). Floristic Composition, Structural Analysis and Land Use/Land Cover Change in Bore-Anferara-Wadera Forest, Southern Ethiopia. PhD Dissertation. Addis Ababa University, Addis Ababa, Ethiopia.
- Ministry of Agriculture (2003). A strategic plan for the sustainable development, conservation and management of the woody biomass resources.

- Ministry of Agriculture (2015). Agricultural Growth Program II. Environmental and Social Management Framework, Addis Ababa, Federal Democratic Republic Of Ethiopia.
- Ministry of Environment and Forestry (2015). Pilot Redd+ Sites Visit Report. Federal Democratic Republic of Ethiopia, Addis Ababa.
- Mirutse Giday (2001). An ethnobotanical study of medicinal plants used by the Zay people in Ethiopia. *CBM:s Skriftserie* **3**: 81–99.
- Mirutse Giday and Gobena Ameni (2003). An Ethnobotanical Survey on Plants of Veterinary Importance in two Districts of Southern Tigray, Northern Ethiopia. *SINET: Ethiop. J. Sci.* **26(2)**: 123–136.
- Mirutse Giday, Zemedede Asfaw, Zerihun Woldu and Tilahun Teklehaymanot (2009). Medicinal plant knowledge of the Bench Ethnic group of Ethiopia: an ethnobotanical investigation. *Journal of Ethnobiology and Ethnomedicine* **5**: 34.
- Mirutse Giday, Zemedede Asfaw and Zerihun Woldu (2010). Ethnomedicinal study of plants used in Sheko ethnic group of Ethiopia. *Journal of Ethnopharmacology* **132**: 75-85.
- Mitiku Haile and Kindeya Gebrehiwot (2000). Efforts to rehabilitate degraded lands: local initiative for planning resource management in Tigray. In: Feoli, E., Pottier, D. and Zerihun Woldu (eds.). *Proceeding of the international workshop on sustainable development of Dryland areas of East Africa*, November 9th–12th 1988, University of Trieste, Italy. pp. 319-330.
- Mittermeier, R. A, Myers, N. and Thomsen, J. B. (1998). Biodiversity hotspots and major tropical wilderness areas: Approaches to setting conservation priorities. *Conservation Biology* **12(3)**: 516-520.
- Moravec, I., Fernandez, E., Vlkova, M. and Milella, L. (2014). Ethnobotany of Medicinal Plants of Northern Ethiopia. *Boletín Latinoamericano Y Del Caribe De Plantas Medicinales Y Aromáticas* **13:2**: 126-134.
- Mueller-Dombois, D. and Ellenberg, H. (1974). Aims and methods of vegetation ecology. John Wiley, New York.
- Mulugeta Lemenih and Demel Teketay (2006). Changes in soil seed bank composition and Density following deforestation and subsequent cultivation of a tropical dry Afromountane forest in Ethiopia. *Tropical Ecology* **47**: 1-12.
- Murphy, P. G. and Lugo, A. E. (1986) Ecology of tropical dry forest. *Annu. Rev. Ecol. Syst.* **17**: 67-88.

- Mushongi, A. A. (2001). Traditional plant concept and plant use by the Pare people of Northern Tanzania: An Ethnobotanical study in semi-arid land vegetation. MSc. Thesis in Dry land Biodiversity, Addis Ababa University, Ethiopia.
- Mutke, J. and Barthlott, W. (2005). Patterns of vascular plant diversity at continental to global scales. *Biol. Skr.* **55**: 521-531.
- Muys, B., Kindeya Gebrehiwot and Bruneel, S. (Eds.). (2004). Symposium on the rehabilitation of dry land forests in Ethiopia: ecology and management. Book of abstracts. Mekelle, Ethiopia, 21-24 September 2004.
- Myers, N., Mittermeier R. A. and Mittermeier, C. G. (2000). Biodiversity hotspots for conservation priorities. *Nature* **403**: 853-858.
- National Metrological Service Agency (2017). Data of Rainfall and Temperature of 20 years (1998-2017). Addis Ababa, Ethiopia.
- Noss, R.F. (1999) Assessing and monitoring forest biodiversity: A suggested framework and indicators. *Forest Ecology and Management* **115**: 135-146.
- Nurya Abdurhman (2010). Ethnobotanical Study of Medicinal Plants Used by Local People in Ofla District, Southern Zone of Tigray Region, Ethiopia. MSc. Thesis. Addis Ababa University, Ethiopia.
- Nyssen, J. (2001). Erosion processes and soil conservation in a tropical mountain catchment under threat of anthropogenic desertification – a case study from Northern Ethiopia. PhD. Dissertation. KULeuven, Leuven.
- Nyssen, J., Poesen J., Moeyersons J., Deckers J., Mitiku H. and Lang, A. (2004). Human impact on the environment in the Ethiopian and Eritrean highlands – a state of the art. *Earth-Science Reviews* **64**: 157-323.
- Økland, R. H. (1990). Vegetation Ecology: theory, methods and application with reference to Fennoscandia. Norway: *Sommerfeltia* Suppl. pp. 1-233.
- Orme, C. D., Davies, R. G. and Burgess, M. (2005). Global hotspots of species richness are not congruent with endemism or threat. *Nature* **436**: 1010-1019.
- Palmer, M.W. (1993) Putting Things in Even Better Order: The Advantages of Canonical Correspondence Analysis *Ecological Society of America* **74:8**: 2215-2230
- Peet, R. K. (1974). The measurement of species diversity. *Ann. Rev.Ecol.Syst.* **5**: 285-307.

- Perera, G. A. D. (2005). Spatial heterogeneity of the soil seed bank in the tropical semideciduous forest at Wasgomuwa National Park, Sri Lanka. *Trop. Ecol.* **46**: 79–89.
- Peters, C. M. (1996). Beyond Nomenclature and Use: A review of Ecological Methods for Ethnobotanists. New York Botanical Garden, New York. pp. 241-278.
- Phillips, O. and Gentry, H.A. (1993) The useful plants of Tambopata, Peru. I: statistical hypotheses tests with a new quantitative technique. *Economic Botany* **47**: 15-32.
- Phillips, O. L., Gentry, A. H., Reynel, C., Wilkin, P. and Galvez-Durand, B. C. (1994). Quantitative ethnobotany and Amazonian conservation. *Conservation Biology* **8**: 225-248.
- Phillips, S (1995). *Poaceae (Gramineae) (vol. 7)*. In: Hedberg I. & Edwards S., Eds. *Flora of Ethiopia and Eritrea*. Department of Systematic Botany, Uppsala, Sweden: The National Herbarium, Addis Ababa, Ethiopia. pp. 438.
- Pianka, E. R. (1966). Latitudinal gradients in species diversity: a review of concepts. *The American Naturalist* **100(910)**: 33-46.
- Pichi-Sermolli, R. E. (1957). Una carta geobotanica dell'Africa Orientale (Eritrea, Ethiopia, Somalia). *Webbia* **13**: 15-132.
- Pielou, E.C. (1977). *Mathematical ecology*. John Wiley and Sons, New York.
- Pimm, S. L. and Raven, P. (2000). Extinction by numbers. *Nature* **403**: 843-845.
- Plotkin, M. J. (1995). The Importance of Ethnobotany for Tropical Forest Conservation. *Ethnobotany: Evolution of a Discipline*. Oracle, AZ, Dioscorides Press.
- Pokhriyal, P., Uniyal, P., Chanuahan, D. S. and Todaria, N. P. (2010). Regeneration status of tree species in forest of phakot and pathri Rao watersheds in Garhwal Himalaya. *Current Science* **98(2)**: 171-175.
- Poorter, L., Bongers, F., van Rompaey, R.S. and de Klerk, M. (1996). Regeneration of canopy tree species at five sites in West African moist forest. *Forest Ecology and Management* **85**: 61-69.
- Popma, J., Bongers, F. and Meave del Castillo, J. (1988). Patterns in the vertical structure of the lowland rainforest of Los Tuxtlas, Mexico. *Vegetatio* **74**: 81 -91.
- Poorter, J., Kingsford, R. and Brock, M. (2007). Seed banks in arid wetlands with contrasting flooding, salinity and turbidity regimes. *Plant Ecology* **188**: 215–234.
- Prance, G. T., Balee, W., Boom, B. M. and Carnerio, R. L. (1987). Quantitative ethnobotany and the case for conservation in Amazonia. *Conserv. Biol.* **1**: 296-310.

- Price, J. N., Wright, B. D., Gross, C. L. and Whalley, W. R. D. B. (2010). Comparison of seedling emergence and seed extraction for estimating the composition of soil seed banks. *Methods Ecol. Evol.* **2**: 151-157.
- Rajan, S.S. (2000) Plant physiology. ANMOL publication pvt.Ltd. New Delhi, pp. 521.
- Ramakrishnan, P. S. (1999). The Impact of globalization on agriculture systems of traditional societies. In: A.K. Dragun and C. Tisdell (Eds.). *Sustainable Agriculture and Environment: Globalization and the Impact of Trade Liberalization*,. Edward Elgar, Chettenham, UK. pp. 185-200.
- Rastogi, A., Godbole, A. and Shengji, P. (1998). Applied Ethnobotany in natural Resources Management –Traditional Home Gardens: Highlights of a Training Workshop Held at Kohima, Nagaland, India from 18-23 June 1997. International Centre for Integrated development, Kathmandu, Nepal.
- R Development Core Team (2017). R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Vienna, Austria or URL: <http://www.R-project.org>.
- Reubens, B., Heyn, M., Kindeya Gebrehiwot, Hermy, M. and Muys, B. (2007). Persistent soilseed banks for natural rehabilitation of dry tropical forests in northern Ethiopia. *Tropicultura* **25**: 204-214.
- Reusing, M. and Kasberger, G. (2000). Wald-monitoring in Äthiopien. *Allgemeine Forstzeitschrift/ Der Wald* **4**: 207-208.
- Rey, P. J., Julio, M. and Alcantara, M. (2000). Recruitment dynamics of a fleshy-fruited plant (*Olea europaea*): Connecting patterns of seed dispersal to seedling establishment. *Journal of Ecology* **88(4)**: 622-633.
- Reyes-García, V., Marti, N., McDade, T.W., Tanner, S. and Vadez, V. (2007). Concepts and methods in studies measuring individual ethnobotanical knowledge. *Journal of Ethnobiology* **27**: 108-203.
- Robertson, H. A. and James, K. R. (2006). Plant establishment from the seed bank of a degraded floodplain wetland: a comparison of two alternative management scenarios. *Plant Ecology* **188**: 145–164.
- Rosenzweig, M. L. (1995). Species diversity in space and time. Cambridge University Press, Cambridge.

- Sanjit, L. and Bhatt, D. (2005). How relevant are the concepts of species diversity and species richness? *Journal of Biosciences* **30(5)**: 557-560.
- Schefus, E., Schouten, S. and Jansen, J. (2003). African vegetation controlled by tropical sea surface temperatures in the mid-Pleistocene period. *Nature* **422**: 418–421.
- Schmitt, C. B., Feyera Senbeta, Tadesse Woldemariam, Rudner, M. and Denich, M. (2013). Importance of regional climates for plant species distribution patterns in moist Afromontane forest. *Journal of Vegetation Science* **24**: 553–568.
- Sebsebe Demissew, Mengistu Wondafrash and Yilma Dellelegn (1996). Forest resources of Ethiopia. In S. Tilahun *et al* (eds), Important Bird Areas of Ethiopia. A First Inventory Ethiopian Wildlife and Natural History Society, Addis Ababa, pp. 10-25.
- Silva, V. A., Andrade, L. and de Albuquerque, U. P. (2006). Revising the Cultural Significance Index: The case of the Fulni-ô in Northeastern Brazil. *Field Methods* **18**: 98-108.
- Silva, F.D.S., Ramos, M.A., Hanazaki, N. and Albuquerque, U.P. (2011). Dynamics of traditional knowledge of medicinal plants in a rural community in the Brazilian semi-arid region. *Brazilian Journal of Pharmacognosy* **21**: 382-391.
- Simon Shibru and Girma Balcha (2004). Composition, structure and regeneration status of woody species in Dindin Natural Forest, Southeast Ethiopia: An implication for conservation. *Ethiopian Journal of Biological Science* **3(1)**: 15–35.
- Simpson, E. H. (1949). Measurement of diversity. *Nature* **163**: 688:688.
- Simpson, R. L., Leck, M. A. and Parker, V. T. (1989). Seed banks: general concepts and Methodological issues. Pp. 3-8. In: M.A. Leck, V. T. Parker and R. L. Simpson (eds). *Ecology of Soil Seed Banks*. Academic Press, San Diego.
- Smith, R. S., Shiel, R. S., Millward, D., Corkhill, P. and Sanderson, R. A. (2002). Soil seed banks and the effects of meadow management on vegetation change in a 10-year meadow field trial. *J. Appl. Ecol.* **39**: 279-293.
- Sørensen, T. (1948). A method of establishing groups of equal amplitude in plant sociology based on similarity of species and its application to analyses of the vegetation on Danish commons. *Kongelige Danske Videnskabernes Selskab* **5 (4)**: 1–34
- Stoffle, R.W., Halmo, D. B., Evans, M. J. and Olmsted, J. E. (1990). Calculating the cultural significance of American Indian plants: Paiute and Shoshone ethnobotany at Yucca Mountain, Nevada. *American Anthropologist* **92**: 416-432.

- Tadesse Beyene (2015). Ethnobotany of medicinal plants in Erob and Gulomahda districts, Eastern Zone of Tigray Region, Ethiopia. PhD Dissertation. Addis Ababa University, Ethiopia.
- Tadesse Woldemariam (2003). Vegetation of the Yayu Forest in Southwest Ethiopia: Impact of Human Use and Implementations for In-situ Conservation of Wild *Coffea Arabica* L. Populations. Doctoral Dissertation, Center for Development and Research, University of Bonn, Bonn.
- Tadesse Woldemariam, Demel Teketay, Edwards, S. and Olsson, M. (2000). Woody plant and avian species diversity in a dry Afromontane forest on the central plateau of Ethiopia: Biological indicators for conservation. *Ethiopian Journal of Natural Resources* **2**: 255-293.
- Teferi Gedif and Hahn, H. (2002). Herbalists in Addis Ababa and Butajira, Central Ethiopia: Mode of service delivery and traditional pharmaceutical practice. *Ethiopian Journal of Health Development* **16**: 191-197.
- Tefera Mengistu, Demel Teketay, Hakan, H. and Yonas Yemshaw (2005). The role of enclosures in the recovery of woody vegetation in degraded dryland hillsides of central and northern Ethiopia. *J. Arid Environ.* **60**: 259–281.
- Tamrat Bekele (1993). Vegetation ecology of remnant Afromontane forests on the central plateau of Shewa, Ethiopia. PhD Dissertation. Uppsala: Opulus press. pp. 64.
- Tamrat Bekele (1994). Phytosociology and Ecology of Humid Afromontane Forest on the Central plateau of Ethiopia. *Journal of Vegetation Science* **5**: 87–98.
- Tengnas, B. (1994). Agroforestry extension manual for Kenya. Nairobi.
- Terborgh, J. and van Schaik, C. P. (1997). Minimizing species loss: The imperative of protection. In R. Kramer et al (eds) Protected Areas and the Defense of Tropical Biodiversity. Oxford University Press, New York, pp. 15-35.
- Ter Braak, C. J. F. (1987). The analysis of vegetation-environment relationships by canonical correspondence analysis. *Vegetation* **69**: 69-77.
- Ter Braak, C. J. F. and Smilauer, P. (1998). CANOCO Release 4. Reference Manual and Users Guide to CANOCO for Windows: Software for Canonical Community Ordination. Microcomputer Power, Ithaca, USA.
- Ter Heerdt, G. N. J., Verweij, G. L., Bekker, R. M. and Bakker, J. P. (1996). An improved method for seed-bank analysis: seedling emergence after removing the soil by sieving. *Functional Ecology* **10**: 144–151.

- Ter Heerdt, G. N. J., Schutter, A. and Bakker, J. P. (1999). The effect of water supply on seed-bank analysis using the seedling-emergence method. *Functional Ecology* **13**: 428–430.
- Ter Steege, H. and Zagt, R. (2002). Density and diversity. *Nature* **417(12)**: 698-699.
- Teshale Sori, Merga Bekana, Girma Adugna and Ensermu Kelbessa (2004). Ethnoveterinary practice of Borena pastoralists, southern Ethiopia. *Intern. J. Appl. Res. Vet. Med.* **2(3)**: 220-225.
- Tewelde Berhan Gebre Egziabher (1988). Vegetation and environment of the mountains of Ethiopia: Implications for utilization and conservation. *Mountain Research and Development* **8**: 211-216.
- Tewelde Berhan Gebre Egziabher (1991). Diversity of the Ethiopian Flora. **In**: Engels, J.M.M., Hawkes, J.G. and Worede, M. (eds.). *Plant genetic resources of Ethiopia*, Cambridge University Press. pp. 75-81.
- Tigray Forestry Action Program (1996). Bureau of Agricultural and Natural Resources Development, main report, vol. I. Mekelle, Tigray, Ethiopia.
- Thompson, K. and Grime, J. P. (1979). Seasonal variation in the seed banks of herbaceous species in ten contrasting habitats. *The Journal of Ecology* **67**: 893–921.
- Thompson, K. (1987). Seeds and seed banks. *New Phytol.* **106 (Suppl.)**: 23–34.
- Thrupp, L. A. (1997). Linking Biodiversity and Agriculture. Challenges and Opportunities for Sustainable Food Security. World resource institute. Washington, D.C.
- Tilahun Teklehaymanot (2009). Ethnobotanical study of knowledge and medicinal plants use by people in Dek Island in Ethiopia. *Journal of Ethnopharmacology* **124**: 69-78.
- Tilahun Teklehaymanot and Mirutse Giday (2007). Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. *Journal of Ethnobiology and Ethnomedicine* **3**: 12.
- Tinsae Bahru, Zemedet Asfaw and Sebsebe Demissew (2012). Indigenous knowledge on fuel wood (charcoal and/or firewood) plant species used by the local people in and around the semi-arid Awash National Park, Ethiopia. *Journal of Ecology and the Natural Environment* **4(5)**: 141-149.
- Tokeshi, M. (1999). Species coexistence: Ecological and evolutionary perspectives. Blackwell Science Ltd, Malden.
- Toledo, L. L and Ramos, M. M. (2011). The soil seed bank in abandoned tropical pastures: source of regeneration or invasion. *Biodiversity* **82**: 663-678.

- Trotter, R. T. and Logan, M. H. (1986). Informants consensus: a new approach for identifying potentially effective medicinal plants In: N.L. Etkin (ed). *Plants in Indigenous Medicine and Diet*, , Redgrave Publishing Company, Bedford Hill, NY, pp. 91-112.
- Turner, N. J. (1988). "The importance of a rose": Evaluating the cultural significance of plants in Thompson and Lillooet Interior Salish. *American Anthropologist* **90**: 272-290.
- Uasuf, A., Tigabu, M. and Oden, P. C. (2009). Soil seed banks and regeneration of neotropical dry deciduous and gallery forests in Nicaragua, Paris. *Bois et Forêts des Tropiques* **299**: 49–62.
- Uniyal, S. K., Singh, K. N., Jamwal, P. and Lal, B. (2006). Traditional use of medicinal plants among the tribal communities Chhota Bhangal, Western Himalaya. *Journal of Ethnobiology and Ethnomedicine* **2**:14.
- van der Maarel, E. (1979). Transformation of cover-abundance values in Phytosociology and its effects on community similarity. *Vegetatio* **39 (2)**: 97-114.
- van Tongeren, O. F. R. (1995). Cluster analysis. In: Jongman, R.H.G., Ter Braak, J.C.F. and van Tongeren, O.F.R., eds. *Data Analysis in Community and Landscape Ecology*, pp. 174-212,.. Cambridge University Press, Cambridge.
- Van der Valk, A. G. and Pederson, R. L. (1989). Seed banks and the management and restoration of natural vegetation. pp. 329-346. In: M.A. Leck, V.T. Parker and R.L. Simpson (eds.). *Ecology of Soil Seed Banks*. Academic Press, San Diego.
- Vivero, J. L., Ensermu Kelbessa and Sebsebe Demissew (2005). The Red List of Endemic Trees and Shrubs of Ethiopia and Eritrea. Fauna and Flora International, United Kingdom. pp. 28.
- Voeks, R. A. (2007). Are women reservoirs of traditional plant knowledge? Gender, ethnobotany and globalization in northeast Brazil. *Singapore Journal of Tropical Geography* **28**: 7-20.
- Voeks, R. A. and Leony, A. (2004). Forgetting the forest: assessing medicinal plant erosion in eastern Brazil. *Economic Botany* **58**: 294-306.
- Walter, K.S. and Gillett, H.J. (eds) (1998). 1997 IUCN Red List of Threatened Plants. WCMC, Cambridge, UK & IUCN, Gland, Switzerland.
- Wang, J., Huang, L., Ren, H., Sun, Z. and Guo, Q. (2015). Regenerative potential and functional composition of soil seed banks in remnant evergreen broad-leaved forests under urbanization in South China. *Community Ecology* **16(1)**: 86-94.
- World Conservation Monitoring Centre (1994). Biodiversity Data Sourcebook. World Conservation Press, Cambridge, UK.

- White, F. (1978). The Afromontane Region. In: Werger, M.J.A. (eds.). *Biogeography and ecology of Africa*, Junk: The Hague, pp. 463-513.
- White, F. (1983). The Vegetation of Africa. A descriptive memoir to accompany the UNESCO/AETFAT/ UNIDO vegetation map of Africa. *Natural Resources Research* **20**: 1-356.
- Whitemore, T. C. (1997). Tropical forest disturbance, disappearance and species loss. In Laurance, W.F. and Bierregaard RO (eds.) *Tropical forest remnants: Ecology, management and conservation of fragmented communities*. The University of Chicago Press, Chicago. pp. 616.
- Whittaker, R. H. (1972). Evolution and measurement of species diversity. *Taxon* **21**: 213-251.
- Whittaker, R.J., Willis, K.J. and Field, R. (2003). Climatic–energetic explanations of diversity: a macroscopic perspective. In *Macroecology: concepts and consequences*, Blackburn, T.M. and Gaston, K.J. (eds), Cambridge University Press; Cambridge. pp. 107-129.
- Wilson, R. T. (1977) The vegetation of central Tigre, Ethiopia in relation to its land use. *Webbia* **32**: 235-270.
- Winter, K. and McClatchey, W. (2008). Quantifying evolution of cultural interactions with plants: implications for managing diversity for resilience in social-ecological systems. *Functional Ecosystems and Communities* **2** (1): 1-10.
- Wood, A., Stedman-Edwards, P. and Mang, J., eds. (2000) *The root causes of biodiversity loss*. Earthscan Publishings Ltd, London.
- World Conservation Monitoring Centre (1998) *Juniperus procera*. 2013 IUCN Red List of threatened species. www.iucnredlist.org. Cited 24 July 2018.
- Wright, B. R. and Clarke, P. J. (2009). Fire, aridity and seed banks. What does seed bank composition reveal about community processes in fire-prone desert? *Journal of Vegetation Science* **20**: 663–674.
- Zemedu Asfaw (1997). The Role of Ethnobotany in Biodiversity Conservation. **In**: *Community Biodiversity Development and Conservation* (CBDC). Ethiopian Project Working paper-1, Biodiversity Institute of Ethiopia. pp. 1-17.
- Zemedu Asfaw (2006). Towards a dynamic indigenous knowledge practice: Optimization of the uses and management of plant resources in the Drylands of Ethiopia. **In**: Nikundiwe, M, A. and Kabigumila, D.L.J (eds). *Dryland ecosystem: challenges and opportunities for sustainable Natural Resources Management*. (). Proceedings of the Regional Workshop held at Arusha, Tanzania. pp. 64-67.
- Zemedu Asfaw and Mesfin Tadesse (2001). Prospects for Sustainable Use and Management of Wild Food Plants in Ethiopia. *Econ. Bot.* **55**(1): 47-62.

- Zent, S. (1996). Behavioral Orientations toward Ethnobotanical Quantification pp. 199-240, **In:** *Selected Guidelines for Ethnobotanical Research: A Field Manual* (Alexiades, N. ed). Scientific Publications Department. The New York Botanical Garden Bronx.
- Zerihun Woldu (1999). Forests in the vegetation types of Ethiopia and their status in the geographical context. **In:** Edwards, S., Abebe Demissie, Taye Bekele and Haase, G. (eds). *proceedings of the national forest genetic resources conservation strategy development workshop*. June 21-22, 1999, Addis Ababa, Ethiopia. pp. 1- 39.
- Zerihun Woldu (2017). Comprehensive Analysis of Vegetation and Ecological Data: concepts and methods. Addis Ababa: Addis Ababa University Press. pp. 482.
- Zerihun Woldu and Backeus, I. f. (1991). The shrubland vegetation in western Shewa, Ethiopia and its possible recovery. *Journal of Vegetation Science* **2**: 173-180.
- Zerihun Woldu, Dragan, M., Feoli, E. and Ferneti, M. (2002). Reducing soil erosion in Northern Ethiopia, Adwa Zone, through a special decision support system (SDSS). *Ethiopian J. Biol. Sci.* **1(1)**: 1-12.

Appendices

Appendix 1. Plant species found in *Wejig-Mahgo-Waren* massif forest (Habit: T = Tree; S = Shrub; H = Herb; WC = Woody climber; HC = Herbaceous climber; Coll.No. = Collection number; MH = Mebrahtu Hishe)

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|--------------------|-----------------|-------|-----------|
| <i>Abutilon longicuspe</i> Hochst. ex A. Rich. | Malvaceae | Sa'da buwak | S | MH 109 |
| <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Chea | T | MH 154 |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | T | MH 120 |
| <i>Acacia polyacantha</i> Willd. | Fabaceae | | T | MH 123 |
| <i>Acacia senegal</i> (L.) Willd. | Fabaceae | Sewansa | T | MH 141 |
| <i>Acacia seyal</i> Del. | Fabaceae | Sa'da chea | T | MH 143 |
| <i>Acacia sieberiana</i> DC. | Fabaceae | | T | MH 155 |
| <i>Acacia tortilis</i> (Forssk.) Hayne | Fabaceae | | T | MH 124 |
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Machelo | H | MH 057 |
| <i>Acokanthera schimperi</i> (A. DC.) Schweinf. | Apocynaceae | Mrenz | T | MH 067 |
| <i>Actiniopteris radiata</i> (Sw.) Link | Actiniopteridaceae | | H | MH 235 |
| <i>Adiantum lunulatum</i> Burm.f. | Adiantaceae | | H | MH 236 |
| <i>Aerva javanica</i> (Burm.f.) Schultes | Amaranthaceae | | S | MH 131 |
| <i>Aerva lanata</i> (L.) Juss. ex Schultes | Amaranthaceae | | H | MH 184 |
| <i>Agrocharis melanantha</i> Hochst. | Apiaceae | | H | MH 277 |
| <i>Allophylus abyssinicus</i> (Hochst.) Radik | Sapindaceae | Tuemay/maara | T | MH 106 |
| <i>Aloe camperi</i> Schweinf. | Aloaceae | Ere | S | MH 302 |
| <i>Anagallis arvensis</i> L. | Primulaceae | | H | MH 182 |
| <i>Andropogon chrysostachyus</i> Steud. | Poaceae | | H | MH 226 |
| <i>Andropogon distachyos</i> L. | Poaceae | | H | MH 227 |
| <i>Andropogon schirensis</i> Hochst. ex A. Rich. | Poaceae | Qeran saeri | H | MH 205 |
| <i>Anthospermum herbaceum</i> L.f. | Rubiaceae | | H | MH 115 |
| <i>Argemone mexicana</i> L. | Papaveraceae | Medafe | S | MH 077 |
| <i>Artemisia abyssinica</i> Sch. Bip. ex A. Rich. | Asteraceae | Chena baria | H | MH 128 |
| <i>Asparagus africanus</i> Lam. | Asparagaceae | Kestenosto | WC | MH 044 |
| <i>Asparagus racemosus</i> Willd. | Asparagaceae | Kestenosto | WC | MH 045 |
| <i>Asplenium aethiopicum</i> (Burm.f.) Bech. | Aspleniaceae | | H | MH 234 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|-----------------|-----------------|-------|-----------|
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | T | MH 019 |
| <i>Barleria eranthemoides</i> R. Br. | Acanthaceae | Melhas sebeyti | S | MH 168 |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | Tebeb | S | MH 013 |
| <i>Berberis holstii</i> Engl. | Berberidaceae | Mucha euf | S | MH 055 |
| <i>Bersama abyssinica</i> Fresen. | Melianthaceae | Mirkuz zibe | T | MH 050 |
| <i>Bidens carinata</i> Cufod. ex Mesfin | Asteraceae | Adey ababa | H | MH 200 |
| <i>Bidens pilosa</i> L. | Asteraceae | Tselim Teneg | H | MH 011 |
| <i>Blepharis integrifolia</i> (L.f.) E.Mey. ex Schinz | Acanthaceae | | H | MH 291 |
| <i>Boscia salicifolia</i> Oliv. | Capparidaceae | Tetem agazen | T | MH 138 |
| <i>Brachiaria brizantha</i> (A. Rich.) Stapf | Poaceae | | H | MH 206 |
| <i>Brachiaria leersioides</i> (Hochst.) Stapf | Poaceae | | H | MH 214 |
| <i>Brassica rapa</i> L. | Brassicaceae | | H | MH 180 |
| <i>Buddleja polystachya</i> Fresen. | Loganiaceae | Metere | T | MH 137 |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | T | MH 002 |
| <i>Calotropis procera</i> (Ait.) Ait.f. | Asclepiadaceae | Gindaea | S | MH 030 |
| <i>Calpurnia aurea</i> (Ait.) Benth. | Fabaceae | Chrenchah | T | MH 001 |
| <i>Capparis tomentosa</i> Lam. | Capparidaceae | Harengama | WC | MH 075 |
| <i>Carissa spinarum</i> L. | Apocynaceae | Egam | S | MH 062 |
| <i>Cassipourea malosana</i> (Baker) Alston | Rhizophoraceae | Tselim om | T | MH 073 |
| <i>Caylusea abyssinica</i> (Fresen.) Fisch. & Mey. | Resedaceae | Hmak chenawi | H | MH 181 |
| <i>Celtis africana</i> Burm. f. | Ulmaceae | Beto Koma | T | MH 094 |
| <i>Cenchrus ciliaris</i> L. | Poaceae | | H | MH 228 |
| <i>Cheilanthes farinosa</i> (Forssk.) Kaulf | Sinopteridaceae | | H | MH 233 |
| <i>Chloris gayana</i> Kunth | Poaceae | | H | MH 223 |
| <i>Cirsium vulgare</i> (Savi.) Ten. | Asteraceae | | H | MH 192 |
| <i>Cissampelos mucronata</i> A. Rich. | Menispermaceae | Hareg mintaro | HC | MH 269 |
| <i>Cissus quadrangularis</i> L. | Vitaceae | Anji afar | HC | MH 032 |
| <i>Clematis simensis</i> Fresen. | Ranunculaceae | Hareghazo | HC | MH 025 |
| <i>Clerodendrum myricoides</i> (Hochst.) Vatke | Lamiaceae | Shewha | S | MH 021 |
| <i>Clutia lanceolata</i> Forssk. | Euphorbiaceae | Hirtmtmo | S | MH 023 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|--|------------------|------------------------|--------------|------------------|
| <i>Colutea abyssinica</i> Kunth & Bouche | Fabaceae | Tyt hibey/tatata | S | MH 008 |
| <i>Combretum molle</i> R. Br. ex G. Don | Combretaceae | Tataso | T | MH 286 |
| <i>Commelina africana</i> L. | Commelinaceae | Gamale | H | MH 158 |
| <i>Commelina benghalensis</i> L. | Commelinaceae | Gamale | H | MH 159 |
| <i>Commicarpus pedunculatus</i> (A.Rich.) Cufod. | Nyctaginaceae | Ezni anchwa | H | MH 022 |
| <i>Conyza hypoleuca</i> A..Rich | Asteraceae | | H | MH 203 |
| <i>Conyza vernonioides</i> (Sch. Rip. ex A. Rich.) Wild | Asteraceae | | S | MH 197 |
| <i>Cordia monoica</i> Roxb. | Boraginaceae | | T | MH 100 |
| <i>Craterostigma longicarpum</i> Hepper | Scrophulariaceae | | H | MH 167 |
| <i>Crotalaria incana</i> L. | Fabaceae | Shamto amrakut | H | MH 280 |
| <i>Cupressus lusitanica</i> Mill. | Cupressaceae | Tshdi ferenji | T | MH 037 |
| <i>Cyanotis barbata</i> D.Don | Commelinaceae | | H | MH 157 |
| <i>Cyathula orthacantha</i> (Aschers.) Schinz | Amaranthaceae | | H | MH 122 |
| <i>Cynanchum abyssinicum</i> Decne. | Asclepiadaceae | Elbi | HC | MH 175 |
| <i>Cynanchum gerrardii</i> (Harv.) Liede | Asclepiadaceae | Armed | HC | MH 121 |
| <i>Cynodon dactylon</i> (L.) Pers. | Poaceae | Tehag | H | MH 218 |
| <i>Cynoglossum lanceolatum</i> Forssk. | Boraginaceae | | H | MH 144 |
| <i>Cyperus dubius</i> Rottb. | Cyperaceae | Kunti | H | MH 268 |
| <i>Cyperus impubes</i> Steud. | Cyperaceae | Saeri htshs | H | MH 267 |
| <i>Cyphostemma adenocaula</i> (Steud. ex A. Rich.) Desc. ex Wild and Drummond | Vitaceae | Aserkuka | HC | MH 017 |
| <i>Dactyloctenium aegyptium</i> (L.) Willd. | Poaceae | Tehag | H | MH 217 |
| <i>Datura stramonium</i> L. | Solanaceae | Mestenagr | H | MH 169 |
| <i>Debregeasia saeneb</i> (Forssk.) Hepper and Wood | Urticaceae | Tseada qotsli | S | MH 069 |
| <i>Dichrostachys cinerea</i> (L.) Wight and Arn. | Fabaceae | Karshamarsha | S | MH 088 |
| <i>Dicrocephala chrysanthemifolia</i> (Bl.) DC. | Asteraceae | | H | MH 296 |
| <i>Discopodium penninervium</i> Hochst. | Solanaceae | Alhim | S | MH 027 |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | MH 059 |
| <i>Dombeya torrida</i> (J.F.Gmel.) P. Bamps | Sterculiaceae | Buyak | T | MH 068 |
| <i>Dovyalis abyssinica</i> (A. Rich) Warb. | Flacourtiaceae | Mongolhats | T | MH 112 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|--|-----------------|-----------------|-------|-----------|
| <i>Dovyalis verrucosa</i> (Hochst.) Warb. | Flacourtiaceae | Teumtegna | T | MH 038 |
| <i>Dregea abyssinica</i> (Hochst.) K. Schum. | Asclepiadaceae | Hareg | WC | MH 166 |
| <i>Dryopteris schimperiana</i> (Hochst. ex A.Br.) C.Chr. | Dryopteridaceae | | H | MH 231 |
| <i>Echinops hispidus</i> Fresen. | Asteraceae | Dander | H | MH 191 |
| <i>Echinops pappii</i> Chiov. | Asteraceae | Dander | S | MH 193 |
| <i>Ehretia cymosa</i> Thonn. | Boraginaceae | Aulaga | T | MH 095 |
| <i>Ekebergia capensis</i> Sparrm. | Meliaceae | Kot | T | MH 042 |
| <i>Eleusine floccifolia</i> (Forssk.) Spreng. | Poaceae | | H | MH 219 |
| <i>Epilobium hirsutum</i> L. | Onagraceae | | H | MH 204 |
| <i>Equisetum ramosissimum</i> Desf. | Equisetaceae | | H | MH 136 |
| <i>Eragrostis papposa</i> (Roem. & Schult.) Steud. | Poaceae | | H | MH 208 |
| <i>Eragrostis pilosa</i> (L.) P. Beauv. | Poaceae | | H | MH 213 |
| <i>Erica arborea</i> L. | Ericaceae | Hasti | T | MH 056 |
| <i>Erythrococca abyssinica</i> Pax | Euphorbiaceae | | S | MH 325 |
| <i>Eucalyptus globulus</i> Labill. | Myrtaceae | Tseada bahirzaf | T | MH 049 |
| <i>Euclea racemosa</i> subsp. <i>schimperii</i> (A. DC.) White | Ebenaceae | Kuleo | T | MH 102 |
| <i>Euphorbia abyssinica</i> Gmel. | Euphorbiaceae | Qulqal | T | MH 300 |
| <i>Euphorbia petitiiana</i> A. Rich. | Euphorbiaceae | Hinzuquzuq | H | MH 176 |
| <i>Ferula communis</i> L. | Apiaceae | Doeg | H | MH 029 |
| <i>Ficus glumosa</i> Del. | Moraceae | Cheqante | T | MH 018 |
| <i>Ficus sur</i> Forssk. | Moraceae | Shamfa | T | MH 085 |
| <i>Ficus sycomorus</i> L. | Moraceae | Sagla | T | MH 104 |
| <i>Galium simense</i> Fresen. | Rubiaceae | | HC | MH 127 |
| <i>Geranium arabicum</i> Forssk. | Geraniaceae | | H | MH 108 |
| <i>Geranium dissectum</i> L. | Geraniaceae | | H | MH 281 |
| <i>Gerbera piloselloides</i> (L.) Cass. | Asteraceae | | H | MH 299 |
| <i>Glycine wightii</i> (Wight and Arn.) Verdc. | Fabaceae | | HC | MH 322 |
| <i>Gomphocarpus purpurascens</i> A. Rich. | Asclepiadaceae | Tseba dimu | S | MH 024 |
| <i>Grewia bicolor</i> Juss. | Tiliaceae | Rewey | T | MH 149 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|------------------|------------------------|--------------|------------------|
| <i>Grewia ferruginea</i> Hochst. ex A. Rich. | Tiliaceae | Rewey | T | MH 148 |
| <i>Grewia mollis</i> A. Juss. | Tiliaceae | Rewey | T | MH 150 |
| <i>Grewia villosa</i> Willd. | Tiliaceae | Rewey | S | MH 147 |
| <i>Hagenia abyssinica</i> (Bruce) J.F. Gmel. | Rosaceae | Habi | T | MH 153 |
| <i>Halleria lucida</i> L. | Scrophulariaceae | | T | MH 282 |
| <i>Haplocarpha schimperi</i> (Sch. Bip.) Beauv. | Asteraceae | | H | MH 129 |
| <i>Harpachne schimperi</i> Hochst. ex A. Rich. | Poaceae | | H | MH 225 |
| <i>Hebenstretia angolensis</i> Rolfe | Scrophulariaceae | | H | MH 276 |
| <i>Helichrysum quartinianum</i> A. Rich. | Asteraceae | | H | MH 195 |
| <i>Helichrysum schimperi</i> (Sch. Bip. ex A. Rich.) Moeser | Asteraceae | | S | MH 202 |
| <i>Helinus mystacinus</i> (Ait.) E. Mey. ex Steud. | Rhamnaceae | Hareg | WC | MH 323 |
| <i>Heliotropium strigosum</i> Willd. | Boraginaceae | Amam gmel | H | MH 066 |
| <i>Helixanthera thomsonii</i> (Sprague) Danser | Loranthaceae | Diqala | S | MH 145 |
| <i>Heteromorpha arborescens</i> (Spreng.) Cham. & Schlecht. | Apiaceae | | S | MH 005 |
| <i>Hibiscus crassinervius</i> Hochst. ex A. Rich. | Malvaceae | | S | MH 174 |
| <i>Hibiscus macranthus</i> Hochst. ex A. Rich. | Malvaceae | Necha | S | MH 151 |
| <i>Huernia macrocarpa</i> (A. Rich.) Sprenger | Asclepiadaceae | Ango | H | MH 033 |
| <i>Hyparrhenia hirta</i> (L.) Stapf | Poaceae | Saeri geza | H | MH 212 |
| <i>Hyparrhenia rufa</i> (Nees) Stapf | Poaceae | Demhala | H | MH 210 |
| <i>Hypericum revolutum</i> Vahl | Hypericaceae | Abedi | T | MH 132 |
| <i>Hypericum roeperianum</i> Schimp. ex A. Rich. | Hypericaceae | | T | MH 183 |
| <i>Hypoestes forskaolii</i> (Vahl) R. Br. | Acanthaceae | Girbia | H | MH 007 |
| <i>Hypoestes triflora</i> (Forssk.) Roem & Schult. | Acanthaceae | Girbia | H | MH 289 |
| <i>Impatiens tinctoria</i> A. Rich | Balsaminaceae | Gursht | H | MH 161 |
| <i>Ischaemum afrum</i> (J.F. Gmel.) Dandy | Poaceae | | H | MH 209 |
| <i>Jasminum abyssinicum</i> Hochst. ex DC. | Oleaceae | Tenbelel | WC | MH 292 |
| <i>Jasminum grandiflorum</i> L. | Oleaceae | Habi tselim | WC | MH 035 |
| <i>Juniperus procera</i> Hochst. ex Endl. | Cupressaceae | Tshdi adi | T | MH 060 |
| <i>Justicia ladanoides</i> Lam. | Acanthaceae | | H | MH 179 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|------------------|-----------------|-------|-----------|
| <i>Justicia schimperiana</i> (Hochst. ex Nees) T. Anders. | Acanthaceae | Shmeda | S | MH 061 |
| <i>Kalanchoe petitiiana</i> A. Rich. | Crassulaceae | Awo awo zbei | H | MH 036 |
| <i>Kickxia elatine</i> (L.) Dumort. | Scrophulariaceae | | H | MH 324 |
| <i>Kleinia odora</i> (Forssk) DC. | Asteraceae | Bierere | S | MH 194 |
| <i>Kniphofia foliosa</i> Hochst. | Asphodelaceae | Ashenda | H | MH 028 |
| <i>Laggera crispata</i> (Vahl) Hepper and Wood | Asteraceae | Tuem chena | H | MH 103 |
| <i>Laggera tomentosa</i> (Sch. Bip. ex A. Rich.) Oliv. & Hiern | Asteraceae | Kaskanso | S | MH 099 |
| <i>Lantana trifolia</i> L. | Verbenaceae | | S | MH 142 |
| <i>Lantana viburnoides</i> (Forssk.) Vahl | Verbenaceae | | S | MH 014 |
| <i>Lathyrus sphaericus</i> Retz. | Fabaceae | | H | MH 295 |
| <i>Leonotis ocymifolia</i> (Burm. f.) Iwarsson | Lamiaceae | | S | MH 058 |
| <i>Leptadenia hastata</i> (Pers.) Decne. | Asclepiadaceae | Qntt | WC | MH 116 |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | Shiwa qerni | S | MH 047 |
| <i>Lippia adoensis</i> Hochst. ex Walp. | Verbenaceae | | S | MH 083 |
| <i>Malva verticillata</i> L. | Malvaceae | Enkftha | H | MH 172 |
| <i>Maytenus arbutifolia</i> (A. Rich.) Wilczek | Celastraceae | Ats Ats | T | MH 092 |
| <i>Maytenus senegalensis</i> (Lam.) Exell | Celastraceae | Qebqeb | T | MH 072 |
| <i>Maytenus undata</i> (Thunb.) Blakelock | Celastraceae | Qfei | T | MH 091 |
| <i>Melinis repens</i> (Willd.) Zizka | Poaceae | | H | MH 224 |
| <i>Myrica salicifolia</i> A. Rich. | Myricaceae | Shinet | T | MH 034 |
| <i>Myrsine africana</i> L. | Myrsinaceae | Qechemo | T | MH 046 |
| <i>Nidorella rsedifolia</i> DC. | Asteraceae | | H | MH 190 |
| <i>Notholaena marantae</i> (L.) Desv. | Sinopteridaceae | | H | MH 232 |
| <i>Nuxia congesta</i> R.Br. ex Fresen. | Loganiaceae | Tekarea | T | MH 041 |
| <i>Ocimum lamiifolium</i> Hochst. ex Benth. | Lamiaceae | Tsomer | S | MH 294 |
| <i>Ocimum stirbeyi</i> Schweinf. and Volk. | Lamiaceae | | S | MH 015 |
| <i>Oenanthe palustris</i> (Chiov.) Norman | Apiaceae | | H | MH 290 |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | MH 079 |
| <i>Opuntia ficus-indica</i> (L.) Miller | Cactaceae | Beles | S | MH 301 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|----------------|-----------------|-------|-----------|
| <i>Osyris quadripartita</i> Decn. | Santalaceae | Qetrets | T | MH 053 |
| <i>Otostegia fruticosa</i> (Forssk.) Schweinf. ex Penzig | Lamiaceae | Chamo/sasa | S | MH 113 |
| <i>Otostegia integrifolia</i> Benth. | Lamiaceae | Chiendog | S | MH 016 |
| <i>Otostegia tomentosa</i> A. Rich. | Lamiaceae | Sasa | S | MH 170 |
| <i>Oxygonum sinuatum</i> (Meisn.) Dammer | Polygonaceae | Kukaito | H | MH 114 |
| <i>Panicum maximum</i> Jacq. | Poaceae | | H | MH 211 |
| <i>Panicum monticola</i> Hook. f. | Poaceae | | H | MH 216 |
| <i>Parthenium hysterophorus</i> L. | Asteraceae | Diha nekel | H | MH 130 |
| <i>Pavetta abyssinica</i> Fresen. | Rubiaceae | Buna hibey | T | MH 089 |
| <i>Pavonia urens</i> Cav. | Malvaceae | Nacha | S | MH 010 |
| <i>Pennisetum thunbergii</i> Kunth | Poaceae | Waz wazo | H | MH 273 |
| <i>Pennisetum villosum</i> Fresen. | Poaceae | Saeri hibey | H | MH 207 |
| <i>Periploca linearifolia</i> Quart.-Dill. & A. Rich. | Asclepiadaceae | Qeyh hareg | WC | MH 135 |
| <i>Phragmanthera regularis</i> (Sprague) M. Gilbert | Loranthaceae | Diqala | S | MH 287 |
| <i>Phyllanthus mooneyi</i> M Gilbert | Euphorbiaceae | | H | MH 297 |
| <i>Phytolacca dodecandra</i> L 'Herit. | Phytolaccaceae | Shibti | S | MH 133 |
| <i>Pittosporum viridiflorum</i> Sims | Pittosporaceae | May liho | T | MH 064 |
| <i>Plantago lanceolata</i> L. | Plantaginaceae | | H | MH 080 |
| <i>Plectranthus lanuginosus</i> (Hochst. ex Benth.) Agnew | Lamiaceae | Guemfer | H | MH 278 |
| <i>Podocarpus falcatus</i> (Thunb.) R. B. ex. Mirb. | Podocarpaceae | Zigba | T | MH 065 |
| <i>Polygala abyssinica</i> Fres. | Polygalaceae | | H | MH 0139 |
| <i>Polygala muratii</i> Jacq.-Felix | Polygalaceae | | H | MH 0185 |
| <i>Polygala steudneri</i> Chod. | Polygalaceae | | S | MH 0173 |
| <i>Prosopis juliflora</i> (Sw.) DC. | Fabaceae | Woyane | T | MH 087 |
| <i>Psydrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | T | MH 098 |
| <i>Pteris dentata</i> Forssk. | Pteridaceae | | H | MH 230 |
| <i>Pterolobium stellatum</i> (Forssk.) Brenan | Fabaceae | Qonteftefe | WC | MH 076 |
| <i>Pupalia lappacea</i> (L.) A. Juss. | Amaranthaceae | Teneg | HC | MH 146 |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | T | MH 040 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|---|----------------|-----------------|-------|-----------|
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | T | MH 118 |
| <i>Rhus retinorrhoea</i> Oliv. | Anacardiaceae | Nefasito/ amus | S | MH 093 |
| <i>Rhus ruspolii</i> Engl. | Anacardiaceae | Atami | S | MH 279 |
| <i>Rhus vulgaris</i> Meikle | Anacardiaceae | Atami | S | MH 119 |
| <i>Rhynchosia ferruginea</i> A. Rich. | Fabaceae | | HC | MH 285 |
| <i>Rhynchosia minima</i> (L.) DC. | Fabaceae | | WC | MH 283 |
| <i>Rhynchosia stipulosa</i> A. Rich. | Fabaceae | | WC | MH 284 |
| <i>Ricinus communis</i> L. | Euphorbiaceae | Gulei | H | MH 084 |
| <i>Rosa abyssinica</i> Lindley | Rosaceae | Qega | WC | MH 051 |
| <i>Rubus apetalus</i> Poir. | Rosaceae | Mongolil | WC | MH 270 |
| <i>Rubus steudneri</i> Schweinf. | Rosaceae | Mongilil | WC | MH 074 |
| <i>Rumex abyssinicus</i> Jacq. | Polygonaceae | Meqmeqo | H | MH 031 |
| <i>Rumex nepalensis</i> Spreng. | Polygonaceae | Dengele | H | MH 071 |
| <i>Rumex nervosus</i> Vahl, | Polygonaceae | Hehot | S | MH 054 |
| <i>Sageretia thea</i> (Osbeck) M.C. Johnston | Rhamnaceae | Qenchil egam | S | MH 063 |
| <i>Salvia merjamie</i> Forssk. | Lamiaceae | | H | MH 171 |
| <i>Sansevieria forskaoliana</i> (Schult.f.) Hepper & Wood | Dracaenaceae | Qacha | H | MH 140 |
| <i>Sarcostemma viminale</i> (L.) R. Br. | Asclepiadaceae | Elbi | H | MH 110 |
| <i>Satureja abyssinica</i> (Benth.) Briq. | Lamiaceae | | H | MH 298 |
| <i>Satureja punctata</i> (Benth.) Briq. | Lamiaceae | | H | MH 052 |
| <i>Scabiosa columbaria</i> L. | Dipsacaceae | | H | MH 177 |
| <i>Senecio hadiensis</i> Forssk. | Asteraceae | Sehum atali | HC | MH 048 |
| <i>Senecio ragazzii</i> Chiov. | Asteraceae | | H | MH 189 |
| <i>Senna occidentalis</i> (L) Link | Fabaceae | Hambohambo | H | MH 164 |
| <i>Sida schimperiana</i> Hochst. ex A. Rich. | Malvaceae | Tsfrer | S | MH 003 |
| <i>Smilax anceps</i> Willd. | Smilacaceae | Hareg nebri | WC | MH 026 |
| <i>Smilax aspera</i> L. | Smilacaceae | Hareg nebri | WC | MH 078 |
| <i>Solanum adoense</i> Hoehst. ex A. Rich. | Solanaceae | Engule abyi | S | MH 039 |
| <i>Solanum anguivi</i> Lam. | Solanaceae | Engule qola | S | MH 086 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|--|------------------|-----------------|-------|-----------|
| <i>Solanum hastifolium</i> Hochst. ex Dunal in DC. | Solanaceae | Alamo kelbi | S | MH 272 |
| <i>Solanum incanum</i> L. | Solanaceae | Engule nieshtey | S | MH 105 |
| <i>Solanum marginatum</i> L.f. | Solanaceae | Engule dogae | S | MH 090 |
| <i>Solanum memphiticum</i> Gmel. | Solanaceae | | H | MH 178 |
| <i>Solanum nigrum</i> L. | Solanaceae | Alamo | H | MH 134 |
| <i>Solanum schimperianum</i> Hochst. ex A. Rich. | Solanaceae | Berbere awald | S | MH 020 |
| <i>Solanum sinaicum</i> Boiss. | Solanaceae | | H | MH 160 |
| <i>Sonchus oleraceus</i> L. | Asteraceae | | H | MH 187 |
| <i>Sporobolus discosporus</i> Nees | Poaceae | | H | MH 220 |
| <i>Sporobolus ioclados</i> (Trin.) Nees | Poaceae | Saeri tefetafo | H | MH 221 |
| <i>Sporobolus pyramidalis</i> P. Beauv. | Poaceae | | H | MH 222 |
| <i>Stephania abyssinica</i> (Quart. Dillon & A. Rich.) Walp. | Menispermaceae | Hareg mintaro | HC | MH 043 |
| <i>Swertia lugardae</i> Bullock | Gentianaceae | | H | MH 186 |
| <i>Tagetes minuta</i> L. | Asteraceae | Chenawi qotsli | H | MH 012 |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | T | MH 101 |
| <i>Tephrosia hochstetteri</i> Chiov. | Fabaceae | | H | MH 117 |
| <i>Tephrosia interrupta</i> Hochst. and Steud. ex Engl. | Fabaceae | | S | MH 288 |
| <i>Thalictrum rhynchocarpum</i> Dill. & A. Rich. | Ranunculaceae | Sre bizu | H | MH 162 |
| <i>Thymus schimperi</i> Ronniger | Lamiaceae | Teshine | H | MH 156 |
| <i>Toddalia asiatica</i> (L.) Lam. | Rosaceae | | WC | MH 271 |
| <i>Tragia cinerea</i> (Pax) M.G. Gilbert & Radcl.-Sm. | Euphorbiaceae | Harengama | HC | MH 004 |
| <i>Trifolium campestre</i> Schreb. | Fabaceae | | H | MH 163 |
| <i>Urtica simensis</i> Steudel | Urticaceae | Amea | H | MH 097 |
| <i>Verbascum sinaiticum</i> Benth. | Scrophulariaceae | Trnake | H | MH 009 |
| <i>Vernonia auriculifera</i> Hiern | Asteraceae | | S | MH 188 |
| <i>Vernonia hymenolepis</i> A. Rich. | Asteraceae | Gudet | S | MH 199 |
| <i>Vernonia leopoldi</i> (Sch. Bip. ex Walp.) Vatke | Asteraceae | | H | MH 201 |
| <i>Vernonia rueppellii</i> Sch. Bip. ex Walp. | Asteraceae | | S | MH 107 |
| <i>Withania somnifera</i> (L.) Dunal | Solanaceae | Gulho | S | MH 082 |

| Scientific name | Family | Vernacular name | Habit | Coll. No. |
|--|---------------|-----------------|-------|-----------|
| <i>Xanthium spinosum</i> L. | Asteraceae | Melhas ansti | H | MH 081 |
| <i>Zehneria scabra</i> (L. f) Sond. | Cucurbitaceae | Hareg resha | HC | MH 070 |
| <i>Ziziphus mucronata</i> Willd. | Rhamnaceae | Qunqura | T | MH 125 |
| <i>Ziziphus spina-christi</i> (L.) Desf. | Rhamnaceae | Qunqura | S | MH 126 |

Appendix 2. Endemic species recorded from the study area, their IUCN status and geographical distributions, (LC = Least Concern; NT = Near Threatened).

| Scientific name | Family | Habit | IUCN status | Altitude | Floristic Region |
|--|----------------|-------|-------------|--------------|--------------------------------|
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | S | NT | 1600-3100 m. | TU, GD, WU, SU, WG, SD |
| <i>Gomphocarpus purpurascens</i> A. Rich. | Asclepiadaceae | S | LC | 1500-2500 m. | EW, TU, GD, GJ, SU, AR, KF, HA |
| <i>Kniphofia foliosa</i> Hochst. | Asphodelaceae | H | LC | 2500-4000 m | TU GD GJ WU SU AR BA,HA |
| <i>Laggera tomentosa</i> (Sch. Bip. ex A. Rich.) Oliv. & Hierm | Asteraceae | S | LC | 2345-2950 m. | TU, GD,GJ, WU, SU, HA |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | S | LC | 1300-2600 m. | TU, GD, GG, SD, BA |
| <i>Lippia adoensis</i> Hochst. ex Walp. | Verbenaceae | S | LC | 1900-2450 m. | TU, GJ, SU, AR, HA, KF, GG |
| <i>Phyllanthus mooneyi</i> M. Gilbert* | Euphorbiaceae | H | LC | 525-2100 m | SU AR IL KF GD,SD |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | T | LC | 1800-3000 m. | TU, WU, SU, WG, AR, BA, HA |
| <i>Rhynchosia stipulosa</i> A. Rich. | Fabaceae | WC | LC | 1000-800 m | TU,WG, GG |
| <i>Thymus schimperi</i> Ronniger | Lamiaceae | H | LC | 2250-4000 m | EW TU GD WU SU AR SD BA HA |
| <i>Urtica simensis</i> Steudel | Urticaceae | H | LC | 1500-3400m | TU, GD, GJ SU AR BA SD |
| <i>Vernonia leopoldi</i> (Sch. Bip. ex Walp.) Vatke | Asteraceae | H | LC | 1850-2850 m. | TU, GD, GJ, SU, WG, KF, HA |
| <i>Vernonia rueppellii</i> Sch. Bip. ex Walp. | Asteraceae | S | LC | 2150-3000 m. | TU, GD, SU, AR, BA, SD, KF, HA |

Source: Hedber et al. (1989,1995,2003,2004,2006,2009a,2009b,; Edwards et al. 1995, 1997,2000)

*1 New records for TU Floristic Region

Appendix 3. Soil seed bank species obtained via germination.

| Scientific name | Family | Habit | Coll. No. |
|--|--------------------|-------|-----------|
| <i>Achyranthes aspera</i> L. | Amaranthaceae | H | MH 243 |
| <i>Actiniopteris radiata</i> (Sw.) Link | Actiniopteridaceae | H | MH 325 |
| <i>Aerva lanata</i> (L.) Juss. ex Schultes | Amaranthaceae | H | MH 256 |
| <i>Amaranthus hybridus</i> L. | Amaranthaceae | H | MH 252 |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | S | MH 262 |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | S | MH 303 |
| <i>Chenopodium murale</i> L. | Chenopodiaceae | H | MH 240 |
| <i>Chenopodium opulifolium</i> Schrader ex Koch & Ziz. | Chenopodiaceae | H | MH 250 |
| <i>Chloris virgata</i> Sw. | Poaceae | H | MH 245 |
| <i>Commelina africana</i> L. | Commelinaceae | H | MH 263 |
| <i>Conyza</i> sp. = Smeds | Asteraceae | H | MH 313 |
| <i>Crassula schimperi</i> Fisch. & Mey. | Crassulaceae | H | MH 324 |
| <i>Cynodon dactylon</i> (L.) Pers. | Poaceae | H | MH 323 |
| <i>Cyperus bulbosus</i> Vahl | Cyperaceae | H | MH 266 |
| <i>Cyperus flavescens</i> L. | Cyperaceae | H | MH 274 |
| <i>Cyperus polystachyos</i> Rottb. | Cyperaceae | H | MH 265 |
| <i>Cyperus pulchellus</i> R.Br. | Cyperaceae | H | MH 264 |
| <i>Cyperus rubicundus</i> Vahl | Cyperaceae | H | MH 254 |
| <i>Digitaria nuda</i> Schumach. | Poaceae | H | MH 247 |
| <i>Digitaria velutina</i> (Forssk.) P. Beauv. | Poaceae | H | MH 242 |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | S | MH 321 |
| <i>Eragrostis cilianensis</i> (All.) Vign. ex Janch. | Poaceae | H | MH 246 |
| <i>Eragrostis cylindriflora</i> Hochst. | Poaceae | H | MH 241 |
| <i>Eragrostis schweinfurthii</i> Chiov. | Poaceae | H | MH 305 |
| <i>Erucastrum abyssinicum</i> (A. Rich.) R.E. Fries | Brassicaceae | H | MH 253 |
| <i>Ficus palmata</i> Forssk. | Moraceae | S | MH 304 |
| <i>Galinsoga parviflora</i> Cav. | Asteraceae | H | MH 238 |
| <i>Galium scioanum</i> Chiov. | Rubiaceae | H | MH 258 |
| <i>Geranium arabicum</i> Forssk. | Geraniaceae | H | MH 261 |
| <i>Harpachne schimperi</i> Hochst. ex A. Rich. | Poaceae | H | MH 249 |
| <i>Helichrysum</i> sp. Mill. | Asteraceae | S | MH 312 |

| Scientific name | Family | Habit | Coll. No. |
|--|------------------|--------------|------------------|
| <i>Hypoestes forskaolii</i> (Vahl) R. Br. | Acanthaceae | H | MH 244 |
| <i>Hypoestes triflora</i> (Forssk.) Roem & Schult. | Acanthaceae | H | MH 317 |
| <i>Kalanchoe petitiiana</i> A. Rich. | Crassulaceae | H | MH 315 |
| <i>Laggera crispata</i> (Vahl) Hepper & Wood | Asteraceae | H | MH 307 |
| <i>Ochthochloa compressa</i> (Forssk.) Hilu | Poaceae | H | MH 316 |
| <i>Ocimum lamiifolium</i> Hochst. ex Benth. | Lamiaceae | S | MH 260 |
| <i>Opuntia ficus-indica</i> (L.) Miller | Cactaceae | H | MH 319 |
| <i>Oxalis procumbens</i> Steud. ex A. Rich | Oxalidaceae | H | MH 259 |
| <i>Plectranthus</i> sp. L Her. | Lamiaceae | H | MH 310 |
| <i>Poa leptoclada</i> Hochst. ex A. Rich. | Poaceae | H | MH 322 |
| <i>Polycarpon tetraphyllum</i> (L.) L. | Caryophyllaceae | H | MH 308 |
| <i>Rostraria cristata</i> (L.) Tzvelev | Poaceae | H | MH 239 |
| <i>Rumex nervosus</i> Vahl | Polygonaceae | S | MH 256 |
| <i>Senecio</i> sp. L. | Asteraceae | H | MH 314 |
| <i>Solanum adoense</i> Hochst. ex A. Rich. | Solanaceae | S | MH 311 |
| <i>Solanum nigrum</i> L. | Solanaceae | H | MH 320 |
| <i>Solanum villosum</i> Mill. | Solanaceae | H | MH 248 |
| <i>Sonchus oleraceus</i> L. | Asteraceae | H | MH 306 |
| <i>Tagetes minuta</i> L. | Asteraceae | H | MH 251 |
| <i>Trifolium campestre</i> Schreb. | Fabaceae | H | MH 255 |
| <i>Verbascum sinaiticum</i> Benth. | Scrophulariaceae | H | MH 318 |
| <i>Vernonia</i> sp. Schreb. | Asteraceae | S | MH 321 |
| <i>Zehneria scabra</i> (L. f) Sond. | Cucurbitaceae | HC | MH 309 |

Appendix 4. Medicinal plants used to treat human ailments (L = Leaf; R/L = Root and Leaf; B= Bark; R = Root; La = Latex; YS = Young Shoot; F = Fruit; L/B = Leaf and Bark; S =Stem; WP = Whole plant; WC = Woody climber; HC = Herbaceous climber)

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|---|---------------|----------------|-------|------------------|---|---|
| <i>Abutilon longicuspe</i> Hochst. ex A. Rich. | Malvaceae | Sa'da buwak | S | L | Cholera | Crush and drink |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | T | R/L L | Cancer Gulhay* | Crush and tie on the infected part Crush and drink |
| <i>Acacia senegal</i> (L.) Willd. | Fabaceae | Sewansa | T | B | Troma | Crush, boil, smoke and drink a cup of glass |
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Machelo | H | R/L L R | Gulhay* Tonsillitis Snakebite | Chew, swallow and drink Chew and swallow Chew |
| <i>Acokanthera schimperi</i> (A. DC.) Schweinf. | Apocynaceae | Merenz | T | L L | Malaria Degehabe* | Crush, drink juiceand apply to nose Crush and drink |
| <i>Aloe camperi</i> Schweinf. | Aloaceae | Ere | S | L | Malaria | Crush, drink juice in a cup of glass |
| <i>Argemone mexicana</i> L. | Papaveraceae | Medafe | H | La. | Wound | Smearing latex to the affected part |
| <i>Artemisia abyssinica</i> Sch. Bip. ex A. Rich. | Asteraceae | Chena baria | H | L | Headache | Crush and apply to nose |
| <i>Asparagus africanus</i> Lam. | Asparagaceae | Kestenosto | WC | R/L | Dandruff | Crush, mix with butter and rub on the head |
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | T | L L L L | Cholera Gulhay* Hangnail Dandruff | Crush and drink in cup of glass Crush and drink Crush, mix with butter and tie on the infected part Crush, mix with butter and rub on the head |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | Tebeb | S | R L YS | Anthrax Tonsillitis Blood clotting Febrile illness | Crush, mix with honey and tie on the infected part Chew and swallow Crush, immediate tie with fresh juice Crush and drink in cup of glass |

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|--|---------------|----------------|--|-----------|-----------------|---|
| <i>Bidens pilosa</i> L. | Asteraceae | Tselim Teneg | H | L | Anthrax | Crush, mix with <i>Commicarpus pedunculatus</i> leaf, <i>Heliotropium strigosum</i> leaf with with honey and rub/tie on the infected part |
| <i>Calpurnia aurea</i> (Ait.) Benth. | Fabaceae | Chrenchah | S | L | Evil eye | Crush, mix with fresh water taken at the morning from the source and wash the whole body |
| <i>Capparis tomentosa</i> Lam. | Capparidaceae | Harengama | WC | R | Evil eye | Crush and smoke |
| <i>Carissa spinarum</i> L. | Apocynaceae | Egam | S | R | Febrile illness | Crush and smoke |
| <i>Clematis simensis</i> Fresen. | Ranunculaceae | Hareghazo | HC | R/L | Chibti* | Crush and drink in cup of glass |
| | | | | R | Cancer | Crush, mix with butter and tie on the infected |
| | | | | R/L | Kidney | Crush, mix with honey and drink in small amount |
| | | | | L | Cancer | Crush and tie |
| | | | | L | Dislocated | Crush and tie |
| | R | Paralyze | Mix with <i>Kniphophia pumila</i> root, <i>Plectranthus punctatus</i> leaf and stored together for three days and wash the damage person | | | |
| <i>Clerodendrum myricoides</i> (Hochst.) Vatke | Lamiaceae | Shewha | S | R | Snakebite | Chew, crush and drink |
| <i>Clutia lanceolata</i> Forssk. | Euphorbiaceae | Hirtmtmo | S | L | Berle* | Crush, mix <i>Olea europaea</i> subsp. <i>cuspidata</i> bark, <i>Asparagus africanus</i> fruit and mix with butter and rub on the infected part |
| <i>Commicarpus pedunculatus</i> (A.Rich.) Cufod. | Nyctaginaceae | Ezni anchwa | H | L | Swollen | Crush and rub/tie on the infected part |
| <i>Crotalaria incana</i> L. | Fabaceae | Shamto amrakut | S | L | Bullet | Crush, dried, mix with butter and rub on the affected part |
| | | | | L | Anthrax | Crush, mix with honey and tie in the infected part |

| <i>Cyphostemma adenocaula</i> (Steud. ex A. Rich.) Descoings ex Wild and Drummond | Vitaceae | Aserkuka | HC | R | Fumigation | Crush and smoke |
|--|---------------|--------------------|-------|-----------|-----------------|--|
| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
| <i>Datura stramonium</i> L. | Solanaceae | Mestenagr | H | L | Hemorrhoid | Crush and tie on the infected part |
| | | | | L | Tuberculosis | Crush and tie on the infected part |
| | | | | L | Werchi* | Crush, mix with butter and rub |
| | | | | L | Wound | Crush and tie on the infected part |
| | | | | F | Toothache | Burn, mix with butter and smoke |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | L | Swollen | Crush and drink |
| | | | | L | Hibri semay* | Crush, mix with honey and rub on the infected part |
| <i>Echinops pappi</i> Chiov. | Asteraceae | Dander | S | R | Headache | Chew |
| <i>Eucalyptus globulus</i> Labill. | Myrtaceae | Tseada bahirzaf | T | L | Asthma | Boil and smoke |
| | | | | L | Febrile illness | Boil and smoke |
| <i>Euclea racemosa</i> subsp. <i>schimperi</i> (A. DC.) White | Ebenaceae | Kuleo | S | R | Goiter | Chew |
| <i>Euphorbia petitiiana</i> A. Rich. | Euphorbiaceae | Hinzuquzq | H | R | Jaundice | Crush, mix with butter and drink |
| <i>Grewia mollis</i> A. Juss. | Tiliaceae | Rewey | T | L | Toothache | Crush and tie |
| | | | | L | Bonbelea* | Crush and drink with cup of glass |
| <i>Heliotropium strigosum</i> Willd. | Boraginaceae | Amam gmel | H | L | Wound | Crush and tie in fresh on the affected part |
| <i>Juniperus procera</i> Hochst. ex Endl. | Cupressaceae | Tshdi adi | T | L | Amoeba | Crush, mix with water, filter and drink |
| <i>Justicia schimperiana</i> (Hochst. ex Nees) T. Anders. | Acanthaceae | Shmeda | S | L | Jaundice | Crush, filter and mix with cheese and drink small cup of glass |
| | | | | | Jaundice | Crush and drink with small amount |
| <i>Kalanchoe petitiiana</i> A. Rich. | Crassulaceae | Awo awo zbei | H | L | Swollen | Crush and drink |
| | | | | | Evulitis | Crush and apply to nose, drink, mix with butter and tie |

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|---|----------------|---------------|-------|-----------|---------------------|---|
| <i>Laggera tomentosa</i> (Sch. Bip. ex A. Rich.) Oliv. and Hiern | Asteraceae | Kaskanso | S | L | Satan | Crush, mix with water and wash every monrning |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | Shiwa qerni | S | L | Wekei* | Crush and drink with cup of glass |
| <i>Myrica salicifolia</i> A. Rich. | Myricaceae | Shinet | T | B | Snakebite Cancer | Crush and drink Crush, dried and smoke through nose |
| <i>Myrsine africana</i> L. | Myrsinaceae | Qechemo | S | B | Cough | Crush, dried/fresh and smoke |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | F | Tapeworm | Crush and drink in cup of glass |
| | | | | L | Febrile illness | Crush and smoke |
| | | | | L/B | Cholera | Crush and drink |
| | | | | S | Typhus | Burn and smoke |
| | | | | L | Shanbu* | Crush and drink |
| | | | | L | Fnfan* | Curl the leaf and insert through anus |
| | | | | L | Eye infection | Crush and smoke |
| <i>Phytolacca dodecandra</i> L 'Herit. | Phytolaccaceae | Shibtu | S | L | Stomachache | Crush, mix with water and drink |
| | | | | R | Jaundice | Crush, filter and drink with small cup |
| <i>Podocarpus falcatus</i> (Thunb.) R. B. ex. Mirb. | Podocarpaceae | Zigba | T | L | Evil eye | Crush, mix with water and wash the body |
| | | | | L | Intelligence | Crush, mix water and wash the whole body |
| | | | | S | Evil eye | Tie on the neck |
| | | | | L | Satan | Crush, mix with water and wash |
| | | | | B | Evil eye | Crush, mix with <i>Rumex nervosus</i> and wash |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | YS | Hebri semay* | Crush, mix white shiny stone and butter and then rub on the infected part |
| <i>Rubus apetalous</i> Poir. | Rosaceae | Mongolil | S | R | Cancer | Crush and tie |
| <i>Rumex nepalensis</i> Spreng. | Polygonaceae | Dengele | H | R/L | Degehabe* | Crush and drink |
| | | | | L | Evil eye | Crush, mix with fresh water and |

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|--|------------------|---------------|-------|-----------|-----------------|---|
| | | | | L | Itching | wash the whole body |
| | | | | | | Crush and wash |
| <i>Solanum marginatum</i> L.f. | Solanaceae | Engule dogae | S | R | Typhus | Crush and drink every three days in small amount |
| <i>Verbascum sinaiticum</i> Benth. | Scrophulariaceae | Trnake | H | R | Jaundice | Crush, filter and drink a cup of glass |
| | | | | R | Jaundice | Crush and drink |
| | | | | L/R | Bullet | Crush and tie |
| | | | | R | Jaundice | Crush, mix with cheese and drink one cup of glass for every three morning |
| <i>Withania somnifera</i> (L.) Dunal | Solanaceae | Gulho | S | R | Malaria | Burn and smoke |
| | | | | L/R | Satan | Crush and smoke |
| | | | | L | Wound | Crush and rub |
| | | | | R | Febrile | Crush and smoke |
| | | | | R | Dislocated | Crush, mix with butter and rub on the affected part |
| <i>Zehneria scabra</i> (Linn. f) Sond. | Cucurbitaceae | Hareg resha | HC | L | Febrile illness | Crush and smoke |

Appendix 5. Medicinal plants used to treat livestock

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|---|---------------|---------------|-------|-----------|---------------|---|
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Machelo | H | R | Eye infection | Chew and smear droplet in to the infected part |
| <i>Aloe camperi</i> Schweinf. | Aloaceae | Ere | S | L | Dislocated | Tie on the tail of the infected animal |
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | T | WP | Degehabe* | Tie the animal on the stem of the live plant of <i>Balanites aegyptiaca</i> |
| <i>Buddleja polystachya</i> Fresen. | Loganiaceae | Metere | S | L | Leeches | Crush and apply to the nose |
| <i>Clutia lanceolata</i> Forssk. | Euphorbiaceae | Hirtmtmo | S | F | Shanbu* | Crush, drink juice and apply to to the nose |
| <i>Cyphostemma adenocaula</i> (Steud. ex A. Rich.) Desc. ex | Vitaceae | Aserkuka | HC | R | Leeches | Crush, filter and apply to the nose |

| Wild and Drummond <i>Ehretia cymosa</i> Thonn. | Boraginaceae | Aulaga | T | S | Dislocated | Part of stem that have formed circle hole at the center and insert in to metal and hold the stem and burn the affected part through the hot metal |
|---|------------------|---------------|-------|-----------|---------------|---|
| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
| <i>Heliotropium strigosum</i> Willd. | Boraginaceae | Amam gmel | H | L | Dislocated | Tie on the tail of the affected animal |
| <i>Hypoestes forskoalii</i> (Vahl) R. Br. | Acanthaceae | Girbia | H | R | Eye infection | Chew and smear in to the infected part |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | Shiwa qerni | S | L | Eye infection | Chew and smear droplet in the infected part |
| <i>Plectranthus punctatus</i> (L. f.) L'Her. | Lamiaceae | Karewo-awalid | H | L | Zgag* | Crush and apply to the nose |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | L | Fnfan* | Curl the leaves and tie via fiber and insert through its anus |
| <i>Verbascum sinaiticum</i> Benth. | Scrophulariaceae | Trnake | H | R/L | Pancreas | Crush, drink and apply to the nose |

Appendix 6. Medicinal plants used to treat both human and livestock

| Scientific name | Family name | Tigrigna name | Habit | Part used | Used for | Preparation and application |
|--------------------------------------|---------------|---------------|-------|-----------|---------------|--|
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | L/R | Dislocated | Crush and tie |
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Machelo | H | R | Eye infection | Crush, juice and smear drop of juice on the infected part |
| <i>Clematis simensis</i> Fresen. | Ranunculaceae | Hareghazo | HC | L | Efni* | Crush, apply fresh without additives, mix with butter and tie for human. For livestock surge and insert it |
| <i>Datura stramonium</i> L. | Solanaceae | Mestenagr | H | L | Dislocated | Crush and tie |
| <i>Erica arborea</i> L. | Ericaceae | Hasti | T | P/N | Anas* | Swallow honey collected from <i>Erica arborea</i> , rub on the infected part |
| <i>Grewia mollis</i> A. Juss. | Tiliaceae | Rewey | T | L | Dislocated | Crush and tie |
| <i>Heliotropium strigosum</i> Willd. | Boraginaceae | Amam gmel | H | L | Dislocated | Crush and tie |
| <i>Malva verticillata</i> L. | Malvaceae | Enkftha | H | L | Anthrax | Crush, mix with honey and tie on the affected part |

Appendix 7. Significance test between gender groups

Group Statistics

| | Gender | N | Mean | Std. Deviation | Std. Error Mean |
|------------------|--------|-----|------|----------------|-----------------|
| Medicinal Plants | Male | 200 | 4.21 | 9.203 | 0.651 |
| | Female | 139 | 0.32 | 1.395 | 0.118 |

Independent Samples Test

| | Levene's Test for Equality of Variances | | t-test for Equality of Means | | | | | | |
|------------------|---|-------|------------------------------|---------|-----------------|-----------------|-----------------------|---|-------|
| | F | Sig. | T | df | Sig. (2-tailed) | Mean Difference | Std. Error Difference | 95% Confidence Interval of the Difference | |
| | | | | | | | | Lower | Upper |
| Medicinal Plants | 69.799 | 0.000 | 4.937 | 337 | 0.000 | 3.886 | 0.787 | 2.338 | 5.435 |
| | | | 5.876 | 212.035 | 0.000 | 3.886 | 0.661 | 2.582 | 5.190 |

Appendix 8. Significance test between age groups

Group Statistics

| | Age | N | Mean | Std. Deviation | Std. Error Mean |
|-----------------|---------------------|-----|------|----------------|-----------------|
| Medicinal plant | Young members (<50) | 252 | 1.61 | 5.990 | 0.377 |
| | Old members (>50) | 87 | 2.66 | 6.220 | 0.667 |

Independent Samples Test

| | Levene's Test for Equality of Variances | | t-test for Equality of Means | | | | | | |
|------------------|---|-------|------------------------------|---------|-----------------|-----------------|-----------------------|---|-------|
| | F | Sig. | T | df | Sig. (2-tailed) | Mean Difference | Std. Error Difference | 95% Confidence Interval of the Difference | |
| | | | | | | | | Lower | Upper |
| Medicinal plants | 4.683 | 0.031 | -1.388 | 337 | 0.166 | -1.044 | 0.752 | -2.524 | 0.436 |
| | | | -1.363 | 144.792 | 0.175 | -1.044 | 0.766 | -2.559 | 0.470 |

Appendix 9. Significance test between education groups

Group Statistics

| | Education | N | Mean | Std. Deviation | Std. Error Mean |
|------------------|------------|-----|------|----------------|-----------------|
| Medicinal plants | Literate | 140 | 1.63 | 4.265 | 0.360 |
| | Illiterate | 199 | 3.55 | 8.289 | 0.588 |

Independent Samples Test

| | Levene's Test for Equality of Variances | | t-test for Equality of Means | | | | | | |
|-----------------|---|-------|------------------------------|--------|-----------------|-----------------|-----------------------|---|--------|
| | F | Sig. | T | df | Sig. (2-tailed) | Mean Difference | Std. Error Difference | 95% Confidence Interval of the Difference | |
| | | | | | | | | Lower | Upper |
| Medicinal plant | 15.527 | 0.000 | -2.521 | 337 | 0.012 | -1.924 | .763 | -3.425 | -0.423 |
| | | | -2.791 | 312.14 | 0.006 | -1.924 | .689 | -3.281 | -0.568 |

Appendix 10. Significance test between informant categories

Group Statistics

| | Informant | N | Mean | Std. Deviation | Std. Error Mean |
|-----------|-----------|-----|-------|----------------|-----------------|
| Medicinal | Key | 30 | 14.57 | 13.736 | 2.508 |
| | General | 309 | .34 | 1.107 | 0.063 |

Independent Samples T- Test

| | Levene's Test for Equality of Variances | | t-test for Equality of Means | | | | | | |
|------------------|---|------|------------------------------|--------|-----------------|-----------------|-----------------------|---|--------|
| | F | Sig. | t | Df | Sig. (2-tailed) | Mean Difference | Std. Error Difference | 95% Confidence Interval of the Difference | |
| | | | | | | | | Lower | Upper |
| Medicinal plants | 392.793 | .000 | 17.858 | 337 | 0.000 | 14.227 | 0.797 | 12.660 | 15.794 |
| | | | 5.671 | 29.037 | 0.000 | 14.227 | 2.509 | 9.097 | 19.357 |

Appendix 11. Wild edible plants (F = Fruit)

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|--|----------------|---------------|-------|-----------|
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | S | F |
| <i>Carissa spinarum</i> L. | Apocynaceae | Egam | S | F |
| <i>Dovyalis abyssinica</i> (A. Rich) Warb. | Flacourtiaceae | Mongolhats | S | F |
| <i>Dovyalis verrucosa</i> (Hochst.) Warb. | Flacourtiaceae | Teumtegna | S | F |
| <i>Ficus sur</i> Forssk. | Moraceae | Shamfa | T | F |
| <i>Ficus sycomorus</i> L. | Moraceae | Sagla | T | F |
| <i>Grewia mollis</i> A. Juss. | Tiliaceae | Rewey | S | F |
| <i>Myrsine africana</i> L. | Myrsinaceae | Qechemo | S | F |
| <i>Opuntia ficus-indica</i> (L.) Miller | Cactaceae | Beles | S | F |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | F |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | F |
| <i>Rosa abyssinica</i> Lindley | Rosaceae | Qega | S | F |
| <i>Rubus apetalous</i> Poir. | Rosaceae | Mongolil | S | F |
| <i>Rumex nervosus</i> Vahl | Polygonaceae | Hehot | S | YS |
| <i>Sageretia thea</i> (Osbeck) M.C. Johnston | Rhamnaceae | Qenchil egam | S | F |
| <i>Solanum nigrum</i> L. | Solanaceae | Alamo | S | F |
| <i>Ziziphus mucronata</i> Willd. | Rhamnaceae | Qunqura | S | F |

Appendix 12. Fuelwood (St. = Stem)

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|----------------|---------------|-------|-----------|
| <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Chea | T | St. |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | St. |
| <i>Acacia senegal</i> (L.) Willd. | Fabaceae | Sewansa | S | St. |
| <i>Acokanthera schimperi</i> (A. DC.) Schweinf. | Apocynaceae | Mrenz | S | St. |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | Tebeb | S | St. |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | S | St. |
| <i>Carissa spinarum</i> L. | Apocynaceae | Egam | S | St. |
| <i>Cassipourea malosana</i> (Baker) Alston | Rhizophoraceae | Tselim om | S | St. |
| <i>Celtis africana</i> Burm. f. | Ulmaceae | Beto Koma | T | St. |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | St. |
| <i>Dovyalis verrucosa</i> (Hochst.) Warb. | Flacourtiaceae | Teumtegna | S | St. |
| <i>Ekebergia capensis</i> Sparrm. | Meliaceae | Kot | S | St. |
| <i>Erica arborea</i> L. | Ericaceae | Hasti | S | St. |
| <i>Juniperus procera</i> Hochst. ex Endl. | Cupressaceae | Tshdi adi | T | St. |
| <i>Maytenus arbutifolia</i> (A. Rich.) Wilczek | Celastraceae | Ats Ats | S | St. |

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|---------------|---------------|-------|-----------|
| <i>Nuxia congesta</i> R.Br. ex Fresen. | Loganiaceae | Tekarea | T | St. |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | St. |
| <i>Podocarpus falcatus</i> (Thunb.) R. B. ex. Mirb. | Podocarpaceae | Zigba | T | St. |
| <i>Psydrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | S | St. |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | St. |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | St. |
| <i>Rumex nervosus</i> Vahl | Polygonaceae | Hehot | S | St. |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | T | St. |

Appendix 13. Farm implements

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|----------------|-----------------|-------|-----------|
| <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Chea | T | St. |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | St. |
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | S | St. |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | S | St. |
| <i>Cassipourea malosana</i> (Baker) Alston | Rhizophoraceae | Tselim om | S | St. |
| <i>Celtis africana</i> Burm. f. | Ulmaceae | Beto Koma | T | St. |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | St. |
| <i>Dovyalis verrucosa</i> (Hochst.) Warb. | Flacourtiaceae | Teumtegna | S | St. |
| <i>Ehretia cymosa</i> Thonn. | Boraginaceae | Aulaga | S | St. |
| <i>Ekebergia capensis</i> Sparrm. | Meliaceae | Kot | S | St. |
| <i>Eucalyptus globulus</i> Labill. | Myrtaceae | Tseada bahirzaf | T | St. |
| <i>Euclea racemosa</i> subsp. <i>schimperi</i> (A. DC.) White | Ebenaceae | Kuleo | S | St. |
| <i>Ficus sycomorus</i> L. | Moraceae | Sagla | T | St. |
| <i>Grewia mollis</i> A. Juss. | Tiliaceae | Rewey | S | St. |
| <i>Maytenus undata</i> (Thunb.) Blakelock | Celastraceae | Qfei | S | St. |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | St. |
| <i>Podocarpus falcatus</i> (Thunb.) R. B. ex. Mirb. | Podocarpaceae | Zigba | T | St. |
| <i>Psydrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | S | St. |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | St. |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | St. |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | T | St. |
| <i>Ziziphus mucronata</i> Willd. | Rhamnaceae | Qunqura | S | St. |

Appendix 14. Building materials

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|----------------|-----------------|-------|-----------|
| <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Chea | T | St. |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | St. |
| <i>Acacia senegal</i> (L.) Willd. | Fabaceae | Sewansa | S | St. |
| <i>Asparagus racemosus</i> Willd. | Asparagaceae | Kestenosto | S | St. |
| <i>Balanites aegyptiaca</i> (L.) Del. | Balanitaceae | Bedeno | S | St. |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | S | St. |
| <i>Cassipourea malosana</i> (Baker) Alston | Rhizophoraceae | Tselim om | S | St. |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | St. |
| <i>Dombeya torrida</i> (J.F.Gmel.) P. Bamps | Sterculiaceae | Buyak | S | St. |
| <i>Ekebergia capensis</i> Sparrmn. | Meliaceae | Kot | S | St. |
| <i>Eucalyptus globulus</i> Labill. | Myrtaceae | Tseada bahirzaf | T | St. |
| <i>Ficus sur</i> Forssk. | Moraceae | Shamfa | T | St. |
| <i>Justicia schimperiana</i> (Hochst. ex Nees) T. Anders. | Acanthaceae | Shmeda | S | St. |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | St. |
| <i>Podocarpus falcatus</i> (Thunb.) R. B. ex. Mirb. | Podocarpaceae | Zigba | T | St. |
| <i>Psydrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | S | St. |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | St. |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | T | St. |
| <i>Ziziphus mucronata</i> Willd. | Rhamnaceae | Qunqura | S | St. |

Appendix 15. Livestock fodder / forage (L = Leaf; P = Pod)

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|--|----------------|---------------|-------|-----------|
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | L/Pod |
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Machelo | H | L |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | Tebeb | S | L |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | S | L |
| <i>Cassipourea malosana</i> (Baker) Alston | Rhizophoraceae | Tselim om | S | L |
| <i>Celtis africana</i> Burm. f. | Ulmaceae | Beto Koma | T | L |
| <i>Clerodendrum myricoides</i> (Hochst.) Vatke | Lamiaceae | Shewha | S | L |
| <i>Cynodon dactylon</i> (L.) Pers. | Poaceae | Tehag | Gr. | L |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | L |
| <i>Erica arborea</i> L. | Ericaceae | Hasti | S | L |
| <i>Hyparrhenia hirta</i> (L.) Stapf | Poaceae | Saeri geza | Gr. | L |
| <i>Hypoestes forskoalii</i> (Vahl) R. Br. | Acanthaceae | Girbia | H | L |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | Shiwa qerni | S | L |

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|---------------|---------------|-------|-----------|
| <i>Maytenus arbutifolia</i> (A. Rich.) Wilcz | Celastraceae | Ats Ats | S | L |
| <i>Maytenus undata</i> (Thunb.) Blakelock | Celastraceae | Qfei | S | L |
| <i>Myrica salicifolia</i> A. Rich. | Myricaceae | Shinet | T | L |
| <i>Myrsine africana</i> L. | Myrsinaceae | Qechemo | S | L |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | L |
| <i>Opuntia ficus-indica</i> (L.) Miller | Cactaceae | Beles | S | L |
| <i>Psyrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | S | L |
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | L |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | L |
| <i>Rumex nervosus</i> Vahl | Polygonaceae | Hehot | S | L |
| <i>Sageretia thea</i> (Osbeck) M.C. Johnston | Rhamnaceae | Qenchil egam | S | L |
| <i>Sida schimperiana</i> Hochst. ex A. Rich. | Malvaceae | Tsfrer | S | L |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | S | L |

Appendix 16. Honeybee forage (N/P = Pollen/Nectar)

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|---|--------------|---------------|-------|-----------|
| <i>Abutilon longicuspe</i> Hochst. ex A. Rich. | Malvaceae | Sa'da buwak | S | N/P |
| <i>Acacia abyssinica</i> Hochst. ex Benth. | Fabaceae | Chea | T | N/P |
| <i>Acacia etbaica</i> Schweinf. | Fabaceae | Seraw | S | N/P |
| <i>Aloe camperi</i> Schweinf. | Aloaceae | Ere | S | N/P |
| <i>Becium grandiflorum</i> (Lam.) Pic.Serm. | Lamiaceae | Tebeb | S | N/P |
| <i>Cadia purpurea</i> (Picc.) Ait. | Fabaceae | Shilean | S | N/P |
| <i>Carissa spinarum</i> L. | Apocynaceae | Egam | S | N/P |
| <i>Dodonaea angustifolia</i> L. f. | Sapindaceae | Tahses | S | N/P |
| <i>Ekebergia capensis</i> Sparrm. | Meliaceae | Kot | S | N/P |
| <i>Erica arborea</i> L. | Ericaceae | Hasti | S | N/P |
| <i>Grewia bicolor</i> Juss. | Tiliaceae | Rewey | S | N/P |
| <i>Hypoestes forskoalii</i> (Vahl) R. Br. | Acanthaceae | Girbia | H | N/P |
| <i>Justicia schimperiana</i> (Hochst. ex Nees) T. Anders. | Acanthaceae | Shmeda | S | N/P |
| <i>Leucas abyssinica</i> (Benth.) Briq. | Lamiaceae | Shiwa qerni | S | N/P |
| <i>Maytenus arbutifolia</i> (A. Rich.) Wilcz | Celastraceae | Ats Ats | S | N/P |
| <i>Myrica salicifolia</i> A. Rich. | Myricaceae | Shinet | T | N/P |
| <i>Olea europaea</i> subsp. <i>cuspidata</i> (Wall. ex G. Don) Cif. | Oleaceae | Awlie | T | N/P |
| <i>Opuntia ficus-indica</i> (L.) Mill. | Cactaceae | Beles | S | N/P |
| <i>Psyrax schimperiana</i> (A. Rich.) Bridson | Rubiaceae | Tsehag | S | N/P |
| <i>Pterolobium stellatum</i> (Forssk.) Brenan | Fabaceae | Qonteftefe | S | N/P |

| Scientific name | Family name | Tigrigna name | Habit | Part used |
|--|--------------------|----------------------|--------------|------------------|
| <i>Rhus glutinosa</i> A. Rich. | Anacardiaceae | Tetaelo | S | N/P |
| <i>Rhus natalensis</i> Krauss | Anacardiaceae | Atami | S | N/P |
| <i>Rosa abyssinica</i> Lindley | Rosaceae | Qega | S | N/P |
| <i>Rumex nervosus</i> Vahl | Polygonaceae | Hehot | S | N/P |
| <i>Senecio hadiensis</i> Forssk. | Asteraceae | Sehum atali | S | N/P |
| <i>Sida schimperiana</i> Hochst. ex A. Rich. | Malvaceae | Tsfrer | S | N/P |
| <i>Teclea nobilis</i> Del. | Rutaceae | Selah | T | N/P |

Appendix 17. Interview items for ethnobotanical information collection.

Informant # _____

- 1 Name of Informant: _____
- 2 Occupation of Informant: _____
- 3 Educational status: _____
- 4 Marital status: _____
- 5 Gender of Informant: _____
- 6 Village (Forest Village): _____
- 7 District : _____

Semi structured interview

General questions:

1. What is the local name of the forest? What does the name refer to?
2. What are the traditional sayings, songs, poems and beliefs associated with the forest?
3. Are you aware of the population growth in the area? If yes, what do you think is the consequence of the population growth?
4. What type of environmental change have you observed in your localities for the past 10 years?
5. Do you own livestock? If yes, when and where do you graze them?
6. Where do you get the main source of fuel for your household?
7. Have you experienced any change in fuel availability during the last 10 years? If yes, what do you think is the reason?

Key informants

I. Cognitive domain on Biodiversity

1. How do you compare a natural and plantation forest and which one do you think is more important to your household?
2. Do you know the indigenous tree/shrub species available in your community? What are they?
3. What do you think are the most destructive activities to *Wejig-Mahgo-Waren* massif forest?
4. Which plant species have been threatened in the forest? What do you think is the reason?

II. Forest product utilization

1. What type of forest products do you use in your household?

2. Can you please tell us if your household is involved in trading some of the forest products? If yes, what type of products?
3. Can you please mention plants used for medicine?
7. What are the main human ailments at your locality?
8. What are the main livestock ailments at your locality?
9. How do you diagnose each ailment?
10. What are the major symptoms of the ailment?
11. How do you control ailments?
12. How do you treat human ailments?
13. How do you treat livestock ailments?
14. Which plant(s) do you use for treating that particular ailment?
15. Local name (vernacular) of the reported plant
16. Morphological description of the plant
17. Plant part/parts of the medicinal plant used
18. Used alone, mixed with water or other materials
19. How plant parts are used
20. Detailed preparation of the remedy
21. Amount of the remedy used (dose)
22. Does the dose differ among men, women, pregnant and children?
23. Any noticeable adverse effect(s)
24. Any antidotes for adverse/side effect(s)
25. Any restriction or taboo in collecting medicinal plants
26. Are there any threats to the medicinal plants?
27. Who are the source of traditional medicine for your practice?
28. How is the knowledge on traditional medicine passed to a family member/younger generation?
29. How does modernization interfere with traditional medicinal knowledge?
30. Can you please mention plants used for food, fuelwood, farm implements, building materials, livestock and bee forage.

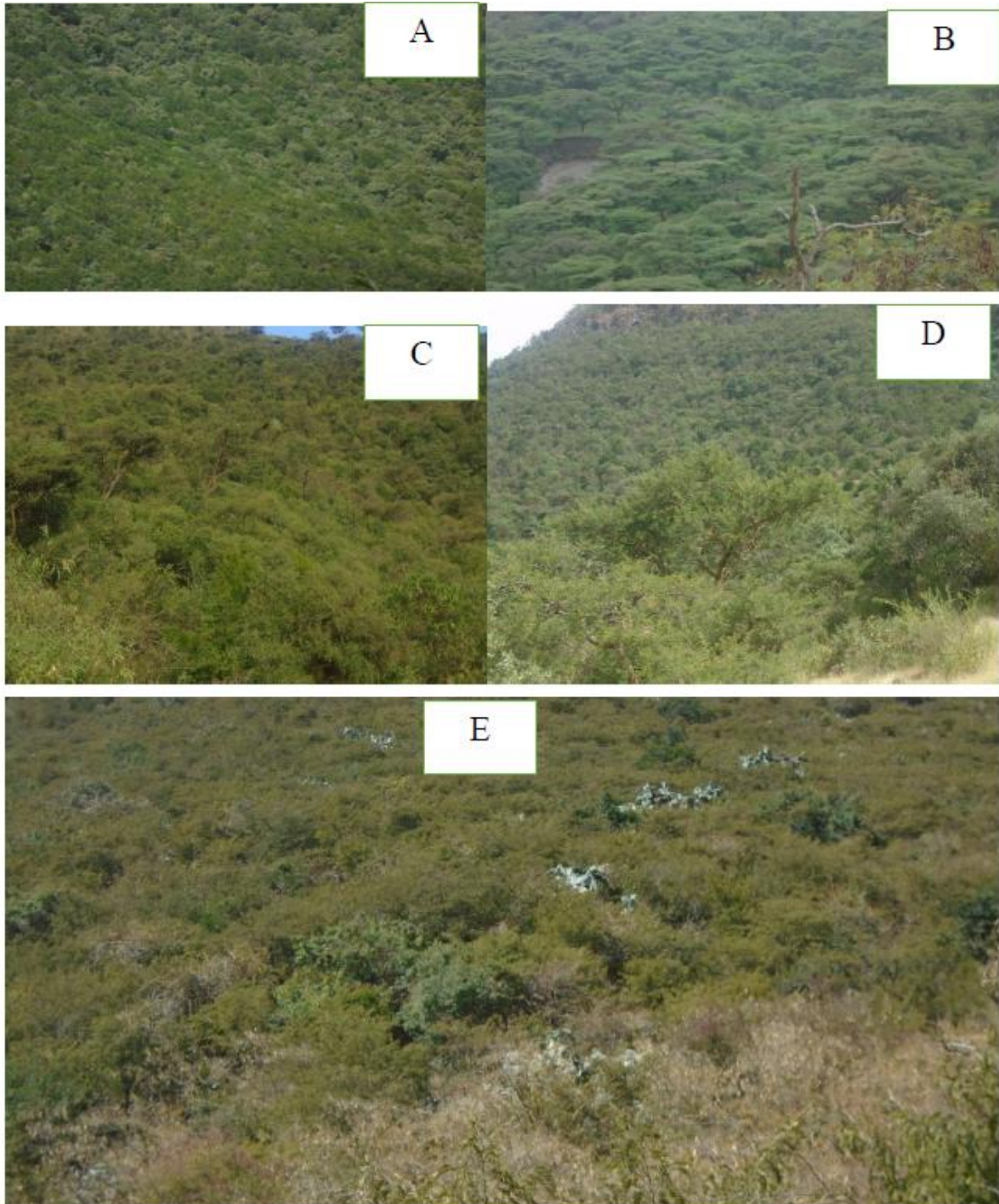
Focus group discussion

I. Cognitive domain on Forest management Issues

1. Before the demarcation of the *Wejig-Mahgo-Waren* massif forest as a biodiversity hotspot area, how did people of the local community traditionally manage the forest resources?
2. Were there any community laws preventing people from entering the forest for:
 - 2.1 Beekeeping
 - 2.2 Grazing
 - 2.3 Fuel wood collecting
 - 2.4 Building material collection
4. How do you compare traditional forest management with the current rules used in managing the forest?
4. Are you satisfied with current rules in managing the forest?
5. Have you ever requested permission to harvest forest products from the forest?
6. Have you ever been involved in making suggestions or decisions towards forest management?
7. How best do you think you participate in forest management activities?
8. Which local institution can work with government in managing the forest for the benefit of the people?
9. Would you please give us any additional information and suggestion?

I thank you very much for your cooperation!

Appendix 18. Plants communities found in *Wejig-Mahgo-Waren* massif forest.



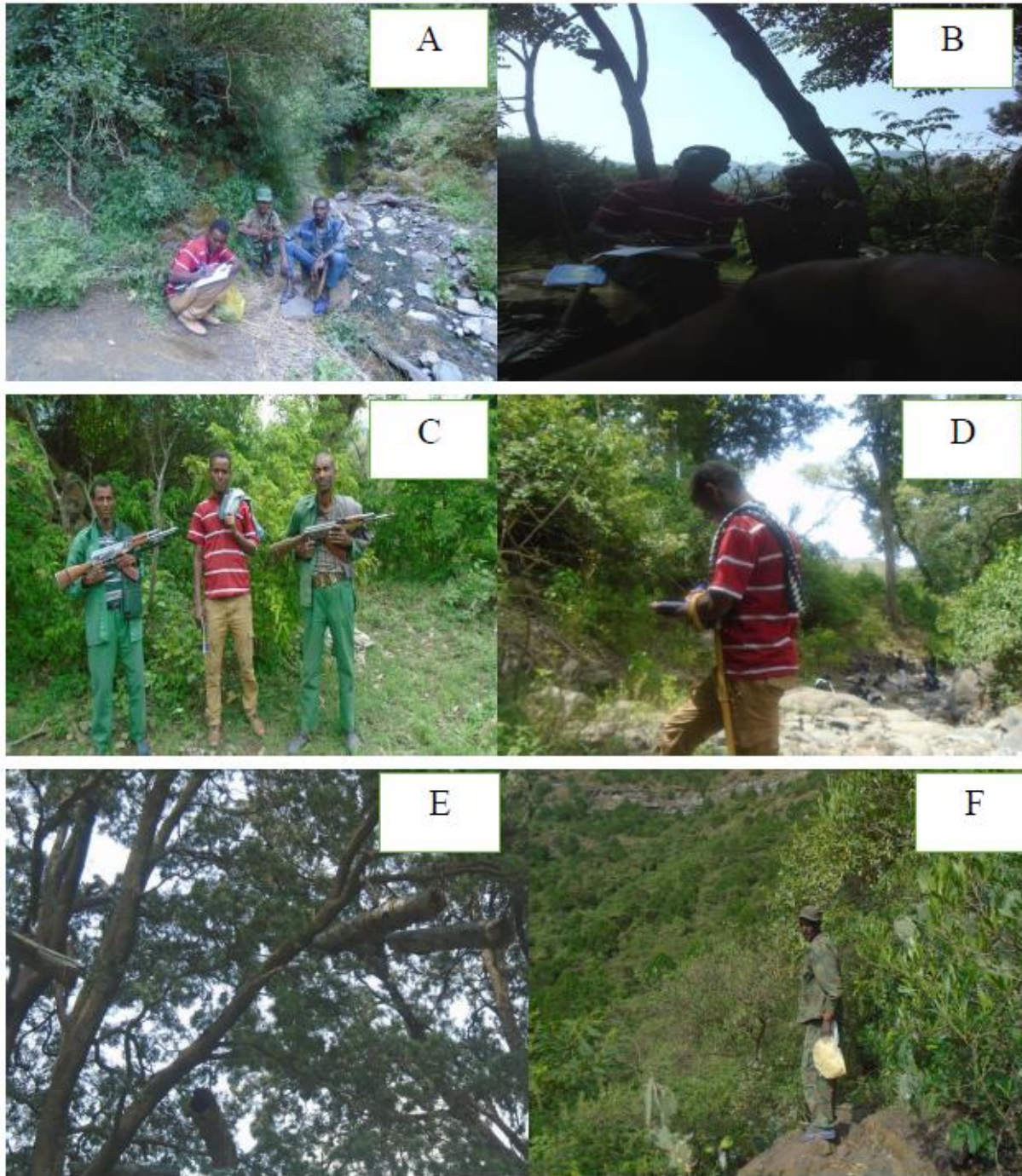
Key: Plant communities found at the study area, (A) *Olea europaea* subsp. *cuspidata* – *Juniperus procera* (B) *Acacia etbaica* – *Acacia tortilis* (C) *Dodonaea angustifolia* – *Acacia abyssinica* (D) *Erica arborea* – *Myrsine africana* (E) *Cadia purpurea* – *Carissa spinarum*.

Appendix 19. Some disturbances factors photos



Key: (A) Fuelwood collection, (B) *Olea europaea* subsp. *cuspidata* sold in the market, (C) Illegal cutting of *Olea europaea* subsp. *cuspidata*, (D) Grazing without permission inside the forest, (E- F) Deforestations for agricultural expansion

Appendix 20. Data recording in the field



Key: (A) Interview with guards inside the forest, (B) Interview with key informant, (C) Guards in Waren forest, (D) Data recording along the riverine forest, (E) *Podocarpus falcatus* with honeybee hive, (F) Guard in Wejig forest.

Appendix 21. Soil seed bank germination from the green house



Key: (A) First sieved soil sample placed on tray, (B) Emerging seedlings, (C) Mature seedlings ready for identification, (D) Visual identification using reproductive structures, (E) Stirring for further germination, (F) Pressed specimens brought to the National Herbarium.

Declaration

I, the undersigned declare that this Dissertation is my original work and it has not been presented to other universities, colleges or institutes for a degree or other purpose. All sources of the materials used have been duly acknowledged.

Name: _____ Signature: _____ Date: _____